

Environmentally Favourable and Unfavorable Bacteria

Abstract

Most of bacteria can be distinguished into three groups: saprophytic; symbiotic and parasitic. Microbial communities have a vast importance to the ecosystem and can be used by humans for health or industrial purposes. Saprophytic bacteria which are the major decomposers of organic matter, can be applied in treatment of metalliferous mine or radioactive environmental wastes, biodiesel production, among others. Symbiotic bacteria live in a mutually beneficial association with other organisms providing essential nutrients to their host organisms. Some bacteria are able to cause diseases (i.e., parasitic bacteria also referred to as pathogens). Antimicrobial peptides and polypeptides such as lectins are promising raw materials for the production of new antibiotics. Lectins are able to interact with carbohydrates in bacterial cell walls and promote antibacterial activity. The aim of this chapter was to describe the importance of bacteria to environments, their use as biological control agents and the application of lectins to control pathogenic bacteria.

Keywords: environmental; bacteria; saprophytic; symbiotic; parasitic; biocontrol; lectins

1. INTRODUCTION

Bacteria are single-celled microorganisms, being classified as prokaryotes. There are over 3.6 billion years bacteria are present on Earth in almost all possible locations of life occurrence. This long co-evolution enabled bacteria to develop several beneficial relationships with the environment since they form part of this system. The number of bacterial species that have been described is low (~7,000 what do you mean by this figure??) in relation to the millions of bacteria that have been predicted to reside on Earth [1].

Comment:

Boldened lines are not clear. Author should explain.

Microbial communities have a vast importance to the ecosystem being important components of the forest ecosystem since they facilitate organic matter decomposition and nutrient cycling in the soil [2]. Free-living bacteria are of importance in agriculture as they abound in the rhizosphere (i.e, the region around the root) and have more than one mechanism of accomplishing increased plant growth, such as the production of enzymes, bioactive factors, antibiotics, metabolites as well as growth promoters [3].

Bacteria can be classified, in terms of their morphology, as bacilli (rods), cocci (spherical), spiral and many others (Figure 1). The bacillus is rod-shaped and can be found as isolated bacilli, diplobacilli or streptobacilli. A coccus is circular and can be isolated as diplococcus, tetracoccus, sarcina micrococcus, streptococcus or staphylococcus. Other bacterial shapes of low occurrence include: spirillum (*Treponema pallidum*), vibrio (*Vibrio cholerae*), transitional forms such as coccobacillus and

involution forms, a survival mechanism to adverse environmental conditions such as spores [4].

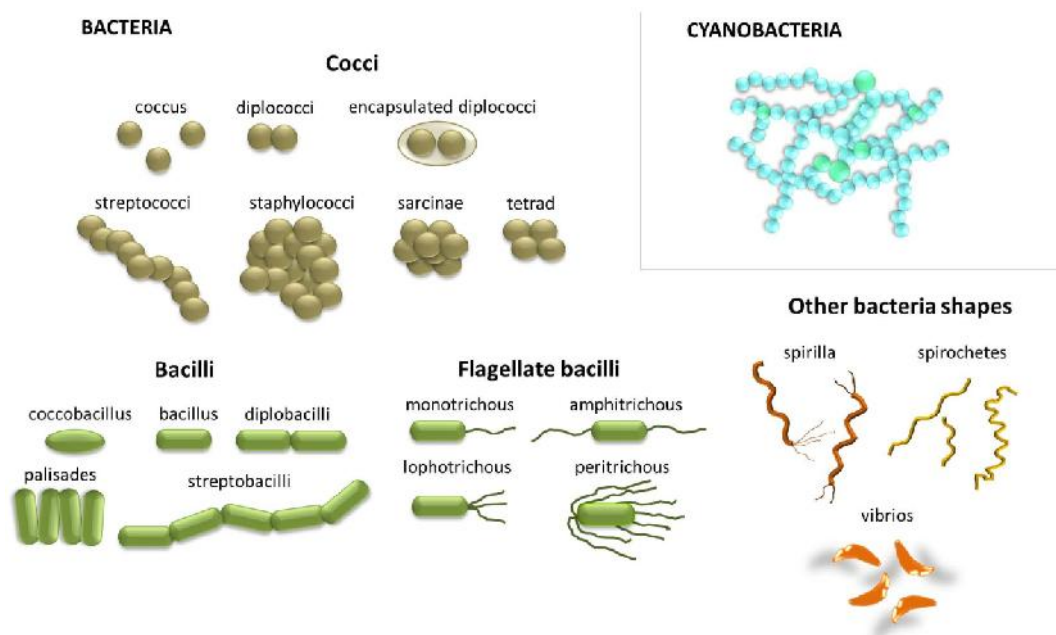


Figure 1: Types and shapes of bacteria.

Comment on figure 1:

What is the source of this figure? Author should state the source and reflect the full citation in the references.

Most bacteria are unable to manufacture their own organic food and hence, are dependent on external sources (i.e., are heterotrophic). These bacteria can be distinguished into three groups: 1) saprophytic; 2) symbiotic and 3) parasitic. Many bacteria that are associated with plants are actually saprophytic and do not harm the plant itself. However, a small number, around 100 species can cause plant diseases and thereby promoting losses in agriculture [5].

