

Review Article**Environmentally Favourable and Unfavorable Bacteria****Abstract**

Most of bacteria can be distinguished into three groups: saprophytic; symbiotic and parasitic. Microbial communities have a vast importance to the ecosystem and can be used by humans for their health or industrial applications. Saprophytic bacteria are the major decomposers of organic matter that can be applied in treatment of metalliferous mine or radioactive environmental wastes, biodiesel production, among others. Symbiotic bacteria live in a mutually beneficial association with other organisms providing essential nutrients to their host organisms. Some bacteria are able to cause diseases, the parasitic bacteria. Antimicrobial peptides and polypeptides such as lectins are promising candidates for being used as new antibiotics. Lectins are able to interact with carbohydrates in bacterial cellular walls and promote antibacterial activity. The aim of this chapter was to describe the importance of bacteria to environments, their use as biological control agents and the application of lectins to control pathogenic bacteria.

Keywords: environmental; bacteria; saprophytic; symbiotic; parasitic; biocontrol; lectins

28 1. INTRODUCTION

29

30 Bacteria are single-celled microorganisms, being classified as prokaryotes.
31 There are over 3.6 billion years bacteria are present on Earth in almost all possible
32 locations of life occurrence. This long co-evolution allowed bacteria to develop several
33 beneficial relationships to the environment and, therefore, beneficial to
34 themselves, since they form part of this system. The number of bacterial species that
35 have been described is low (~7,000) in relation to the millions of bacteria that have been
36 predicted to reside on Earth; the lack of knowledge about the ecology of each of the
37 known species is greater [1].

38 Microbial communities have a vast importance to the ecosystem and to
39 agriculture, being important components of the forest ecosystem since they facilitate
40 organic matter decomposition and nutrient cycling in the soil [2]. Free-living bacteria of
41 beneficial importance to agriculture and abound in the rhizosphere, the region around
42 the root, have more than one mechanism of accomplishing increased plant growth, such
43 as the production of enzymes, bioactive factors, antibiotics, metabolites as well as
44 growth promoters [3].

45 Bacteria can be classified, in terms of their morphology, as bacilli (rods), coccus
46 (spherical), spiral and many others (Figure 1). The bacillus is rod-shaped and can be
47 found as isolated bacilli, diplobacilli or streptobacilli. Coccus is circular and can be
48 isolated as dipococcus, tetracoccus, sarcina micrococcus, streptococci, and
49 staphylococci. Other bacterial shapes of lowest occurrence can occur as spirillum
50 (*Treponema pallidum*), as vibrio (*Vibrio cholerae*), transitional forms such as
51 coccobacillus and involution forms, a survival mechanism to adverse environmental
52 conditions such as spores [4].

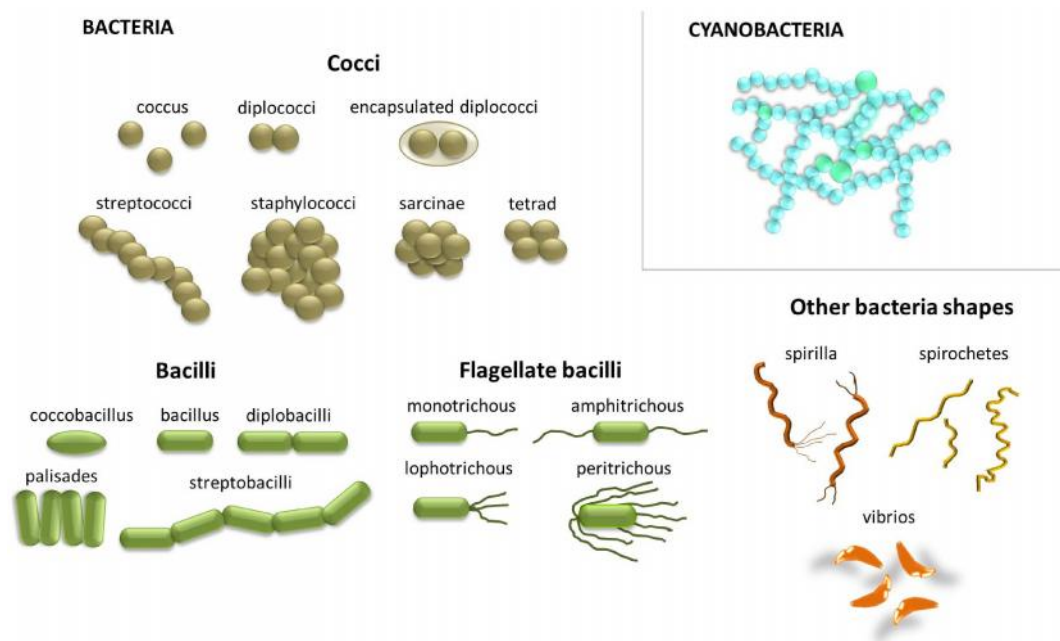


Figure 1: Types and shapes of bacteria.

Most bacteria are unable to manufacture their own organic food and hence dependent on external source (heterotrophic). These bacteria can be distinguished into three groups: 1) saprophytic; 2) symbiotic and 3) parasitic. Many bacteria that are associated with plants are actually saprophytic and do not harm the plant itself. However, a small number, around 100 species, is able to cause diseases, promoting losses in agriculture [5].

Microorganisms may be beneficial to humans microbiotes, complex infective bacteria that inhabit sites in and on the human body such as gut, skin, and oral cavity. Special situations as in patients whose normal innate defenses fail to function properly can lead to an imbalance of an individual species that are pathogens in the classical sense such as *Enterobacter sp.*, *Escherichia coli* and *Pseudomonas aeruginosa* [6].

The control of bacteria growth is a way to avoid ecosystem imbalance and disease caused by some of these microorganisms. However, massive use of antibiotics

69 for this purpose has led to bacterial resistance, generated by selection processes
70 including increase in the frequency of resistance bacterial genes [7]. As an alternative,
71 antimicrobial polypeptides such as lectins have been isolated and characterized from
72 tissues and organisms from every kingdom and phylum [8]. The complete
73 understanding of mechanisms of action from new alternatives to biological control may
74 provide models and strategies for developing novel antimicrobial agents, that may also
75 increase immunity, restore potency or amplify the mechanisms of conventional
76 antibiotics, and minimize antimicrobial resistance mechanisms among pathogens.

77

78 **2. Saprophytic bacteria**

79 The saprophytic bacteria are the major decomposers of organic matter (Figure
80 2), breaking down complex mixtures into simple soluble forms, freeing their atoms to
81 be re-used by other bioprocesses [9]. The ability of some acidophilic bacteria to
82 withstand raised concentrations of certain metals through biological oxi-reduction
83 reactions has been applied in a variety of industrial fields such as treatment of
84 metalliferous mine wastes, acid mine waters and sulphurous flue gases. The Matsuo
85 Mine in Japan applied this biological treatment system using *Thiobacillus ferrooxidans*
86 to treat 28m³/min of mine water at pH 2.5 oxidising more than 95 % of soluble ferrous
87 iron [10]. Microbial systems can detoxify the metal ions either by extracellular
88 biomineralization, biosorption, complexation or precipitation or intracellular
89 bioaccumulation. The cell wall reductive enzymes or soluble secreted enzymes can be
90 involved in the reductive process of metal ions by bacteria [11].

