

The simultaneous determination of some water-soluble vitamins in gum of *Acacia nilotica* by high performance liquid chromatography

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Abstract

A rapid, simple and precise method by HPLC (high performance liquid chromatography) has been developed for simultaneous determination of water-soluble vitamins as thiamine(B₁), nicotinamide(B₃), panthotenic(B₅) , pyridoxine(B₆) and biotin(B₈) in gum of *Acacia nilotica* using enzymatic hydrolysis. The method uses a C₁₈ column (4.6*150 mm, 5μm). Mobile phase such as methanol 0.1M, sodium dihydrogen phosphate (pH = 2.5), (10:90 v/v) is found most suitable for rapid separation and identification of this water – soluble vitamins. Good linearity was observed between the concentration of analytes and peak area (r = 0.9999). Each vitamin was quantitatively determined at its maximum wavelength. Recovery percentages ranged from 97% to 99%.

Keywords: Water – soluble vitamins; Gum, Acacia nilotica; HPLC.

1. Introduction

Acacia gums have a complex and branched structure, which makes them have good adhesive and cohesive properties. These properties are useful in pharmaceutical preparations. They are used as dental and other adhesives and as bulk laxatives. These hydrophilic polymers are useful as tablet binders, emulsifiers, suspending agents, gelling agents, stabilizers, thickeners, protective colloids and suspending agents keeping tablets [1]. They can also be

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used as tablet disintegrants [2]. Their adhesive property could be used in the apparatus of colostomies and also in fixing dental prosthesis [3].

For internal use, they help in the preparation of medicines to soothe coughs, diarrhea, dysentery and hemorrhages; for external use, they calm inflammations, so the presence of vitamins in *Acacia* gums is very important since vitamins are essential for human health. [4]

As far as we know, other researchers have not reported the presence or absence of vitamins B₁, B₃, B₅, B₆, B₈ in *Acacia* gum, particularly that of *Acacia nilotica*.

These vitamins are very important for the production of energy (B₁), normal growth and development (B₃), the regulation of neurotransmitters (messengers of nerve impulses) (B₅), physical balance and regulation of blood sugar (B₆) and the processing of several products such as glucose and fatty acids (B₈). [5]

Due to the nutritional importance of these vitamins, microbiological assay and several analytical methodologies have been developed for the determination of these substances in food, pharmaceutical supplements and biological fluids [6-10]. There are many analytical methods for performing the assay of vitamins in food, pharmaceutical and physiological specimens such as spectrophotometry [6,11-13] spectrophotofluorimetry [7], voltammetry [8], the gas chromatography [15-17] and high performance liquid chromatography [18-28].

Normally, it is necessary to determine more than one vitamin; the analytical method must be able to determine multiple components in complex samples, which can lead to interference in chemical analysis.

The aim of this study is to develop a rapid and reliable technique for the simultaneous determination of five water- soluble vitamins (B₁, B₃, B₅, B₆, and B₈) in gum of *Acacia nilotica* by HPLC using the enzymatic hydrolysis.

57 2. Experimental

58 2.1. Reagents and chemicals

59 Methanol was of HPLC grade. Other chemicals were of reagent grade. Sodium
60 dihydrogen phosphate (NaH_2PO_4), sodium acetate, sulfuric acid (H_2SO_4) and
61 acetic acid ($\text{C}_2\text{H}_4\text{O}_2$) (Sigma). Purified water was obtained from a Millipore
62 Milli-Q system.

63 Standards of thiamine, nicotinamide, pantothenic, pyridoxine and biotin were
64 purchased from Sigma.

65 Taka-diastase enzyme from *Aspergillus oryzae* powder, slightly beige was
66 obtained from Sigma. All chemicals and reagents used are of HPLC and were
67 used without further purification.

68 All solutions were filtered through a 0.45 μm membrane (Millipore), protected
69 from light and stored at 4°C.

70 The mobile phase of the HPLC system was comprised of pure methanol and
71 sodium dihydrogenphosphate NaH_2PO_4 (10:90 v/v).

72 2.2. Chromatographic conditions

73 The HPLC system (Agilent) was equipped with a pump type technology
74 Agilent 1200 series, a vacuum degassing unit model G1322A, a UV-VIS
75 spectrometer to 8 wavelengths, a fluorescence detector (G1321 Agilent 1200
76 Series), an analytical C_{18} column (Agilent) (4 * 150mm, 5 μm). During the
77 analysis the column was equilibrated at 30°C and a manual injector uses an
78 injection valve sample seven lane Rheodyne 7725i. The chromatographic peaks
79 were recorded and elaborated automatically by employing a computerized
80 program 'Agilent ChemStation'.