Microorganisms may be beneficial to human microbiota, complex infective bacteria that inhabit sites in and on the human body such as gut, skin, and oral

68 cavity. Special situations as in patients whose normal innate defenses fail to
69 function properly can lead to an imbalance of an individual species that are
70 pathogens in the classical sense such as *Enterobacter sp.*, *Escherichia coli* and
71 *Pseudomonas aeruginosa* [6].

72 **Comments:**

73 - The above highlighted paragraph is talking about normal
74 microflora of humans. That being the case, it is not well written.
75 The last lines (lines 67 and 68 suggest that only the three organisms
76 are opportunistic pathogens. There are many others. Author should
77 recast the entire paragraph.

78 The control of bacteria grown is a way to avoid ecosystem imbalance and
79 disease caused by some of these microorganisms. However, massive use of antibiotics
80 for this purpose has led to bacterial resistance, generated by selection processes
81 including increase in the frequency of resistance bacterial genes [7]. As an alternative,
82 antimicrobial polypeptides such as lectins have been isolated and characterized from
83 tissues and organisms from every kingdom and phylum [8]. The complete
84 understanding of mechanisms of action from new alternatives to biological control may
85 provide models and strategies for developing novel antimicrobial agents, that may also
86 increase immunity, restore potency or amplify the mechanisms of conventional
87 antibiotics, and minimize antimicrobial resistance mechanisms among pathogens.

88

89 **1. Saprophytic bacteria**

90 The saprophytic bacteria are the major decomposers of organic matter (Figure
91 2), breaking down complex mixtures into simple soluble forms and freeing
92 their atoms to be re-used by other bioprocesses [9]. The ability of some acidophilic

93 bacteria to withstand raised concentrations of certain metals through biological oxi-
94 reduction reactions has been applied in a variety of industrial fields such as treatment of
95 metalliferous mine wastes, acid mine waters and sulphurous flue gases (reference ??).
96 The Matsuo Mine in Japan applied this biological treatment system using *Thiobacillus*
97 *ferrooxidans* to treat 28m³/min of mine water at pH 2.5 oxidising more than 95 % of
98 soluble ferrous iron [10]. Microbial systems can detoxify the metal ions either by
99 extracellular biomineralization, biosorption, complexation, precipitation or intracellular
100 bioaccumulation. The cell wall reductive enzymes or soluble secreted enzymes can be
101 involved in the reductive process of metal ions by bacteria [11].

102 Some environmental factors such as availability of iron, sulphide and a micro-
103 aerobic environment are important for proliferation of the magnetotactic bacteria such
104 as *Magnetospirillum magneticum*. Magnetic minerals produced by these bacteria such
105 as greigite and biogenic magnetite form a post-depositional remnant magnetization that
106 is indicative of rapid local environmental change (reference??). These biomarkers are
107 used by archeologists to establish the chronology and environmental history of a place.
108 Linford. [12] discovered bacterial magnetosomes at the village of Yarnton (Oxford,
109 UK) suggesting a transformation of the previously dry river valley to an active flood
110 plain (reference??).

111 Radiation-resistant micro-organisms have been used in the treatment of highly
112 radioactive environmental wastes due to their ability to transform, detoxify, or
113 immobilize a variety of metallic and organic pollutants [13, 14] and also used for
114 decontamination of acid mine drainage waters through anaerobic degradation [15]. The
115 techniques traditionally applied for the treatment of radioactive environmental residuals
116 have been based on chemical methods of neutralization and precipitation (reference??).
117 These quick and effective techniques have several disadvantages, such as the need for

118 building additional plant treatments, the high cost of the chemical reagents used and the
119 generation of an important volume of sludges which need to be relocated [15].

120 Some bacteria, when subjected to any form of environmental stress, produce a
121 signal transduction cascade in which certain promoters are induced, leading to
122 expression of proteins that adjust to the ecological impact of altering the environment
123 (reference??). Bioluminescent bacteria are being used as tools to detect some special
124 compounds that are toxic and/or are of current interests as inorganic and organic
125 pollutants of water, soil and air, as well as to monitor the level of toxicity of the
126 effluents from industries into urban wastewaters, effluents from plant treatments, and
127 water (reference??). Recombinant bioluminescent bacterial strains are increasingly
128 receiving attention as environmental biosensors due to their advantages, such as high
129 sensitivity and selectivity, low costs, ease of use and short measurement times
130 (reference??). Exposure of a recombinant *Escherichia coli* strain, containing a fusion of
131 a promoter to the *Vibrio fischeri lux* genes (Ecolum-5), to a toxic or lethal condition
132 (give example of such conditions) results in a decrease in bioluminescence [16]. The
133 toxicity of benzene in air was determined using the Ecolum-5 [17].

134 Biodiesel production is been stimulated as a result of search for renewable fuels.
135 The transesterification of vegetable oils or animal fats, with ethanol or methanol
136 generates glycerol as the main byproduct. With the increasing production of biodiesel,
137 glycerol is becoming of great environmental and economical concern due to its toxicity
138 to aquatic organisms [18]; fermentation processes of this byproduct can result in value-
139 added products, such as 1,3-propanediol (1,3-PD) and ethanol. The 1, 3-PD has many
140 applications in polymers, cosmetics, foods, adhesives, lubricants, laminates, solvents,
141 antifreeze, and in medicine; ethanol could be used in the esterification of biodiesel.
142 Several bacterial strains have been isolated and characterized for their ability to convert

143 this raw glycerol into 1, 3-propanediol (1, 3-PD) and ethanol [19, 20]. Rossi. [21]
144 showed that a *Klebsiella pneumoniae* strain was able to simultaneously produce up to
145 9.4 g/L of 1,3-PD with yields of 0.41 mol product mol⁻¹ glycerol and 6.1 g/L of ethanol
146 with yields of 0.14 mol product mol⁻¹ glycerol.