91 Some environmental factors such as availability of iron and sulphide and a
92 micro-aerobic environment are important for proliferation of the magnetotactic bacteria
93 *Magnetospirillum magneticum*. Magnetic minerals produced by these bacteria such as

greigite and biogenic magnetite form a post-depositional remanent magnetization that is indicative of rapid local environmental change. These biomarkers are used by archeologists to establish the chronology and environmental history of a place. Linford et al. [12] discovered bacterial magnetosomes at the village of Yarnton (Oxford, UK) suggesting a transformation of the previously dry river valley to an active flood plain.

Radiation-resistant micro-organisms have been used in the treatment of highly radioactive environmental wastes for their ability to transform, detoxify, or immobilize a variety of metallic and organic pollutants [13, 14] or used for decontamination of acid mine drainage waters through anaerobic degradation [15].

The techniques traditionally applied for the treatment of radioactive environmental residuals have been based on chemical methods of neutralization and precipitation. These quick and effective techniques have several disadvantages, such as the need for building additional plant treatments, the high cost of the chemical reagents used and the generation of an important volume of sludges which need to be relocated [15].

Some bacteria, when in contact with environmental stress, produce a signal transduction cascade in which certain promoters are induced, leading to expression of proteins that adjust to the ecological impact of altering the environment. Bioluminescent bacteria is being used as tools to detect some special compounds that are toxic and/or are of current interests as inorganic and organic pollutants from water, soil and air, as well as to monitor the level of toxicity of the influents from industries into urban wastewaters, effluents from plant treatments, and water. Recombinant bioluminescent bacterial strains are increasingly receiving attention as environmental biosensors due to their advantages, such as high sensitivity and selectivity, low costs, ease of use and short measurement times. Exposure of recombinant *Escherichia coli* strain, containing a

119 fusion of a promoter to the *Vibrio fischeri lux* genes (Ecolum-5), to a toxic or lethal
120 condition results in a decrease in bioluminescence [16]. The toxicity of benzene in air
121 was determined using the Ecolum-5 [17].

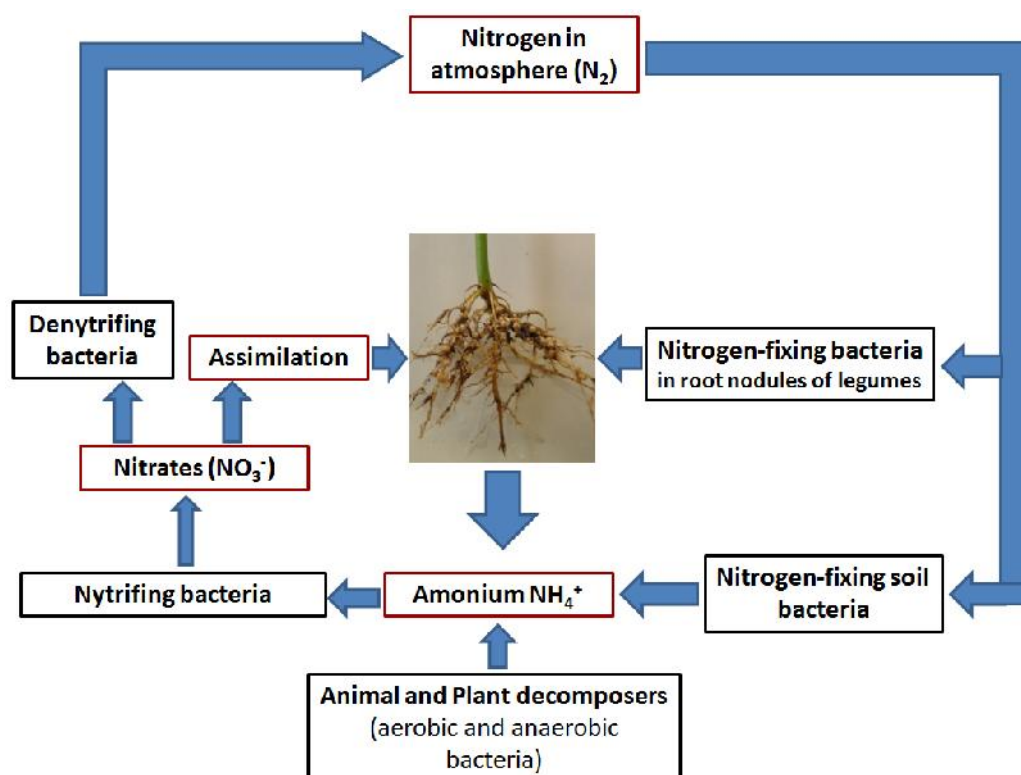
122 Biodiesel production is been stimulated as a result of search for renewable fuels.
123 The transesterification of vegetable oils or animal fats, with ethanol or methanol
124 generate glycerol as the main byproduct. With the increasing production of biodiesel,
125 glycerol is becoming of great environmental and economical concern due to its toxicity
126 to aquatic organisms [18]; fermentation processes of this byproduct can result in value-
127 added products, such as 1,3-propanediol (1,3-PD) and ethanol. The 1,3-PD has many
128 applications in polymers, cosmetics, foods, adhesives, lubricants, laminates, solvents,
129 antifreeze, and in medicine; ethanol could be used in the esterification of biodiesel,
130 contributing to develop cheaper and environmentally cleaner processes. Several
131 bacterial strains have been isolated and characterized for their ability to convert this raw
132 glycerol into 1,3-propanediol (1,3-PD) and ethanol [19, 20]. Rossi et al. [21] showed
133 that a *Klebsiella pneumoniae* was able to simultaneously produce up to 9.4 g/L of 1,3-
134 PD with yields of 0.41 mol product mol⁻¹ glycerol; and 6.1 g/L of ethanol with yields of
135 0.14 mol product mol⁻¹ glycerol.

136

137 **3. Symbiotic bacteria**

138 Symbiotic bacteria live in a mutually beneficial association with other
139 organisms. Such bacteria derive the essential nutrients from their host organisms and in
140 that process help the host through some of their biological activities. Plant growth-
141 promoting bacteria can positively provide the plant with compounds that is synthesized
142 by the bacterium or by facilitating the uptake of nutrients from the environment by the
143 plant. Nitrogen-fixing bacteria from *Rhizobium* genus can fix atmospheric nitrogen and

144 supply it to plants (Figure 2). Most of biological nitrogen fixation (80%) is carried out
 145 by diazotrophic bacteria, such as the *Rhizobium* genus, in symbiosis with legumes.
 146 Moreover, some bacteria which are free-living in soil (e.g., cyanobacteria,
 147 *Pseudomonas*, *Azospirillum*, and *Azotobacter*) may fix significant amounts of nitrogen
 148 [22].



149

150 Figure 2: Importance of saprophytic and symbiotic bacteria to the nitrogen cycle.

151

152 Other bacteria can synthesize siderophores, chelating agents which has more
 153 affinity to metals than plant siderophores and can solubilize and sequester iron from the
 154 soil providing it to plant cells. Antibiotics, which antagonize phytopathogenic fungi and
 155 pathogenic bacteria, and synthesize phytohormones, including auxins and cytokinins
 156 can enhance various stages of plant growth, and enzymes that can modulate plant
 157 growth and development [23, 24, 25].

