1 2 3 4 5 6 7 8 9 10 11 13 14 15 16

17 18

19 20

21 22

23

24 25

26 27

28 29

30

31

32

33 34

35

36 37

Saprophytic, Symbiotic and Parasitic Bacteria: Importance to Environment, Biotecnological Applications and Biocontrol

Francis Soares Gomes¹, Emmanuel Viana Pontual¹, Luana Cassandra Breitenbach Barroso Coelho¹, and Patrícia Maria Guedes Paiva^{1*}

¹ Departamento de Bioquímica, Centro de Ciências Biológicas, Universidade Federal de Pernambuco, Avenida Moraes Rego, S/N, Cidade Universitária, Recife-PE, 50670-420, Brazil.

ABSTRACT

Microbial communities have a vast importance to the ecosystem being of use by humans for health or industrial purposes. Most bacteria can be distinguished into three groups: saprophytic; symbiotic and parasitic. Saprophytic bacteria, which are the major decomposers of organic matter, can be applied in treatment of metalliferous mine, radioactive environmental wastes, biodiesel production, among others. Symbiotic bacteria live in a mutually beneficial association with other organisms providing essential nutrients to their host organisms. However, some bacteria are able to cause diseases (i.e, parasitic bacteria also referred to as pathogens). To control the growth of these parasitic bacteria, antimicrobial peptides and polypeptides such as lectins are promising raw materials for the production of new antibiotics. Lectins are able to interact with carbohydrates in bacterial cell walls and promote antibacterial activity. The aim of this review was to describe the importance of bacteria to environments, their use as biological control agents and the application of lectins to control pathogenic bacteria.

Keywords: environmental, bacteria, saprophytic, symbiotic, parasitic, biocontrol, lectins

1. INTRODUCTION

Bacteria are single-celled microorganisms classified as prokaryotes. There are over 3.6 billion years bacteria are present on Earth, colonizing almost every possible occurrence of life. This long co-evolution enabled bacteria to develop several beneficial relationships with the environment since they form part of this system. The number of bacterial species that have been described is low (~7,000) in relation to the millions of bacteria that have been predicted to reside on Earth [1].

Microbial communities have vast importance to the ecosystem. They are important components of the forest ecosystem since they facilitate organic matter decomposition and nutrient cycling in the soil [2]. Free-living bacteria are of importance in agriculture as they abound in the rhizosphere (i.e., the region around the root) and have more than one mechanism of accomplishing increased plant growth, such as the production of enzymes, bioactive factors, antibiotics, metabolites as well as growth promoters [3].

Bacteria in terms of their morphology are classified as bacilli (rods), cocci (spherical), spiral and many others (Figure 1). The bacillus is rod-shaped and found as isolated bacilli, diplobacilli or streptobacilli. A coccus is circular and can be isolated as diplococcus, tetracoccus, sarcina micrococcus, streptococcus or staphylococcus. Other bacterial shapes

of low occurrence include spirillum (*Treponema pallidum*), vibrio (*Vibrio cholerae*), transitional forms such as coccobacillus and involution forms, a survival mechanism to adverse environmental conditions such as spores [4].

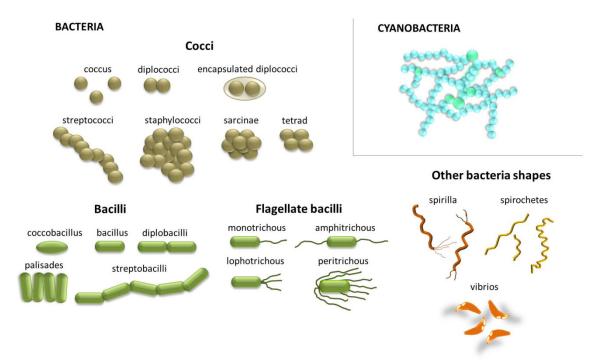


Fig. 1: Types and shapes of bacteria (cyanobacteria, cocci, bacilli, spiral). Source: photo courtesy of Dr. Rosiely Felix Bezerra.