81 The analyzes were performed by gradient elution of wavelength at room
82 temperature, at a flow rate of 1 mL / min. The total execution time required is
83 less than 20 min.

84 The program of wavelength changes during elution time for five vitamins
85 determination in gum of *Acacia nilotica* shown in Table2.

86 **2.3. Standard solutions**

87 The vitamin stock solution: 100 mg / L were prepared by dissolving 10 mg of
 88 each standard in 100 mL of methanol in dark volumetric flasks. These
 89 solutions are stable hang at least one month when stored in the dark at
 90 4°C. Working solutions were prepared from stock solutions by appropriate
 91 dilution with methanol and protected from light. The following table illustrates
 92 the calibration of the analytical method:

93 Table1. Concentration of the standards used for plotting the calibration curve
 94 of five vitamins (B₁, B₃, B₅, B₆, B₈)

Vitamins	Concentrations (mg/L)
Thiamine B ₁	1.0, 5.0, 10.0, 15.0, 20.0, 30.0
Nicotinamide B ₃	1.0, 5.0, 10.0, 15.0, 20.0, 30.0
Pantothenic B ₅	1.0, 5.0, 10.0, 15.0, 20.0, 30.0
Pyridoxine B ₆	1.0, 5.0, 10.0, 15.0, 20.0, 30.0
Biotin B ₈	1.0, 2.0 ,5.0 ,10.0 ,15.0, 20.0, 30.0

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96 **2.4 Sample preparation**

97 1g of gum *Acacia nilotica* which is a fine powder was accurately weighed in a
 98 250 mL erlenmeyer flask, 10mL of sulfuric acid (1N) was added. The mixture
 99 was thoroughly shaken, after the pH was adjusted to 4.5 with sodium acetate
 100 (2.5M), then 500mg of the enzyme Taka-diastrase stirring was added .The
 101 solution was incubated at 37°C and protected from light all night. The
 102 following content was filtered on 0.45µm filter. Finally, 20 µL of the extract
 103 was injected into the HPLC system for analysis.

104

105 **3. Results and discussion**

106 The determination of vitamins in gum of *Acacia nilotica* is a complex
 107 analytical problem for several reasons: gum of *Acacia nilotica* is a very
 108 complex matrix, vitamins that are micro constituents and vitamins are easily
 109 destroyed by strong acids or alkalis, which is why we find that the enzymatic
 110 hydrolysis is a good solution for these problems.

111 First, scan analysis of standard vitamins was performed to check the optimum
 112 conditions for the detection. Wavelengths were changed according to the
 113 elution time of each vitamin, as is shown in Table 2.

114 Table 2. Program of wavelength changes during elution time for five water-
 115 soluble determinations in gum *Acacia nilotica*.

Vitamins	Time (min)	Wavelengths (nm)
Biotin (B ₈)	0.0 - 2.5	204
Nicotinamide (B ₃)	2.6 - 3.8	261
Thiamine (B ₁)	3.9 - 4.5	234
Pyridoxine (B ₆)	4.6 - 5.0	275
Pantothenic acid (B ₅)	5.1 - 7.0	210

116 The mobile phase was composed of methanol and sodium dihydrogen
 117 phosphate NaH₂PO₄ (10:90) v/v) for the determination of vitamins B₁, B₃, B₅,
 118 B₆, and B₈ in gum *Acacia nilotica*. A study of pH and the proportion of
 119 methanol and NaH₂PO₄ were necessary to improve the resolution in the gum of
 120 *Acacia nilotica* formulae. When the proportion of methanol is 20%, vitamins
 121 are eluted in less than 5 min, but there is an overlap peak of certain vitamins.

122 The pH of the mobile phase is extremely important for the separation of
123 vitamins in order to overcome this problem, a decrease in the proportion of
124 methanol by 10 %, which has the effect of providing a higher resolution but
125 against party, the time of analysis.

126 A choice of pH = 2.5 implies that most vitamins are of molecular form since
127 the pH is less than the pka of all vitamins (B₁, B₃, B₅, B₆, B₈). Figure 1 shows
128 the chromatogram of vitamins B₁, B₃, B₈ in gum of *Acacia nilotica*.

129 We note from the figure that gum of *Acacia nilotica* contains a wide range of
130 vitamin B₈. The peak of vitamin B₈ was detected at a retention time of about
131 2.2 to 2.4 min (Fig.1, 2), with minor variations on a daily basis due to
132 temperature fluctuations in the laboratory [29]. No other peaks were observed
133 at 204 nm.

134 All calculations prove that vitamin B₈ is in the order of 12,000 ppm. A part
135 from vitamin B₈, there are vitamins B₁ and B₃ but with low levels. We can
136 conclude that the method gives a good resolution of vitamin B₈.