158 Bacteria may be beneficent when they are associated with other organisms in the
159 removal of contaminants from the environment, a process called bioremediation [26].
160 Glick et al [23] showed that the synergistic use of the plant *Brassica campestris* and the
161 bacterium *Enterobacter cloacae* added to the roots lead to an increase in the number of
162 seeds that germinated and the amount of biomass that the plants were able to attain due
163 to reduction in the level of ethylene, an inhibitor of root elongation. Furthermore, the
164 bacterium synthesizes antibiotics, inhibiting the proliferation and invasion of
165 phytopathogens.

166 Additionally, bacteria can remove from the environment many potentially toxic
167 compounds like metals, organic compounds (such as petroleum hydrocarbons and
168 pesticides), inorganic (such as arsenic, sodium, nitrate, ammonia or phosphate) and
169 radioactive (such as uranium, cesium or strontium) compounds [23]. The bacterium
170 *Kluyvera ascorbata* protects *B. campestris* against high levels of nickel in the soil,
171 producing siderophores [27].

172 A group of bacteria named microbial flora (Figure 3) is able to beneficially
173 affects the host animal with contributions to nutrition, health and development by
174 secreting Vitamin K, B12 and folate; preventing colonization by pathogens by
175 competing for attachment sites or for essential nutrients in the oral cavity, intestine,
176 skin, and vaginal epithelium; excrete ammonium that can be used to synthesize proteins
177 and nucleic acids; and synthesize and excrete enzymes that act in the digestion of
178 carbohydrates [28]. The genera present in the intestinal tract (probiotic bacteria)
179 generally seem to be those from the environment or the diet. Probiotics in aquaculture
180 can prevent pathogens proliferating in the intestinal tract, on the superficial structures,
181 or in the water.

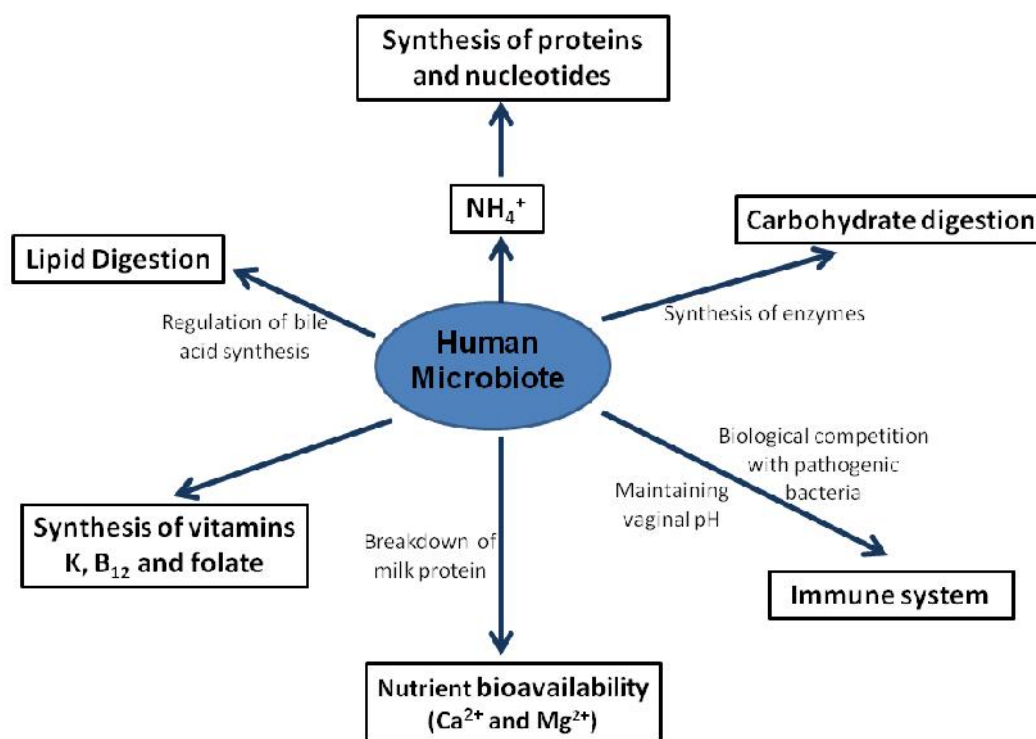


Figure 3: Benefits of human microbiote.

Probiotics can be used as biological control agents [29, 30]. This activity has been reported with lactic acid bacteria (*Lactobacillus* and *Pediococcus* genus) as biocontrol agents against the phytopathogenic and spoilage bacteria and fungi [31]. A bacterial strain (*Alteromonas haloplanktis*) isolated from the gonads of Chilean scallop displayed *in vitro* inhibitory activity against the pathogens *V. ordalii*, *V. parahaemolyticus*, *V. anguillarum*, *V. alginolyticus*, and *Aeromonas hydrophila* [32].

The exact modes of action of the probiotics were not completely elucidated, but it is suggested that microbial populations may release chemical substances such as antibiotics, lysozymes, proteases, hydrogen peroxide and organic acids that have a bactericidal or bacteriostatic effect on other microbial populations. Rouse et al. [33] showed that a lactic strains *Pediococcus pentosaceus* produced antifungal. Other possibilities of modes of action are by competition for nutrients as iron or adhesion sites

197 on gut or other tissue surfaces, by enhancement of the immune systems of animals
198 against infections by viruses, bacteria, fungi, and parasites or by improvement of water
199 quality [29].

200 Bacteria as *Pedobacter* sp. can act on many species of microalgae of red tide
201 plankton as, for example, *Microcystis aeruginosa* [34] function for controlling harmful
202 blooms and further studies will provide new insights into its role in water environment
203 with prospects to use this algicidal bacteria as microbial pesticides.

204 Bacteria in the aquatic environment are used as food by adults and larvae of
205 bivalve, providing nitrogen and carbon and recycling organic and mineral matter
206 released by bivalves. Marine bacteria excrete various substances, including amino acids,
207 carbohydrates, and vitamins of group B. Bacteria of bivalve microflora are important in
208 the digestive process, metabolism and metamorphosis [35]. Prieur et al. [36] isolated
209 cellulolytic bacteria from the digestive tract of the bivalve *Teredo navalis* that had the
210 ability to degrade mannose and galactose. Belkin et al. [37] showed that the associated
211 bacteria from gill tissues of a mussel from deep ocean *Bathymodiolus thermophilus*
212 were capable of CO₂ fixation indicating that the symbiotic bacteria have an autotrophic
213 metabolism. Bivalve bacteria that live in sulphide-enriched habitats are important in the
214 degradation of the organic matter through anaerobic metabolism [36].

215 Among the environmental factors that induce or influence metamorphosis of
216 many marine invertebrates, the occurrence of bacterial film and organic particles
217 trapped within the film could also be used as food by larvae ready to metamorphose;
218 alternatively, bacteria living in the biofilm could synthesize certain compounds such as
219 low and high molecular weight polysaccharides, low molecular weight peptides and
220 neurotransmitters, diffusible into the environment, which could induce metamorphosis.
221 Water-soluble chemical cues produced by the biofilm of two bacterial strains

222 *Macrocooccus* sp. and *Bacillus* sp. induced larval settlement of the green-lipped mussel,
223 *Perna canaliculus* [38].