147

148 2. Symbiotic bacteria

149 Symbiotic bacteria live in a mutually beneficial association with other
150 organisms. Such bacteria derive the essential nutrients (what are the essential
151 nutrients??) from their host organisms and in return, help the host through some of their
152 biological activities. Plant growth-promoting bacteria can positively provide the plant
153 with compounds which are synthesized by the bacteria or by facilitating the uptake of
154 nutrients from the environment by the plant (reference??). Nitrogen-fixing bacteria of
155 *Rhizobium* genus can fix atmospheric nitrogen and supply it to plants (Figure 2). Most
156 of biological nitrogen fixation (80%) is carried out by diazotrophic bacteria, such as the
157 *Rhizobium* genus, in symbiosis with legumes. Moreover, some bacteria which are free-
158 living in soil (e.g., cyanobacteria, *Pseudomonas*, *Azospirillum*, and *Azotobacter*) may
159 fix significant amounts of nitrogen [22].

160 Comments:

161 Lines 151 (from nitrogen fixing bacteria) and 154 (up to legumes) are not
162 well structured.

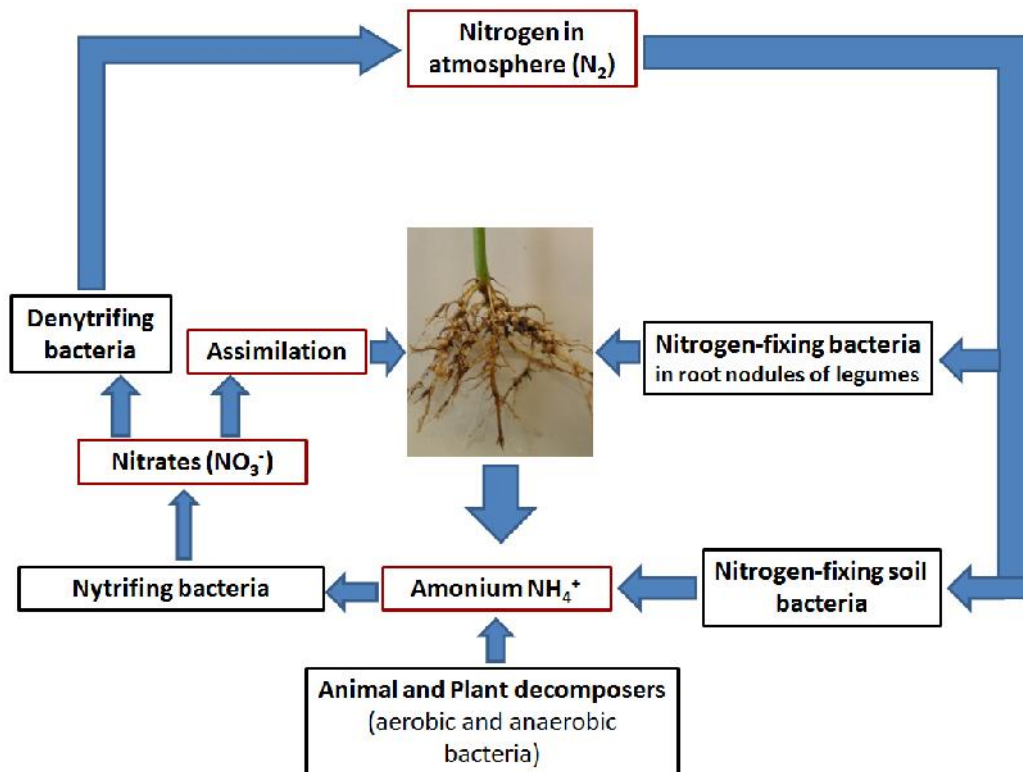


Figure 2: Importance of saprophytic and symbiotic bacteria to the nitrogen cycle.

Comment on figure 2:

What is the source of this figure? Author should state the source and reflect the full citation in the references.

Other bacteria can synthesize siderophores, chelating agents which has more affinity to metals than plant siderophores and can solubilize and sequester iron from the soil providing it to plant cells. Antibiotics, which antagonize phytopathogenic fungi and pathogenic bacteria, and synthesize phytohormones, including auxins and cytokinins can enhance various stages of plant growth, and enzymes that can modulate plant growth and development [23, 24, 25].

Comments:

Lines 163 to 168 (highlighted) do not make sense.

177

178 Bacteria may be **beneficial** when they **go into association** with other organisms
179 **(give examples of these organisms)** in the removal of contaminants from the
180 environment, a process called bioremediation [26]. **Glick. [23] reported** that the
181 **symbiotic association of the bacterium *Enterobacter cloacae* in the roots of the plant**
182 ***Brassica campestris* led** to an increase in the number of seeds that germinated and the
183 amount of biomass that the **plant was** able to attain due to reduction in the level of
184 ethylene, an inhibitor of root elongation. Furthermore, the bacterium synthesized
185 antibiotics **which inhibited** the proliferation and invasion of phytopathogens.