137 ***3.1. Characteristic of the HPLC method***

138 The proposed method allows the resolution of various forms of vitamin B
139 especially B₈ in gum of *Acacia nilotica* by HPLC with UV detection.

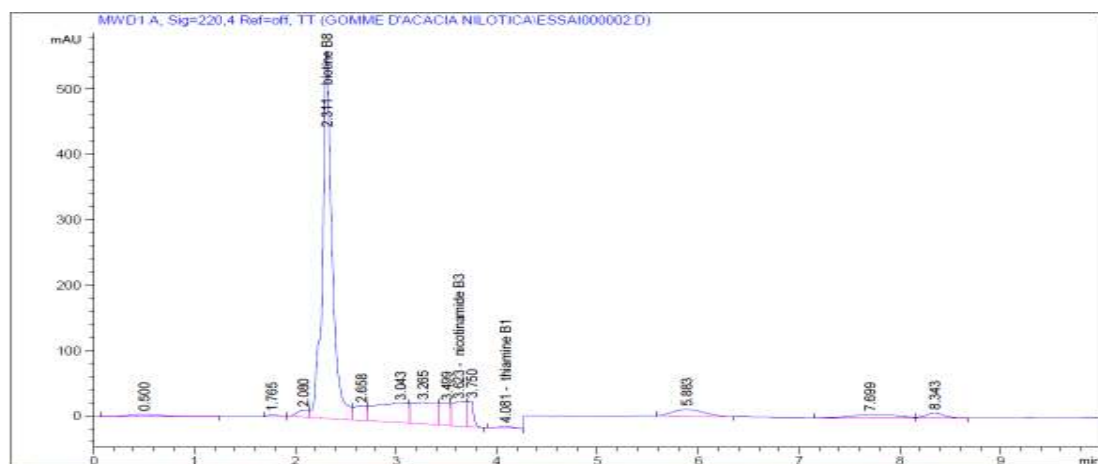
140 A reliable chromatographic assay requires an acceptable resolution, reasonable
141 retention times and good peak symmetry. Accordingly, in preliminary studies
142 optimal chromatographic conditions were investigated in gradient elution
143 system with varying wavelengths. The advantage of gradient elution is that the
144 bandwidth can be nearly constant at both early and tardative analytes.
145 Therefore an elution system of five wavelengths has been developed with a
146 beneficial effect on the sensitivity of biotin.

147 Representative chromatograms with other chromatographic parameters are
148 shown in Figure 1, 2 and Table 3.

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Amount (mg/L)



Retention time (min)

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154 **Figure 1 and 2** .Typical chromatograms of vitamin (B₈), vitamin (B₃), and
 155 vitamin (B₁)

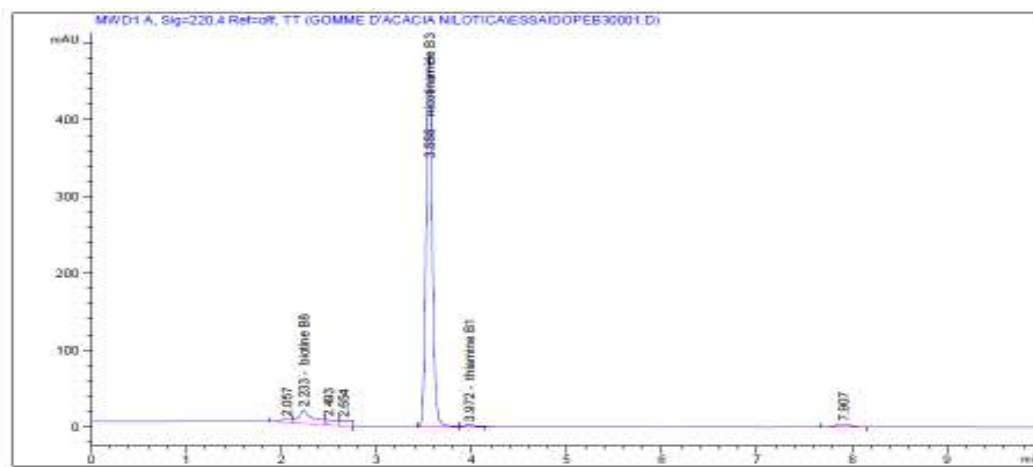
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Amount (mg/L)



Retention time (min)

Table 3. High performance liquid chromatographic parameters of the *Acacia gum* separation using the gradient elution system $t_0 = t_m = 0.56$ min.

Vitamins	Chromatographic parameters			
	T	t_R (min)	R_S	k'
Biotin B ₈	2.306	3.117	1	2.412(B3-B8)
Nicotinamide B ₃	3.512	5.27	1.5	2.087 (B1-B3)
Thiamine B ₁	4.242	6.575	1.9	3.872 (B1-B8)

t_R : retention time reduced; k' : Retention factor; R_S : Resolution factor; T: The asymmetric peak

In practice, care must be taken to keep values k' inferior to 10 for a period of reasonable analysis, values between 2 and 5 are the correct values.