224

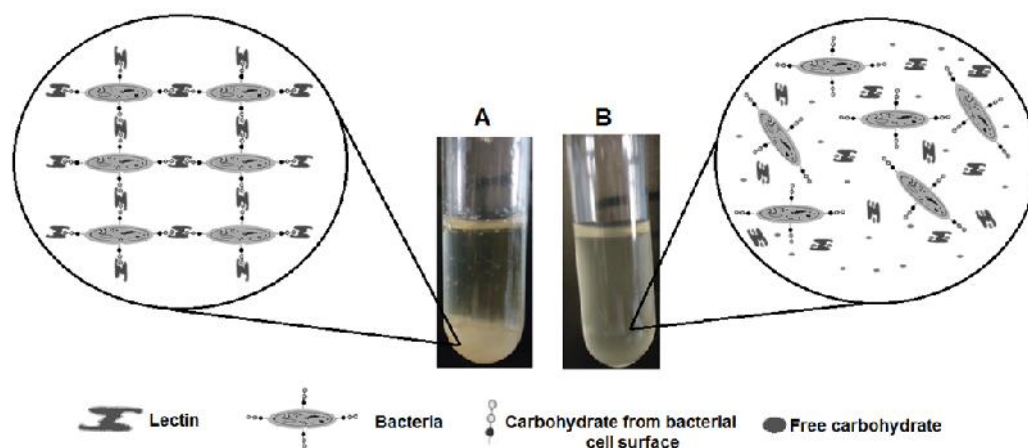
225 **4. Parasitic bacteria**

226

227 Parasitic bacteria occur in the body of animals and in plants obtaining their
228 organic food or releasing poisonous secretions called toxins. Many of these toxins are
229 specific to target-organism. Thus, the majority of bacterial pathogens are highly
230 specialized for a limited number of eukaryotic host organisms. Plant pathogenic bacteria
231 (Figure 4) are responsible for some of the most devastating losses of major agricultural
232 crops and vital fruit trees, causing millions of dollars in damage annually [39].
233 *Ralstonia solanacearum* is a soil borne bacterium, capable of inducing disease on more
234 than 250 plant species, invading its roots, colonizing the xylem vessels, and causing a
235 lethal wilting known as bacterial wilt disease [40]. Seeds of cashew, cocoa, coffee,
236 pumpkin, and tomato are protected from this bacteria because they produce oligo- and
237 poly-saccharides that block the pathogen lectins from *R. solanacearum*, inhibiting its
238 binding to xylem cell wall glycans [41].

239

240



241

242 Figure 4: (A) Agglutination of *Staphylococcus aureus* by incubation with the *Schinus*
 243 *terebinthifolius* leaf lectin (SteLL). (B) Inhibition of agglutination in the presence of the
 244 carbohydrate inhibitor of SteLL (N-acetyl-glucosamine). The circles show
 245 schematizations of the phenomena occurring in the assay tubes.

246

247 Bacterial canker is another disease caused by phytopathogen that has economic
 248 impact. Bacterial canker of grapevine caused by *Xanthomonas campestris* pv. *viticola*
 249 can be manifested in various parts of the plant. In leaves, the symptoms are small, dark
 250 and angular leaf spots that may coalesce and dry up, causing necrotic areas and leaf
 251 blight. Cankers were often observed on petioles, stems and rachis and were also
 252 observed in grapes [42].

253 Blackleg, a major bacterial disease of potato, is caused by bacteria
 254 *Pectobacterium carotovorum* subsp. *carotovorum*. This bacterium can cause rotting of
 255 potato tubers (soft rot) during storage. Control of potato blackleg is hampered by the
 256 absence of effective tools and strategies and by the dispersing ability of the bacterium,
 257 being spread via surface and rain water, by aerosols and also, by insects [43].

258 *Acidovorax citrulli* is the bacterial causal agent of bacterial fruit blotch, a
 259 devastating disease of melon (*Cucumis melo*) and other Cucurbitaceae, and its

destructive potential stems from the fact that, under favorable conditions, infection spreads rapidly throughout the field [44]. Symptoms of bacterial fruit blotch including water-soaking and coalescing reddish-brown lesions on cotyledons and reddish-brown lesions on leaves that developed along the venation [45].

Among pathogenic bacteria to humans, there is *Staphylococcus aureus*, which is a coccus (spherical) microorganism usually with irregular distribution in clusters like bunches of grapes that is responsible for many infections in humans such as endocarditis, acute hematogenous osteomyelitis, meningitis or pulmonary infection. *Escherichia coli*, which is in the form of rod-shaped (a bacillus), is part of the normal flora and accidentally cause diseases (urinary tract infection, diarrhea, meningitis and septicemia); *Pseudomonas aeruginosa*, a mobile aerobic bacillus widely distributed in nature and is found in small groups of normal intestinal flora and on human skin [4].

5. Antibacterial lectins

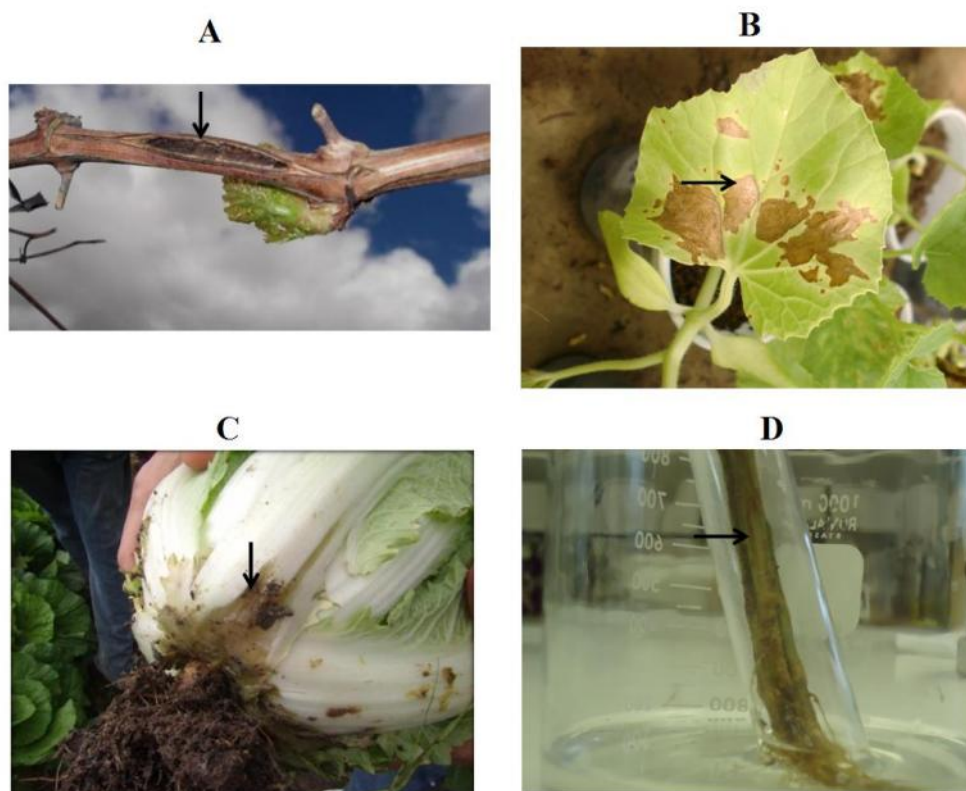
Antimicrobial peptides and polypeptides are promising candidates for using as new antibiotics. The carbohydrate recognizing proteins known as lectins are noteworthy since they are able to interact with carbohydrates in bacterial cellular walls [46-48].