186 Additionally, bacteria can remove from the environment many potentially toxic
187 compounds like metals, organic compounds (such as petroleum hydrocarbons and
188 pesticides), inorganic **compounds** (such as **compounds of arsenate**, sodium, nitrate,
189 ammonia or phosphate) **as well as radioactive compounds of uranium, cesium or**
190 **strontium** [23]. The bacterium *Kluyvera ascorbata* protects *B. campestris* against high
191 levels of nickel in the soil **which produce** siderophores [27].

192 A group of bacteria **called** microbial flora (Figure 3) **are** able to beneficially
193 **affect** the host animal with contributions to nutrition, health and development by
194 **secreting Vitamin K, B12 and folate; preventing colonization by pathogens by**
195 **competing for attachment sites or for essential nutrients in the oral cavity,**
196 **intestine, skin, and vaginal epithelium; excrete ammonium that can be used to**
197 **synthesize proteins and nucleic acids; and synthesize and excrete enzymes that act**
198 **in the digestion of carbohydrates [28]. The genera present in the intestinal tract**
199 **(probiotic bacteria) generally seem to be those from the environment or the diet.**
200 **Probiotics in aquaculture can prevent pathogens proliferating in the intestinal**
201 **tract, on the superficial structures, or in the water.**

Comments:

Lines 186 to 195 (boldened) on normal flora are poorly structured.

Author should recast these lines and merge them with line 68 which

ended earlier lines on same normal flora

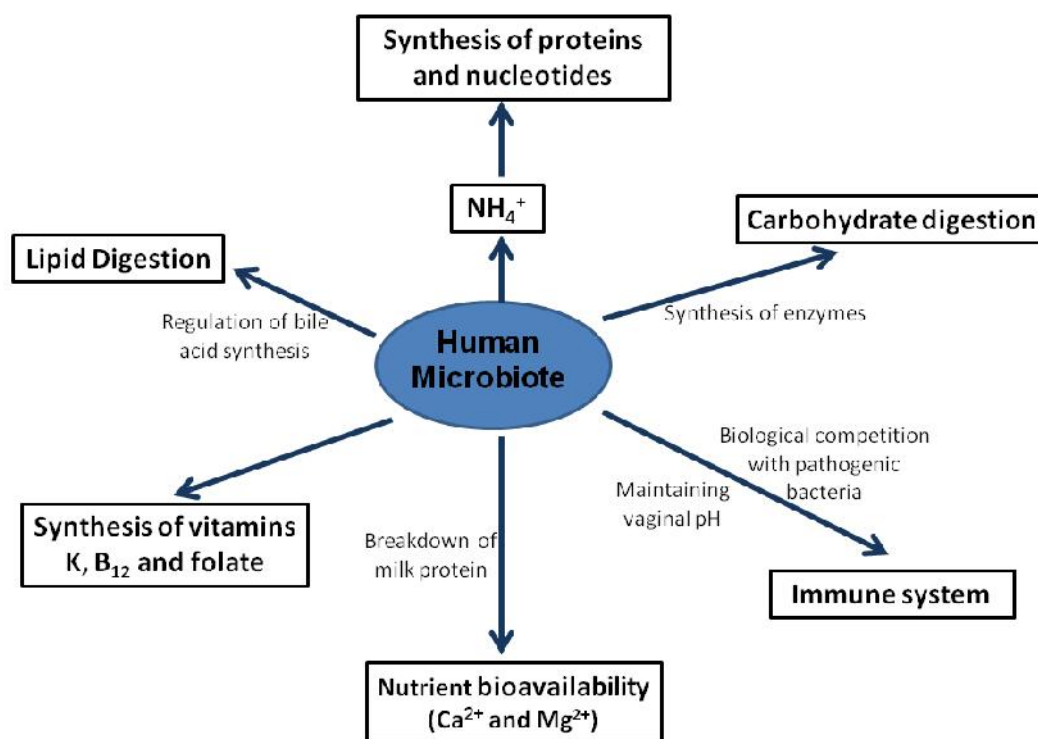


Figure 3: Benefits of human microbiote.

Comment on figure 3:

What is the source of this figure? Author should state the source and reflect the full citation in the references.

Probiotics can be used as biological control agents [29, 30]. This activity (which activity??) has been reported with lactic acid bacteria (*Lactobacillus* and *Pediococcus* genus) as biocontrol agents against the phytopathogenic and spoilage bacteria and fungi

217 [31]. A bacterial strain (*Alteromonas haloplanktis*) isolated from the gonads of Chilean
218 scallop displayed *in vitro* inhibitory activity against the pathogens *V. ordalii*, *V.*
219 *parahaemolyticus*, *V. anguillarum*, *V. alginolyticus*, and *Aeromonas hydrophila* [32].
220 The exact modes of action of the probiotics are not well understood, but it is suggested
221 that microbial populations may release chemical substances such as antibiotics,
222 lysozymes, proteases, hydrogen peroxide and organic acids that have a bactericidal or
223 bacteriostatic effects on other microbial populations. Rouse. [33] reported that a lactic
224 (what??) strain *Pediococcus pentosaceus* produced antifungal (what??). Other modes
225 of action are by competition for nutrients as iron or adhesion sites on gut or other tissue
226 surfaces, by enhancement of the immune systems of animals against infections by
227 viruses, bacteria, fungi, and parasites or by improvement of water quality (water quality,
228 how do you mean??) [29].