Most bacteria are unable to manufacture their own organic food and hence, are dependent on external sources (i.e., are heterotrophic). These bacteria can be distinguished into three groups: 1) saprophytic; 2) symbiotic and 3) parasitic. Many bacteria that are associated with plants are actually saprophytic and do not harm the plant itself. However, a small number, around 100 species can cause plant diseases and thereby promoting losses in agriculture [5].

Microorganisms may be beneficial to human microbiota, complex infective bacteria that inhabit sites in and on the human body such as gut, skin, and oral cavity. Special situations as in patients whose normal innate defenses fail to function properly can lead to an imbalance of an individual species that are pathogens in the classical sense such as Enterobacter sp., Escherichia coli and Pseudomonas aeruginosa, among others [6].

The control of bacteria grown is a way to avoid ecosystem imbalance and disease caused by some of these microorganisms. However, massive use of antibiotics for this purpose has led to bacterial resistance, generated by selection processes including increase in the frequency of resistance bacterial genes [7]. As an alternative, antimicrobial polypeptides such as lectins have been isolated and characterized from tissues and organisms from every kingdom and phylum [8]. The complete understanding of mechanisms of action from new alternatives to biological control may provide models and strategies for developing novel antimicrobial agents that may also increase immunity, restore potency or amplify the mechanisms of conventional antibiotics, and minimize antimicrobial resistance mechanisms among pathogens.

1. Saprophytic bacteria

 The saprophytic bacteria are the major decomposers of organic matter (Figure 2), breaking down complex mixtures such as cellulose, hemicelluloses, lignin and proteins into simple soluble forms (catabolic reactions) and freeing their atoms to be re-used by other bioprocesses (anabolic reactions) [9]. The biological relevance of saprophytic bacteria to the environment has been used in several biotechnological applications. The ability of some acidophilic bacteria to withstand raised concentrations of certain metals through biological oxi-redution reactions has been applied in a variety of industrial fields such as treatment of metalliferous mine wastes, acid mine waters and sulphurous flue gases [10, 11]. The Matsuo Mine in Japan applied this biological treatment system using *Thiobacillus ferrooxidans* to treat 28m³/min of mine water at pH 2.5 oxidising more than 95 % of soluble ferrous iron [10]. Microbial systems can detoxify the metal ions by either extracellular biomineralization, biosorption, complexation, precipitation or intracellular bioaccumulation. The cell wall reductive enzymes or soluble secreted enzymes can be involved in the reductive process of metal ions by bacteria [11].

Some environmental factors such as availability of iron, sulphide and a micro-aerobic environment are important for proliferation of the magnetotactic bacteria such as Magnetospirillum magneticum. Magnetic minerals produced by these bacteria such as greigite and biogenic magnetite form a post-depositional remnant magnetization that is indicative of rapid local environmental change [12]. Archeologists to establish the chronology and environmental history of a place use these biomarkers. Linford. [12] discovered bacterial magnetosomes at the village of Yarnton (Oxford, UK) suggesting a transformation of the previously dry river valley to an active flood plain.

Radiation-resistant microorganisms have been used in the treatment of highly radioactive environmental wastes due to their ability to transform, detoxify, or immobilize a variety of metallic and organic pollutants [13, 14] and used for decontamination of acid mine drainage waters through anaerobic degradation [15]. The techniques traditionally applied for the treatment of radioactive environmental residuals have been based on chemical methods of neutralization and precipitation [13-15]. These quick and effective techniques have several disadvantages, such as the need for building additional plant treatments, the high cost of the chemical reagents used and the generation of an important volume of sludges that need to be relocated [15].

Some bacteria, when subjected to any form of environmental stress, produce a signal transduction cascade in which certain promoters are induced, leading to expression of proteins that adjust to the ecological impact of altering the environment [16, 17]. Bioluminescent bacteria have being used as tools to detect some special compounds that are toxic and/or are of current interests as inorganic and organic pollutants of water, soil and air, as well as to monitor the level of toxicity of the effluents from industries into urban wastewaters, effluents from plant treatments, and water [16, 17]. Recombinant bioluminescent bacterial strains are increasingly receiving attention as environmental biosensors due to their advantages, such as high sensitivity and selectivity, low costs, ease of use and short measurement times. Exposure of a recombinant E. coli strain, containing a fusion of a promoter to the Vibrio fisheri lux genes (Ecolum-5), to a toxic or lethal condition (DNA, superoxide or protein/membrane damage) results in a decrease in bioluminescence [16]. The toxicity of benzene in air was determined using the Ecolum-5 [17].