Lectins can agglutinate cells and precipitate polysaccharides, glycoproteins or glycolipids, without structural modifications [49]. The presence of lectins in a sample can be evaluated in a microtiter plate through incubation with erythrocytes. The linkage between lectins and glycoconjugates from erythrocyte surface maintains the cells agglutinated and suspended in solution.

Gomes et al. [50] isolated an antimicrobial chitin-binding-lectin from leaves of the medicinal plant *Schinus terebinthifolius*. The authors reported that this lectin

285 showed antibacterial activity against *E. coli* (MIC, minimal inhibitory concentration, of
286 28.5 µg/ml; MBC, minimal bactericidal concentration, of 115 µg/ml), *K. pneumoniae*
287 (MIC of 3.59 µg/ml; MBC of 115 µg/ml), *Proteus mirabilis* (MIC of 3.59 µg/ml; MBC
288 of 14.37 µg/ml), *P. aeruginosa* (MIC of 1.79 µg/ml; MBC of 14.37 µg/ml), *Salmonella*
289 *enteritidis* (MIC of 0.45 µg/ml; MBC of 115 µg/ml), and *S. aureus* (MIC of 1.79 µg/ml;
290 MBC of 7.18 µg/ml).

291 Lectins can also promote agglutination of bacterial cells. Figure 5 shows the
292 agglutination of *Staphylococcus aureus* promoted by *S. terebinthifolius* leaf lectin
293 (SteLL) in assay tubes. The agglutination occurs through linkage between the
294 carbohydrate binding sites of lectin and glycoconjugates from bacterial surface as
295 schematized in Figure 5A. The bacterial agglutination was inhibited in the presence of
296 the carbohydrate inhibitor of SteLL (N-acetyl-glucosamine) as shown in Figure 5B. The
297 inhibition of bacteria agglutination by lectin after incubation with free carbohydrates or
298 glycoconjugates ensures that lectin binding to bacteria involves the carbohydrate-
299 binding sites.



300
 301 Figure 5. Diseases caused by phytopathogenic bacteria. Bacterial canker of grapevine
 302 caused by *Xanthomonas campestris* pv. *viticola* (A); blotch of melon caused by
 303 *Acidovorax citrulli*, showing reddish-brown lesions on leaves (B); rooting of *Brassica*
 304 *pekinensis* by *Pectobacterium carotovorum* subsp. *carotovorum* (C); tomato wilt
 305 disease caused by *Ralstonia solanacearum*, showing colonization of xylem vessels (D).

306
 307 Oliveira et al. [51] isolated an antibacterial lectin from *Eugenia uniflora* seeds
 308 (EuniSL), which inhibited the growth of *Staphylococcus aureus*, *Pseudomonas*
 309 *aeruginosa* and *Klebsiella* sp. with MIC of 1.5 µg/ml. Additionally, EuniSL also
 310 inhibited the growth of *Bacillus subtilis*, *Streptococcus* sp. and *Escherichia coli*,
 311 although less efficiently (MIC of 16.5 µg/ml). The authors also showed that EuniSL was
 312 able to agglutinate *S. aureus*, *Streptococcus* sp., *Klebsiella* sp. and *P. aeruginosa*.

313 MuHL, a chitin-binding-lectin isolated from *Myracrodruon urundeuva*
 314 heartwood, was able to inhibit the growth and agglutinate the Gram-positive bacteria *S.*
 315 *aureus* (MIC of 0.58 µg/ml; MAC, minimal agglutinating concentration, of 2.34 µg/ml),
 316 *Enterococcus (Streptococcus) faecalis* (MIC of 2.34 µg/ml; MAC of 4.68), *B. subtilis*
 317 (MIC of 2.34 µg/ml; MAC of 4.68 µg/ml), and *Corynebacterium callunae* (MIC of 1.17
 318 µg/ml; MAC of 4.68 µg/ml), as well as the Gram-negative bacteria *E. coli* (MIC of 9.37
 319 µg/ml; MAC of 9.37 µg/ml), *Klebsiella pneumoniae* (MIC of 9.37 µg/ml; MAC of 9.37
 320 µg/ml) and *P. aeruginosa* (MIC of 4.68 µg/ml; MAC of 9.37 µg/ml) [52]. *M. urundeuva*
 321 heartwood is very resistant to the deteriorative biological agents and the authors pointed
 322 out that antibacterial activity of MuHL may be involved in this resistance.

323 A lectin isolated from leaf of *Phthirusa pyrifolia* also showed antibacterial
 324 potential, being active against Gram-positive (*Staphylococcus epidermidis*,
 325 *Enterococcus (Streptococcus) faecalis* and *Bacillus subtilis*) and Gram-negative
 326 (*Klebsiella pneumoniae*) bacteria, and the MIC values ranged from 250 µg/ml to >2000
 327 µg/ml [53]. WSMoL, a water-soluble lectin purified from seeds of *Moringa oleifera*,
 328 reduced the growth of *S. aureus* and *E. coli* and was also active against ambient lake
 329 water bacteria [54].

330 Antimicrobial lectins have also been isolated from animals. Nunes et al. [55]
 331 purified a lectin from *Bothrops leucurus* snake venom, which inhibited the growth of
 332 the Gram-positive bacteria *S. aureus*, *E. faecalis* and *B. subtilis* with MIC of 31.25,
 333 62.25 and 125 µg/ml, respectively. Table 1 lists antibacterial lectins and species against
 334 which they are active.

335

336

337

338 Table 1. Antimicrobial activity of lectins.

Lectin	Source	Antibacterial activity
Lectins from plants		
EuniSL	Seeds of <i>E. uniflora</i>	<i>S. aureus</i> , <i>P. aeruginosa</i> and <i>Klebsiella</i> sp
MuHL	Heartwood of <i>M. urundeuva</i>	<i>S. aureus</i> , <i>E. faecalis</i> , <i>B. subtilis</i> , <i>C. callunae</i> , <i>E. coli</i> , <i>K. pneumoniae</i> and <i>P. aeruginosa</i>
PpyLL	Leaves of <i>P. pyrifolia</i>	<i>S. epidermidis</i> , <i>E. faecalis</i> , <i>B. subtilis</i> and <i>K. pneumonia</i>
WSMoL	Seeds of <i>M. oleifera</i>	<i>S. aureus</i> , <i>E. coli</i> and bacteria from ambient lake water
SteLL	Leaves of <i>S. terebinthifolius</i>	<i>E. coli</i> , <i>K. pneumonia</i> , <i>P. mirabilis</i> , <i>P. aeruginosa</i> , <i>S. enteritidis</i> and <i>S. aureus</i>
Lectin from animal		
BIL	<i>B. leucurus</i> snake venom	<i>S. aureus</i> , <i>E. faecalis</i> and <i>B. subtilis</i>

339 References: [50-52, 54, 55].

340

341 Moura et al. [56] overviewed processes involved in biofilm formation and in
 342 biocorrosion of pipes for oil transportation, which occurs due to fixation of bacteria that
 343 release metabolites and form biofilms inducing or accelerating corrosion. The authors
 344 highlighted five groups of bacteria (EPS-producing bacteria, acid-producing bacteria,
 345 sulfur-oxidizing bacteria, iron-precipitating bacteria and sulfate-reducing bacteria) as
 346 promoters of biocorrosion. In addition, authors pointed out the use of biocides,
 347 protective coatings (antifouling) and corrosion inhibitors as the main methods applied
 348 by industry to prevent corrosive bacteria spreading. It was then suggested that plant
 349 compounds, including lectins, are interesting candidates to be used as controlling
 350 biocorrosion agents.