229 **Comments:**

230 - Lines 214 to 216 not well written

231

232 Bacteria such as *Pedobacter spp* can act on many species of microalgae of
233 red tide plankton as, for example, *Microcystis aeruginosa* [34] function for
234 controlling harmful blooms and further studies will provide new insights into its
235 role in water environment with prospects to use this algicidal bacteria as microbial
236 pesticides.

237 **Comments:**

238 Lines 232 to 236 (boldened) are confusing. Author should re-structure
239 them.

240

241 Bacteria in the aquatic environment are used as food by adults and larvae of
242 bivalve (bivalves are what??), providing nitrogen and carbon and recycling organic and
243 mineral matter released by bivalves. Marine bacteria excrete various substances,
244 including amino acids, carbohydrates, and vitamin B (which type of Vitamin B??).
245 Bacteria of bivalve microflora are important in the digestive process, metabolism and
246 metamorphosis [35]. Prieur. [36] isolated cellulolytic bacteria from the digestive tract
247 of the bivalve *Teredo navalis* that had the ability to degrade mannose and galactose.
248 Belkin. [37] showed that certain bacteria can associate with the gill tissues of a mussel
249 in deep ocean - *Bathymodiolus thermophilus* and help to fix CO₂ and thus aid
250 autotrophic metabolism. Bivalve bacteria that live in sulphide-enriched habitats are
251 important in the degradation of the organic matter through anaerobic metabolism [36].

252 Among the environmental factors that induce or influence metamorphosis of
253 many marine invertebrates, the occurrence of bacterial films and organic particles
254 trapped within the films could also be used as food by larvae ready to metamorphose.
255 Alternatively, bacteria living in the biofilms could synthesize certain compounds such
256 as low and high molecular weight polysaccharides, low molecular weight peptides and
257 neurotransmitters, diffusible into the environment, which could induce metamorphosis.
258 Water-soluble chemical compounds produced by the biofilms of two bacterial strains
259 *Macrococcus sp.* and *Bacillus sp.* induced larval settlement of the green-lipped mussel,
260 *Perna canaliculus* [38].

261

262 4. Parasitic bacteria

263 Parasitic bacteria occur in the body of animals and in plants and obtain their
264 organic food or release poisonous secretions called toxins. Many of these toxins act
265 specifically on some organisms. Thus, the majority of bacterial pathogens are highly

specialized for a limited number of eukaryotic host organisms. Plant pathogenic bacteria (Figure 4) are responsible for some of the most devastating losses of major agricultural crops and vital fruit trees, causing millions of dollars in damage annually [39]. *Ralstonia solanacearum* is a soil borne bacterium, capable of inducing disease on more than 250 plant species by invading their roots, colonizing the xylem vessels and causing a lethal wilting known as bacterial wilt disease [40]. Seeds of cashew, cocoa, coffee, pumpkin and tomato are protected from this bacterial phytopathogen because they produce oligo- and poly-saccharides that block the pathogen lectins from binding to xylem cell wall glycans [41].

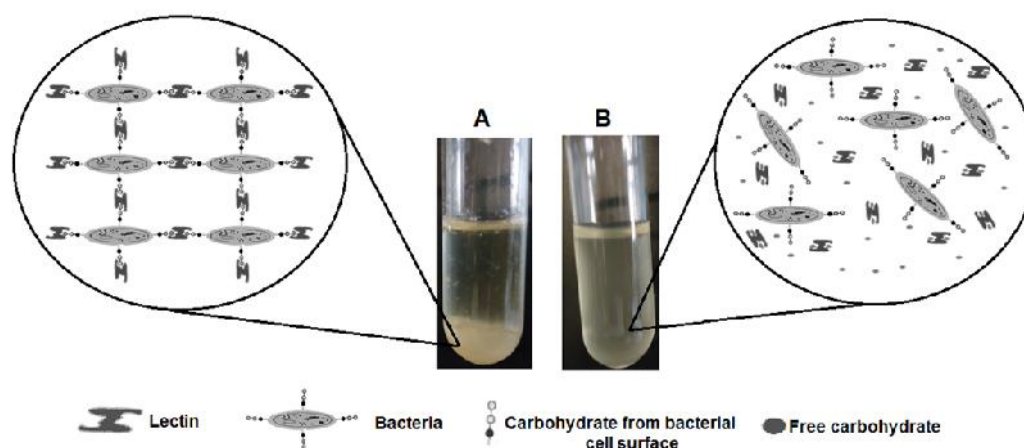


Figure 4: (A) Agglutination of *Staphylococcus aureus* by incubation with the *Schinus terebinthifolius* leaf lectin (SteLL). (B) Inhibition of agglutination in the presence of the carbohydrate inhibitor of SteLL (N-acetyl-glucosamine). The circles show schematizations of the phenomena occurring in the assay tubes.

Comment on figure 4:

What is the source of this figure? Author should state the source and reflect the full citation in the references.