3.1.1 Linearity

Six working solutions were prepared for each analyte whose range is between 1 and 30 mg /L for B₁, B₃, B₅, B₆ and seven solutions between 1 and 30 mg/L for B₈. The analysis was performed in triplicate to determine the linearity of the assay. The regression lines were calculated by the method of least squares of the areas of the peaks relative to the analyte.

The equations corresponding to the five regression analytes were

$$B_1: y = 25.82754x + 6.26753$$

$$B_3: y = 33.21959x - 1.42661$$

$$B_5: y = 7.15590x + 5.776 \cdot 10^{-1}$$

$$B_6: y = 13.70389x + 9.16444 \cdot 10^{-1}$$

$$B_8: y = 11.89793x + 2.37982$$

x: Amount et y: Area

They were consistently linear in the already mentioned range for all compounds.

181 The linearity was checked by analysis of variance of the regression (Table 4).
 182 A value of r above 0.9949 for all vitamins, ($P < 0.001$) except for thiamine with
 183 $r = 0.9781$. The coefficient of determination (r^2) is more than 95.66% for
 184 thiamine and 99.66% higher than for others. Six determinations of the same
 185 sample were performed to assess the accuracy of the method.

186 Table 4: Linearity of standard curves of vitamins B₁, B₃, B₅, B₆, and B₈

Vitamins	r	r^2	F_{exp}^a	DF^b	P
Biotin (B ₈)	0.9949	99.88	21247.5	1.5	($P < 0.001$)
Nicotinamide(B ₃)	0.9998	99.96	165492.10	1.5	($P < 0.001$)
Thiamine (B ₁)	0.9781	95.66	100026.5	1.5	($P < 0.001$)
Pyridoxine (B ₆)	0.9998	99.96	27417.53	1.5	($P < 0.001$)
Pantothenic acid (B ₅)	0.9999	99.98	7686.97	1.5	($P < 0.001$)

187 $F^a (1.5; 0.001) = 6.61$. F_{tab} and F_{exp} are tabulated and experimental
 188 Snedecor's F values, respectively in ANOVA analysis, DF^b , degrees of freedom

189 3.1.2 Accuracy and precision

190 Six determinations of the same sample were performed to assess the accuracy
 191 of the method.

192 The following table illustrates the accuracy of the method for the determination
 193 of vitamins B₁, B₃, B₅, B₆, and B₈ in gum of *Acacia nilotica*.

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198 Table 5. Peak area range and concentration, correlation data of the calibration curves
 199 and quantification limit of determined vitamins in gum of *Acacia nilotica*.

Vitamins	Concentration range (mg/L)	Surface broad peak	Correlation coefficient	Detection limits(mg/L)	Quantification limits (mg/L)
Biotin (B ₈)	1-30	59.83426- 348.10776	0.9949	0.006	0.022
Nicotinamide (B ₃)	1-30	31.45451- 1000.048	0.9998	0.008	0.028
Thiamine (B ₁)	1-30	18.14213- 714.6253	0.9781	0.012	0.042
Pyrodixine (B ₆)	1-30	14.31208- 409.0012	0.9998	0.002	0.007
Pantotenic acid (B ₅)	1-30	8.05888- 214.6708	0.9999	0.001	0.0035

200

201 3.1.3 Recovery

202 The recovery rate was tested by the standard addition procedure. One level was
 203 used for each water-soluble vitamin in gum samples (Table 6).Mean recoveries
 204 obtained were always satisfactory-higher than 99% for biotin, higher than
 205 nicotinamide 98.8 %,and higher than thiamine.

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Table 6. Study of determining recovery rate by the addition of 500µL of vitamins B₁, B₃ and B₈

N ₀ test	Biotin			Nicotinamide			Thiamine		
	Found	Recovery	%	Found	Recovery	%	Found	Recovery	%
	value	%	RSD	value	%	RSD	value	%	RSD
	(mg/L)			(mg/L)			(mg/L)		
1	1994.7	99.0	0.23	5.04	98.8	0.91	2.81e ⁻¹	97.2	1.26

4. Conclusion

The proposed method has been applied to the determination of water-soluble vitamins such as thiamine (B₁), nicotinamide (B₃), pantothenic acid (B₅), pyridoxine (B₆), biotin (B₈) in gum of *Acacia nilotica* by enzymatic hydrolysis. The chromatographic separation was performed on a C₁₈ reverse phase, and vitamins are detected at different wavelengths by UV-visible. This method is rapid, simple, and reliable and saves a significant amount of reagent.

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