351

352 **6. Bacteria as biological control agents**

353

354 Pesticides are used to control organisms considered harmful [57]. However, the
355 main problem of the use of chemical pesticides is the development of resistance,
356 resulting in reduced efficiency of the product and increased environmental risk.
357 Pesticides are one of the causes of water pollution, and some pesticides are persistent
358 organic pollutants which contribute to soil contamination [58].

359 Bacteria have been studied as safer and more eco-friendly alternatives for the
360 control of postharvest decays caused by fungi. The bacteria used for this purpose
361 usually act as antagonistic microorganisms probably through competition for nutrients
362 and space as well as by production of antibiotics, direct parasitism, and possibly
363 induced resistance in the harvested commodity. *B. subtilis* has been used as antagonist
364 for phytopathogenic fungi that attack fruits [59].

365 Investigations carried out by Yoshiyama and Kimura [60] showed that seven
366 bacteria strains belonging to *Bacillus* genus, isolated from digestive tract of the
367 Japanese honeybee (*Apis cerana japonica*), inhibited the development of the Gram-
368 positive bacterium *Paenibacillus larvae*, the causal agent of American foulbrood. This
369 disease is contagious and affects the larval and pupal stages of honeybees. The authors
370 suggested that these *Bacillus* strains can be used for control of this disease by acting as
371 antagonists of *P. larvae*.

372 An alternative is the use of *Bacillus thuringiensis*, a bacterium that produce
373 toxins with hemolytic and cytolytic activity. This versatile pathogen is capable of
374 infecting protozoa, nematodes, flatworms, mites and insects [61]. *B. thuringiensis* is
375 characterized by the production of crystals composed of protein known as
376 deltaendotoxins that are toxic to insect pests [62].

377 *B. thuringiensis* (Bt), before 1976, was used exclusively to the control of insect
 378 pests in agriculture. The discovery of a pathogenic strain against Diptera, called Bt
 379 israelensis (Bti) initiated the use of this bacterium in the control of disease vector.
 380 Insects such as *Aedes aegypti* (Culicidae), vector of dengue and yellow fever, and
 381 insects of the Simuliidae family, transmitters of filariasis, are included in the Diptera
 382 order. The use of bacteria for biological control of insect larvae from Culicidae and
 383 Simuliidae family has highlighted by having more kinds of formulation (granules,
 384 powder or liquid), genetic stability, not toxic to humans, besides being more
 385 advantageous considering social and environmental costs of using non-selective
 386 insecticides in aquatic ecosystems [63].

387 A strategy used in Brazil as part of the National Program of Dengue Control is
 388 the biological control with *Bacillus thuringiensis* serovar *israelensis* (Bti). The
 389 endotoxin Cry1AC, produced during Bti sporulation, is digested by enzymes of larvae
 390 midgut releasing larvicidal toxins; tablet containing spore and crystals (15%, w/w) of *B.*
 391 *thuringiensis* was able to cause 100 % mortality of larvae and was suggested for use in
 392 programs to control dengue vector [64]. Cry1AC has an *N*-acetylgalactosamine-specific
 393 lectin domain that binds glycoconjugates at insect midgut [65, 66]. Another example of
 394 larvicidal protein produced by bacterial strains is that from *Bacillus sphaericus*, which
 395 was lethal to the 3rd instar larvae of *Culex pipiens*, the vector of the West Nile fever and
 396 the Rift Valley fever [67].

397

398 7. Conclusion

399 The purpose of this review was to approach the biological importance of bacteria to
 400 distinct environments, how they are used as biological control agents and the
 401 importance of lectins to control pathogenic bacteria that affect animals (including

humans) and plants. Moreover, this review showed biotechnological applications of bacteria in many areas of human interest. This review was motivated by the lack of knowledge about the ecology of bacteria and by the success of plant lectins as antimicrobial agents.

Competing Interest

Authors have declared that no competing interests exist.

REFERENCES

[1] Philippot L, Andersson SG, Battin TJ, Prosser JI, Schimel JP, Whitman WB, et al. The ecological coherence of high bacterial taxonomic ranks. *Nat Rev Microbiol.* 2010; 8(7): 523-529.

[2] Hynes HM, Germida JJ. Impact of clear cutting on soil microbial communities and bioavailable nutrients in the LFH and Ae horizons of Boreal Plain forest soils. *Forest Ecol Manag.* 2013; 306: 88-95.

[3] Babalola OO. Beneficial bacteria of agricultural importance. *Biotechnol lett.* 2010; 32(11): 1559-1570.

[4] Trabulsi LR, Alterthum F. *Microbiologia*, 4th ed. São Paulo: Atheneu; 2004. Portuguese.

426 [5] Siddiqui ZA, Nesha R, Singh N, Alam S. Interactions of Plant-Parasitic Nematodes
427 and Plant-Pathogenic Bacteria. In: Maheshwari DK editor. Bacteria in Agrobiolgy:
428 Plant Probiotics. Springer Berlin Heidelberg. 251-267; 2012.

429

430 [6] Rogers GB, Hoffman LR, Carroll MP, Bruce KD. Interpreting infective microbiota:
431 the importance of an ecological perspective. Trends Microbiol. 2013; 21(6): 271-276.

432

433 [7] El-Baky RMA, Ahmed HR, Gad GFM. Prevalence and conjugal transfer of
434 vancomycin resistance among clinical isolates of *Staphylococcus aureus*. Adv Res.
435 2014; 2(1): 12-23

436

437 [8] Yeaman, M R and Yount, N Y. Mechanisms of antimicrobial peptide action and
438 resistance. Pharmacol Rev. 2003; 55(1): 27-55.

439

440 [9] Deyn GB, Quirk H, Bardgett RD. Plant species richness, identity and productivity
441 differentially influence key groups of microbes in grassland soils of contrasting fertility.
442 Biology lett. 2011; 7(1): 75-78.

443

444 [10] Smith FE. The Treatment of Metalliferous Industrial Effluents in Japan. In
445 Proceedings of Sudbury, 95: 915-924; 1995.

446

447 [11] He S, Guo Z, Zhang Y, Zhang S, Wang J, Gu N. Biosynthesis of gold nanoparticles
448 using the bacteria *Rhodopseudomonas capsulata*. Mater Lett. 2007; 61(18): 3984-3987.

449

450 [12] Linford N, Linford P, Platzman E. Dating environmental change using magnetic
451 bacteria in archaeological soils from the upper Thames Valley, UK. *J Archaeol Sci.*
452 2005; 32(7): 1037-1043.

453

454 [13] Daly MJ. Engineering radiation-resistant bacteria for environmental biotechnology.
455 *Curr Opin Biotech.* 2000; 11(3): 280-285.