285 Bacterial canker is another disease caused by a phytopathogen that has negative
286 economic impact. Bacterial canker of grapevine caused by *Xanthomonas campestris* pv.
287 *viticola* can manifest in various parts of the plant. In leaves, the symptoms are small,
288 dark and angular leaf spots that may coalesce and dry up, causing necrotic areas and leaf
289 blight. Cankers were often observed on petioles, stems and rachis and were also
290 observed in grapes [42]. Blackleg, a major bacterial disease of potato, is caused by
291 bacterial organism - *Pectobacterium carotovorum* subsp. *carotovorum*. This bacterium
292 can cause rotting of potato tubers (soft rot) during storage. Control of potato blackleg is
293 hampered by the absence of effective tools and strategies and by the dispersing ability
294 of the bacterium, being spread via surface and rain water, by aerosols and also, by
295 insects [43].

296 *Acidovorax citrulli* is the bacterial causal agent of bacterial fruit blotch, a
297 devastating disease of melon (*Cucumis melo*) and other Cucurbitaceae, and its
298 destructive potential stems from the fact that, under favorable conditions, infection
299 spreads rapidly throughout the field [44]. Symptoms of bacterial fruit blotch including
300 water-soaking and coalescing reddish-brown lesions on cotyledons and reddish-brown
301 lesions on leaves that developed along the venation [45].

302 Among pathogenic bacteria to humans, there is *Staphylococcus aureus*, which is
303 a coccus (spherical) microorganism usually with irregular distribution in clusters like
304 bunches of grapes that is responsible for many infections in humans such as
305 endocarditis, acute hematogenous osteomyelitis, meningitis or pulmonary infection.
306 *Escherichia coli*, which is a rod-shaped (a bacillus) organism, is part of the normal flora
307 and can accidentally (or opportunistically) cause diseases (such as urinary tract
308 infection, diarrhea, meningitis and septicemia). *Pseudomonas aeruginosa*, a mobile

309 aerobic bacillus organism widely distributed in nature, is found in small groups of
310 normal intestinal flora and on human skin [4].

311

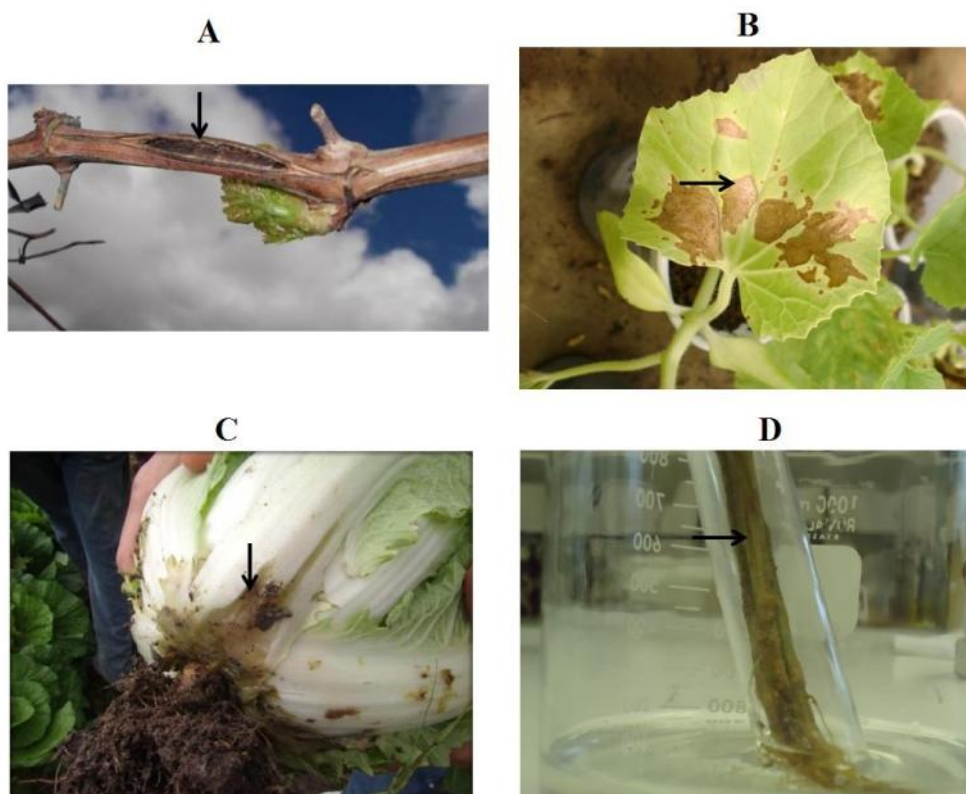
312 **5. Antibacterial lectins**

313 Antimicrobial peptides and polypeptides are promising candidates for use as raw
314 materials for producing new antibiotics. The carbohydrate recognizing proteins known
315 as lectins are noteworthy since they are able to interact with carbohydrates on bacterial
316 cellular walls [46-48]. Lectins can agglutinate cells and precipitate polysaccharides,
317 glycoproteins or glycolipids, without structural modifications [49]. The presence of
318 lectins in a sample can be evaluated in a microtiter plate through incubation with
319 erythrocytes. The linkage between lectins and glycoconjugates from erythrocyte surface
320 maintains the cells agglutinated and suspended in solution.

321 Gomes. [50] isolated an antimicrobial chitin-binding-lectin from the leaves of
322 the medicinal plant *Schinus terebinthifolius*. The authors reported that this lectin
323 showed antibacterial activity against *E. coli* (MIC, minimal inhibitory concentration, of
324 28.5 µg/ml; MBC, minimal bactericidal concentration, of 115 µg/ml), *K. pneumoniae*
325 (MIC of 3.59 µg/ml; MBC of 115 µg/ml), *Proteus mirabilis* (MIC of 3.59 µg/ml; MBC
326 of 14.37 µg/ml), *P. aeruginosa* (MIC of 1.79 µg/ml; MBC of 14.37 µg/ml), *Salmonella*
327 *enteritidis* (MIC of 0.45 µg/ml; MBC of 115 µg/ml), and *S. aureus* (MIC of 1.79 µg/ml;
328 MBC of 7.18 µg/ml).