Biodiesel production is been stimulated because of search for renewable fuels. The transesterification of vegetable oils or animal fats, with ethanol or methanol generates glycerol as the main byproduct. With the increasing production of biodiesel, glycerol is becoming of great environmental and economical concern due to its toxicity to aquatic organisms [18]; fermentation processes of this byproduct can result in value-added products, such as 1,3-propanediol (1,3-PD) and ethanol. The 1, 3-PD has many applications in polymers, cosmetics, foods, adhesives, lubricants, laminates, solvents, antifreeze, and in

medicine; ethanol could be used in the esterification of biodiesel. Several bacterial strains have been isolated and characterized for their ability to convert this raw glycerol into 1, 3-propanediol (1, 3-PD) and ethanol [19, 20]. Rossi. [21] showed that a *Klebsiella pneumoniae* strain was able to simultaneously produce up to 9.4 g/L of 1,3-PD with yields of 0.41 mol product mol⁻¹ glycerol and 6.1 g/L of ethanol with yields of 0.14 mol product mol⁻¹ glycerol.

2. Symbiotic bacteria

Symbiotic bacteria live in a mutually beneficial association with other organisms. Such bacteria derive the essential nutrients (proteins, carbohydrates and lipids) from their host organisms and in return, help the host through some of their biological activities. Plant growth-promoting bacteria can positively provide the plant with compounds which are synthesized by the bacteria or by facilitating the uptake of nutrients from the environment by the plant [22, 23]. Nitrogen-fixing bacteria of *Rhizobium* genus can fix atmospheric nitrogen and supply it to plants (Figure 2). Most of biological nitrogen fixation (80%) is carried out by diazotrophic bacteria, such as the *Rhizobium* genus, in symbiosis with legumes. Moreover, some bacteria which are free-living in soil (e.g., cyanobacteria, *Pseudomonas, Azospirillum*, and *Azotobacter*) may fix significant amounts of nitrogen [22].

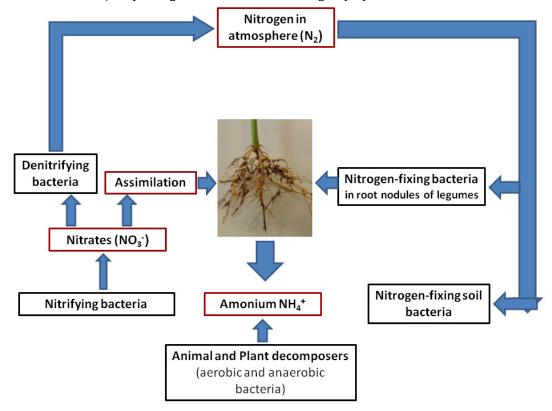


Figure 2: Importance of saprophytic and symbiotic bacteria to the nitrogen cycle. Source: authors.

Other bacteria can synthesize many compounds with positive effect on plants such as siderophores, chelating agents which has more affinity to metals than plant siderophores and can solubilize and sequester iron from the soil providing it to plant cells; antibiotics, which antagonize phytopathogenic fungi and pathogenic bacteria; phytohormones, including

auxins and cytokinins, enhancing various stages of plant growth; and enzymes that can modulate plant growth and development [23, 24, 25].

Bacteria may be beneficial when they go into association with other organisms (plants and animals) in the removal of contaminants from the environment, a process called bioremediation [26]. Glick. [23] reported that the symbiotic association of the bacterium Enterobacter cloacae in the roots of the plant Brassica campestris led to an increase in the number of seeds that germinated and the amount of biomass that the plant was able to attain due to reduction in the level of ethylene, an inhibitor of root elongation. Furthermore, the bacterium synthesized antibiotics which inhibited the proliferation and invasion of phytopathogens.

Additionally, bacteria can remove from the environment many potentially toxic compounds like metals, organic compounds (such as petroleum hydrocarbons and pesticides), inorganic compounds (such as compounds of arsenate, sodium, nitrate, ammonia or phosphate) as well as radioactive compounds of uranium, cesium or strontium [23]. The bacterium *Kluyvera ascorbata* protects *B. campestris* against high levels of nickel in the soil, which produce siderophores [27].