456

457 [14] Fredrickson JK, Gorby YA. Environmental processes mediated by iron-reducing
458 bacteria. *Curr Opin Biotech.* 1996; 7(3): 287-294.

459

460 [15] Garcia C, Moreno DA, Ballester A, Blazquez ML, Gonzalez F. Bioremediation of
461 an industrial acid mine water by metal-tolerant sulphate-reducing bacteria. *Miner Eng.*
462 2001; 14(9): 997-1008.

463

464 [16] Lee JH, Mitchell RJ, Kim BC, Cullen DC, Gu MB. A cell array biosensor for
465 environmental toxicity analysis. *Biosens Bioelectron.* 2005; 21(3): 500-507.

466

467 [17] Girotti S, Ferri EN, Fumo MG, Maiolini, E. Monitoring of environmental
468 pollutants by bioluminescent bacteria. *Anal Chim Acta.* 2008; 608(1): 2-29.

469

470 [18] Ding Z, Hao A, Wang Z. Water-in-gasoline microemulsions stabilized by
471 polyglycerol esters. *Fuel.* 2007; 86(4): 597-602.

472

473 [19] Chen X, Xiu Z, Wang J, Zhang D, Xu P. Stoichiometric analysis and experimental
474 investigation of glycerol bioconversion to 1, 3-propanediol by *Klebsiella pneumoniae*
475 under microaerobic conditions. *Enzyme Microb Tech.* 2003; 33(4): 386-394.

476

477 [20] Metsoviti M, Paramithiotis S, Drosinos EH, Galiotou-Panayotou M, Nychas GJE,
478 Zeng AP et al. Screening of bacterial strains capable of converting biodiesel-derived
479 raw glycerol into 1, 3-propanediol, 2, 3-butanediol and ethanol. *Eng Life Sci.* 2012;
480 12(1): 57-68.

481

482 [21] Rossi DM, da Costa JB, de Souza EA, Peralba MDCR, Ayub MAZ. Bioconversion
483 of residual glycerol from biodiesel synthesis into 1, 3-propanediol and ethanol by
484 isolated bacteria from environmental consortia. *Renew Energ.* 2012; 39(1): 223-227.

485

486 [22] Orr CH, James A, Leifert C, Cooper JM, Cummings SP. Diversity and activity of
487 free-living nitrogen-fixing bacteria and total bacteria in organic and conventionally
488 managed soils. *Appl Environ Microb.* 2011; 77(3): 911-919.

489

490 [23] Glick BR. Phytoremediation: synergistic use of plants and bacteria to clean up the
491 environment. *Biotechnol Adv.* 2003; 21(5): 383-393.

492

493 [24] Sokolova MG, Akimova GP, Vaishlya OB. Effect of phytohormones synthesized
494 by rhizosphere bacteria on plants. *Appl Biochem Microb.* 2011; 47(3): 274-278.

495

496 [25] Schalk IJ, Hannauer M, Braud A. New roles for bacterial siderophores in metal
497 transport and tolerance. *Environ Microbiol.* 2011; 13(11): 2844-2854.

498

499 [26] Guo H, Luo S, Chen L, Xiao X, Xi Q, Wei W et al. Bioremediation of heavy
500 metals by growing hyperaccumulaor endophytic bacterium *Bacillus* sp. L14.
501 Bioresource Technol. 2010; 101(22): 8599-8605.

502

503 [27] Ma W, Zalec K, Glick BR. Effects of the bioluminescence-labeling of the soil
504 bacterium *Kluyvera ascorbata* SUD165/26. FEMS Microbiol Ecol. 2001; 35: 137–44.

505

506 [28] Cho I, Blaser MJ. The human microbiome: at the interface of health and disease.
507 Nat Rev Genet. 2012; 13(4): 260-270.

508

509 [29] Verschuere L, Rombaut G, Sorgeloos P, Verstraete W. Probiotic bacteria as
510 biological control agents in aquaculture. Microbiol Mol Biol R. 2000; 64(4): 655-671.

511

512 [30] Dalié DKD, Deschamps AM, Richard-Forget F. Lactic acid bacteria–Potential for
513 control of mould growth and mycotoxins: A review. Food Control. 2010; 21(4): 370-
514 380.

515

516 [31] Trias R, Bañeras L, Montesinos E, Badosa, E. Lactic acid bacteria from fresh fruit
517 and vegetables as biocontrol agents of phytopathogenic bacteria and fungi. Int
518 Microbiol. 2010; 11(4): 231-236.

519

520 [32] Riquelme, C Araya R, Vergara N, Rojas A, Guaita M, Candia M. Potential
521 probiotic strains in the culture of the Chilean scallop *Argopecten purpuratus* (Lamarck,
522 1819). Aquaculture. 1997; 154(1): 17-26.

523

524 [33] Rouse S, Harnett D, Vaughan A, van Sinderen D. Lactic acid bacteria with
525 potential to eliminate fungal spoilage in foods. J Appl Microbiol. 2008; 104:915–923.

526

527 [34] Yang L, Maeda H, Yoshikawa T, Zhou GQ. Algicidal effect of bacterial isolates of
528 *Pedobacter* sp. against cyanobacterium *Microcystis aeruginosa*. Water Sci Eng. 2012;
529 5(4): 375-382.

530

531 [35] Betcher MA, Fung JM, Han AW, O'Connor R, Seronay R, Concepcion GP et al.
532 Microbial distribution and abundance in the digestive system of five shipworm species
533 (Bivalvia: Teredinidae). PLoS One. 2012; 7 (9): e45309

534

535 [36] Prieur D, Mevel G, Nicolas JL, Plusquellec A, Vigneulle M. Interactions between
536 bivalve molluscs and bacteria in the marine environment. Oceanogr. Mar. Biol. Annu.
537 Rev. 1990; 28: 277-352.

538

539 [37] Belkin S, Wirsén CO, Jannasch HW. A new sulfur-reducing, extremely
540 thermophilic eubacterium from a submarine thermal vent. Appl Environ Microb. 1986;
541 51(6): 1180-1185.

542

543 [38] Alfaro AC, Young T, Ganesan AM. Regulatory effects of mussel (*Aulacomya*
544 *maoriana* Iredale 1915) larval settlement by neuroactive compounds, amino acids and
545 bacterial biofilms. Aquaculture. 2011; 322: 158-168.

546

547 [39] Nadarasah G, Stavrinides J. Insects as alternative hosts for phytopathogenic
548 bacteria. FEMS Microbiol Rev. 2011; 35(3): 555-575.

549

550 [40] N'Guessan CA, Brisse S, Roux-Nio ACL, Poussier S, Koné D, Wicker, E.
551 Development of variable number of tandem repeats typing schemes for *Ralstonia*
552 *solanacearum*, the agent of bacterial wilt, banana Moko disease and potato brown rot. J
553 Microbiol Meth. 2013; 92: 366-374.

554

555 [41] Rachmaninov O, Zinger-Yosovich KD, Gilboa-Garber N. Preventing *Ralstonia*
556 *solanacearum* adhesion with glycans from cashew, cocoa, coffee, pumpkin, and tomato
557 seed extract. Can J Microbiol. 2012; 58(7): 856-862.