329 Lectins can also promote agglutination of bacterial cells. Figure 5 shows the
330 agglutination of *Staphylococcus aureus* promoted by *S. terebinthifolius* leaf lectin
331 (SteLL) in assay tubes. The agglutination occurs through linkage between the
332 carbohydrate binding sites of lectin and glycoconjugates from bacterial surface as
333 schematized in Figure 5A. The bacterial agglutination was inhibited in the presence of

334 the carbohydrate inhibitor of SteLL (N-acetyl-glucosamine) as shown in Figure 5B. The
335 inhibition of bacterial agglutination by lectins after incubation with free carbohydrates
336 or glycoconjugates ensures that the binding of lectins to bacteria involves the
337 carbohydrate-binding sites.



338
339 Figure 5. Diseases caused by phytopathogenic bacteria. Bacterial canker of grapevine
340 caused by *Xanthomonas campestris* pv. *viticola* (A); blotch of melon caused by
341 *Acidovorax citrulli*, showing reddish-brown lesions on leaves (B); rooting of *Brassica*
342 *pekinensis* by *Pectobacterium carotovorum* subsp. *carotovorum* (C); tomato wilt
343 disease caused by *Ralstonia solanacearum*, showing colonization of xylem vessels (D).

344 **Comment on figure 5:**

345 **What is the source of this figure? Author should state the source and**
346 **reflect the full citation in the references.**

347 Oliveira. [51] isolated an antibacterial lectin from *Eugenia uniflora* seeds
348 (EuniSL), which inhibited the growth of *Staphylococcus aureus*, *Pseudomonas*
349 *aeruginosa* and *Klebsiella spp* with MIC of 1.5 µg/ml. Additionally, EuniSL also
350 inhibited the growth of *Bacillus subtilis*, *Streptococcus spp* and *Escherichia coli*,
351 although less efficiently (MIC of 16.5 µg/ml). The authors also showed that EuniSL was
352 able to agglutinate *S. aureus*, *Streptococcus spp*, *Klebsiella spp* and *P. aeruginosa*.

353 MuHL, a chitin-binding-lectin isolated from *Myracrodruon urundeuva*
354 heartwood, was able to inhibit the growth and agglutinate the gram-positive bacterium
355 *S. aureus* (MIC of 0.58 µg/ml; MAC, minimal agglutinating concentration, of 2.34
356 µg/ml), *Enterococcus (Streptococcus) faecalis* (MIC of 2.34 µg/ml; MAC of 4.68), *B.*
357 *subtilis* (MIC of 2.34 µg/ml; MAC of 4.68 µg/ml), and *Corynebacterium callunae* (MIC
358 of 1.17 µg/ml; MAC of 4.68 µg/ml), as well as the Gram-negative bacteria *E. coli* (MIC
359 of 9.37 µg/ml; MAC of 9.37 µg/ml), *Klebsiella pneumoniae* (MIC of 9.37 µg/ml; MAC
360 of 9.37 µg/ml) and *P. aeruginosa* (MIC of 4.68 µg/ml; MAC of 9.37 µg/ml) [52]. *M.*
361 *urundeuva* heartwood is very resistant to the deteriorative biological agents and the
362 authors pointed out that antibacterial activity of MuHL may be involved in this
363 resistance.

364 A lectin isolated from the leaf of *Phthirusa pyrifolia* also showed antibacterial
365 potentials being active against gram-positive bacteria such as *Staphylococcus*
366 *epidermidis*, *Enterococcus (Streptococcus) faecalis* and *Bacillus subtilis* and the gram-
367 negative bacterium, *Klebsiella pneumoniae* with the MIC values ranging from 250
368 µg/ml to >2000 µg/ml [53]. WSMoL, a water-soluble lectin purified from seeds of
369 *Moringa oleifera*, reduced the growth of *S. aureus* and *E. coli* and was also active
370 against ambient lake water bacteria [54].

Antimicrobial lectins have also been isolated from animals. Nunes. [55] purified a lectin from *Bothrops leucurus* snake venom, which inhibited the growth of the gram-positive bacteria *S. aureus*, *E. faecalis* and *B. subtilis* with MIC of 31.25, 62.25 and 125 µg/ml, respectively. Table 1 lists antibacterial lectins and species against which they are active.

376

377

378

Table 1. Antimicrobial activity of lectins.