558

559 [42] Nascimento AR, Michereff SJ, Mariano RDL, Gomes, AM. Elaboração e validação
560 de escala diagramática para cancro bacteriano da videira. Sum Phytopathol. 2005;
561 31(1): 59-64. Portuguese

562

563 [43] Czajkowski R, Grabe GJ, Van der Wolf JM. Distribution of *Dickeya* spp. and
564 *Pectobacterium carotovorum* subsp. *carotovorum* in naturally infected seed potatoes.
565 Eur J Plant Pathol. 2009; 125(2): 263-275.

566

567 [44] Walcott RR, Gitaitis, RD. Detection of *Acidovorax avenae* subsp. *citrulli* in
568 watermelon seed using immunomagnetic separation and the polymerase chain reaction.
569 Plant Dis, 2000; 84(4): 470-474.

570

571 [45] Walcott RR, Fessehaie A, Castro AC. Differences in pathogenicity between two
572 genetically distinct groups of *Acidovorax avenae* subsp. *citrulli* on cucurbit hosts. J
573 Phytopathol. 2004; 152(5): 277-285.

574

575 [46] Gaidamashvili M, Van Staden J. Interaction of lectin-like proteins of South
576 African medicinal plants with *Staphylococcus aureus* and *Bacillus subtilis*. J
577 Ethnopharmacol. 2002; 80(2): 131-135.

578

579 [47] Ratanapo S, Ngamjunyaporn W, Chulavatnatol M. Interaction of a mulberry leaf
580 lectin with a phytopathogenic bacterium, *P. syringae* pv *mori*. Plant Sci. 2001; 160(4):
581 739-744.

582

583 [48] Tasumi S, Yang WJ, Usami T, Tsutsui S, Ohira T, Kawazoe I et al. Characteristics
584 and primary structure of a galectin in the skin mucus of the Japanese eel *Anguilla*
585 *japonica*. Dev Comp Immunol. 2004; 28(4): 325-335.

586

587 [49] Paiva PMG, Pontual EV, Napoleão TH, Coelho LCBB. Effects of plant lectins and
588 trypsin inhibitors on development, morphology and biochemistry of insect larvae. In:
589 Pourali, H.; Raad, V.N (Eds.). Larvae: Morphology, Biology and Life Cycle. Nova
590 Science Publishers, Inc., New York, 37–55; 2012.

591

592 [50] Gomes FS, Procópio TF, Napoleão TH, Coelho LCBB, Paiva PMG. Antimicrobial
593 lectin from *Schinus terebinthifolius* leaf. J Appl Microbiol. 2013; 114(3): 672-679.

594

595 [51] Oliveira MDL, Andrade CAS, Santos-Magalhães NS, Coelho LCBB, Teixeira JA,
596 Carneiro-da-Cunha MG et al. Purification of a lectin from *Eugenia uniflora* L. seeds
597 and its potential antibacterial activity. Lett Appl Microbiol. 2008; 46(3): 371-376.

598

599 [52] Sá RA, Gomes FS, Napoleão TH, Santos ND, Melo CM, Gusmão NB et al.
600 Antibacterial and antifungal activities of *Myracrodruon urundeuva* heartwood. Wood
601 Sci Technol 2009; 43(1-2): 85-95.

602

603 [53] Costa RMPB, Vaz AFM, Oliva MLV, Coelho LCBB, Correia MTS, Carneiro-da-
604 Cunha MG. A new mistletoe *Phthirusa pyrifolia* leaf lectin with antimicrobial
605 properties. Process Biochem. 2010; 45(4): 526-533.

606

607 [54] Ferreira RS, Napoleão TH, Santos AF, Sá RA, Carneiro-da-Cunha MG, Morais
608 MMC et al. Coagulant and antibacterial activities of the water-soluble seed lectin from
609 *Moringa oleifera*. Lett Appl Microbiol. 2011; 53(2): 186-192.

610

611 [55] Nunes EDS, de Souza MAA, Vaz AFM, Santana GMS, Gomes FS, Coelho LCBB
612 et al. Purification of a lectin with antibacterial activity from *Bothrops leucurus* snake
613 venom. Comp Biochem Phys B. 2011; 159(1): 57-63.

614

615 [56] Moura MC, Pontual EV, Paiva PMG, Coelho LCBB. An Outline to Corrosive
616 Bacteria. In: Microbial pathogens and strategies for combating them: science,
617 technology and education. Formatex Research Centre, Badajoz. 2013: (in press).

618

619 [57] Hasheesh WS, Mohamed RT. Bioassay of two pesticides on *Bulinus truncatus*
620 snails with emphasis on some biological and histological parameters. Pestic Biochem
621 Phys. 2011; 100(1): 1-6.

622

623 [58] Fu JM, Mai BX, Sheng GY, Zhang G, Wang XM, Peng P et al. Persistent organic
624 pollutants in environment of the Pearl River Delta, China: an overview. Chemosphere.
625 2003; 52: 1411–1422.

626

627 [59] Sharma RR, Singh D, Singh R. Biological control of postharvest diseases of fruits
628 and vegetables by microbial antagonists: A review. Biol control. 2009; 50(3): 205-221.

629

630 [60] Yoshiyama M, Kimura K. Bacteria in the gut of Japanese honeybee, *Apis cerana*
631 *japonica*, and their antagonistic effect against *Paenibacillus larvae*, the causal agent of
632 American foulbrood. J Invertebr Pathol. 2009; 102(2): 91-96.

633

634 [61] Khan MQ, Abbasi MW, Zaki MJ, Khan SA. Evaluation of *Bacillus thuringiensis*
635 isolates against root-knot nematodes following seed application in okra and mungbean.
636 Pak. J. Bot. 2010; 42(4): 2903-2910.

637

638 [62] Khedher SB, Zouari N, Messaddeq N, Schultz P, Jaoua S. Overproduction of delta-
639 endotoxins by sporeless *Bacillus thuringiensis* mutants obtained by nitrous acid
640 mutagenesis. Curr Microbiol. 2011; 62(1): 38-43.

641

642 [63] Stevens MM, Hughes PA, Mo J. Evaluation of a commercial *Bacillus*
643 *thuringiensis* var. *israelensis* formulation for the control of chironomid midge larvae

644 (Diptera: Chironomidae) in establishing rice crops in south-eastern Australia. J
645 Invertebr Pathol. 2013; 112: 9–15

646

647 [64] Araújo AP, de Melo-Santos MAV, Carlos SDO, Rios EMMM, Regis L. Evaluation
648 of an experimental product based on *Bacillus thuringiensis* sorovar. *Israelensis* against
649 *Aedes aegypti* larvae (Diptera: Culicidae). Biol Control, 2007; 41(3): 339-347.

650

651 [65] Gill SS, Cowles EA, Pietrantonio PV. The mode of action of *Bacillus thuringiensis*
652 endotoxins. Annu Rev Entomol, 1992; 37(1): 615-634.

653

654 [66] Burton SL, Ellar DJ, Li J, Derbyshire DJ. *N*-acetylgalactosamine on the putative
655 insect receptor aminopeptidase N is recognised by a site on the domain III lectin-like
656 fold of a *Bacillus thuringiensis* insecticidal toxin. J Mol Biol. 1999; 287(5): 1011-1022.

657

658 [67] Rashad FM, Saleh WD, Nasr M, Fathy HM. Identification of mosquito larvicidal
659 bacterial strains isolated from north Sinai in Egypt. AMB Express. 2012; 2(1): 1-15.