Lectins	Sources	Antibacterial activities
Lectins from plants		
EuniSL MuHL	Seeds of <i>E. uniflora</i> Heartwood of <i>M. urundeuva</i>	<i>S. aureus</i> , <i>P. aeruginosa</i> and <i>Klebsiella</i> sp <i>S. aureus</i> , <i>E. faecalis</i> , <i>B. subtilis</i> , <i>C. callunae</i> , <i>E. coli</i> , <i>K. pneumoniae</i> and <i>P. aeruginosa</i>
PpyLL	Leaves of <i>P. pyrifolia</i>	<i>S. epidermidis</i> , <i>E. faecalis</i> , <i>B. subtilis</i> and <i>K. pneumonia</i>
WSMoL	Seeds of <i>M. oleifera</i>	<i>S. aureus</i> , <i>E. coli</i> and bacteria from ambient lake water
SteLL	Leaves of <i>S. terebinthifolius</i>	<i>E. coli</i> , <i>K. pneumonia</i> , <i>P. mirabilis</i> , <i>P. aeruginosa</i> , <i>S. enteritidis</i> and <i>S. aureus</i>
Lectin from animal		
BIL	<i>B. leucurus</i> snake venom	<i>S. aureus</i> , <i>E. faecalis</i> and <i>B. subtilis</i>

References: [50-52, 54, 55].

381

Moura. [56] overviewed the processes involved in biofilm formation and in biocorrosion of pipes used for oil transportation, which occurs due to fixation of bacteria that release metabolites and form biofilms thus inducing or accelerating corrosion. The authors highlighted five groups of bacteria (EPS-producing bacteria,

386 acid-producing bacteria, sulfur-oxidizing bacteria, iron-precipitating bacteria and
387 sulfate-reducing bacteria) as promoters of biocorrosion. In addition, authors pointed out
388 the use of biocides, protective coatings (antifouling) and corrosion inhibitors as the
389 main methods applied by industries to prevent corrosive bacteria from spreading. It was
390 then suggested that plant compounds, including lectins should be used for controlling
391 biocorrosion.

392

393 **6. Bacteria as biological control agents**

394

395 Pesticides are used to control organisms that are considered harmful [57].
396 However, the main problem with the use of chemical pesticides is the development of
397 resistance, resulting in reduced efficiency of the product and increased environmental
398 risk. Pesticides are one of the causes of water pollution, and some pesticides are
399 persistent organic pollutants which contribute to soil contamination [58].

400 Bacteria have been studied as safer and more eco-friendly alternatives for the
401 control of postharvest decays caused by fungi. The bacteria used for this purpose
402 usually act as antagonistic microorganisms probably through competition for nutrients
403 and space as well as by production of antibiotics, direct parasitism, and possibly
404 induced resistance in the harvested commodity. *B. subtilis* has been used as an
405 antagonist for phytopathogenic fungi that attack fruits [59].

406 Investigations carried out by Yoshiyama and Kimura [60] showed that seven
407 bacterial strains belonging to *Bacillus* genus, isolated from the digestive tract of the
408 Japanese honeybee (*Apis cerana japonica*), inhibited the development of the gram-
409 positive bacterium *Paenibacillus larvae*, the causal agent of American foulbrood. This
410 disease is contagious and affects the larval and pupal stages of honeybees. The authors

411 suggested that these *Bacillus* strains can be used for control of this disease by acting as
412 antagonists of *P. larvae*. An alternative is the use of *Bacillus thuringiensis*, a bacterium
413 that produces toxins with hemolytic and cytolytic activities. This versatile pathogen is
414 capable of infecting protozoans, nematodes, flatworms, mites and insects [61]. *B.*
415 *thuringiensis* is characterized by the production of crystals composed of proteins known
416 as deltaendotoxins that are toxic to insect pests [62].

417 *B. thuringiensis* (Bt), before 1976, was used exclusively for the control of insect
418 pests in agriculture. The discovery of a pathogenic strain against Diptera, called Bt
419 israelensis (Bti) initiated the use of this bacterium in the control of the vector disease.
420 Insects such as *Aedes aegypti* (of the Culicidae family) - vector of dengue and yellow
421 fever and insects of the Simuliidae family, transmitters of filariasis, are included in
422 the Diptera order. The use of bacteria for biological control of insect larvae from
423 Culicidae and Simuliidae family has been highlighted by having more kinds of
424 formulation (granules, powder or liquid), genetic stability, not toxic to humans, besides
425 being more advantageous considering social and environmental costs of using non-
426 selective insecticides in aquatic ecosystems [63].

427 A strategy used in Brazil as part of the National Program of Dengue Control is
428 the biological control with *Bacillus thuringiensis* serovar *israelensis* (Bti). The
429 endotoxin Cry1AC, produced during Bti sporulation, is digested by enzymes of larvae
430 midgut releasing larvicidal toxins; tablet containing spore and crystals (15%, w/w) of *B.*
431 *thuringiensis* was able to cause 100 % mortality of larvae and was suggested for use in
432 programs to control dengue vector [64]. Cry1AC has an *N*-acetylgalactosamine-specific
433 lectin domain that binds glycoconjugates at insect midgut [65, 66]. Another example of
434 larvicidal protein produced by bacterial strains is that from *Bacillus sphaericus*, which

435 was lethal to the 3rd instar larvae of *Culex pipiens*, the vector of the West Nile fever and
436 the Rift Valley fever [67].

437

438 **7. Conclusion**

439 The purpose of this review was to re-evaluate the biological importance of
440 bacteria to the environments, how they are used as biological control agents and the
441 importance of lectins in controlling pathogenic bacteria that affect animals (including
442 humans) and plants. Moreover, this review re-emphasizes biotechnological applications
443 of bacteria in many areas of human interest. This review was motivated by the lack of
444 adequate knowledge about the ecology of bacteria and use of plant lectins as
445 antimicrobial agents.

446

447 **Competing Interest**

448 Authors have declared that no competing interests exist.

449

450

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