A group of bacteria called microbial flora (Figure 3) are able to affect beneficially the host animal with contributions to nutrition, health and development. Microbial flora can secret vitamins; participate in the synthesis of proteins and nucleic acids; and act in the digestion of carbohydrates. The genera present in the intestinal tract (probiotic bacteria) generally seem to be those from the environment or the diet. Furthermore, microbial flora prevent colonization by pathogens by competing for attachment sites or for essential nutrients in the oral cavity, intestine, skin, and vaginal epithelium. Probiotics in aquaculture of genus *Lactobacillus* can prevent pathogens proliferating in the intestinal tract, on the superficial structures, or in the water [28].

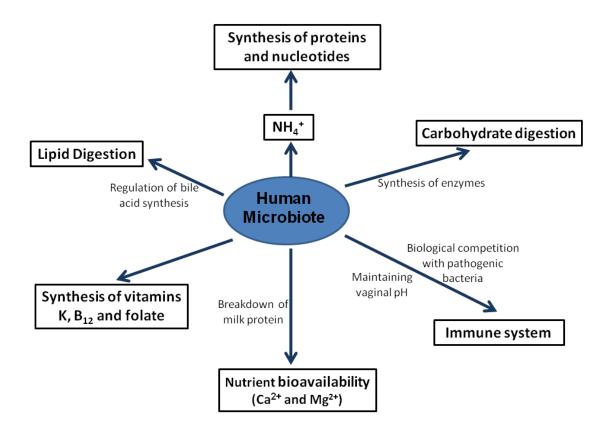


Fig. 3: Benefits of human microbiote. Source: authors.

Probiotics can be used as biological control agents of other bacteria and fungi [29, 30]. This function has been reported with lactic acid bacteria (*Lactobacillus* and *Pediococcus* genus) as biocontrol agents against the phytopathogenic and spoilage bacteria and fungi [31]. A bacterial strain (*Alteromonas haloplanktis*) isolated from the gonads of Chilean scallop displayed *in vitro* inhibitory activity against the pathogens *V. ordalii, V. parahaemolyticus, V. anguillarum, V. alginolyticus*, and *Aeromonas hydrophila* [32]. The exact modes of action of the probiotics are not well understood, but it is suggested that microbial populations may release chemical substances such as antibiotics, lysozymes, proteases, hydrogen peroxide and organic acids that have a bactericidal or bacteriostatic effects on other microbial populations. Rouse. [33] reported that the lactic acid bacteria *Pediococcus pentosaceous* produced antifungal peptides (not completely characterized), with potential applications in the food industry to prevent fungal spoilage of food. Other modes of action are by competition for nutrients as iron or adhesion sites on gut or other tissue surfaces, or by enhancement of the immune systems of animals against infections by viruses, bacteria, fungi, and parasites [29].

Some bacteria may play an important role in the control of harmful algae blooms. Bacteria such as *Pedobacter* spp can act on many species of microalgae of red tide plankton such as, for example, *Microcyctis aeruginosa* [34]. Further studies will provide new insights into its role in water environment with prospects to use these algicidal bacteria as microbial pesticides.

Bacteria of microflora from mollusks such as bivalve are important in the digestive process, metabolism and metamorphosis [35]. Bacteria in the aquatic environment are used as food by adults and larvae of bivalve, providing nitrogen and carbon and recycling organic and mineral matter released by these aquatic organisms. Furthermore, marine bacteria, such as

cyanobacteria, excrete various substances, including amino acids, carbohydrates, and vitamin B12. Prieur. [36] isolated cellulolytic bacteria from the digestive tract of the bivalve Teredo navalis that had the ability to degrade mannose and galactose. Belkin. [37] showed that certain bacteria can associate with the gill tissues of a mussel in deep ocean - Bathymodiolus thermophilus and help to fix CO₂ and thus aid autotrophic metabolism. Bivalve bacteria that live in sulphide-enriched habitats are important in the degradation of the organic matter through anaerobic metabolism [36].

Among the environmental factors that induce or influence metamorphosis of many marine invertebrates, the occurrence of bacterial films and organic particles trapped within the films could also be used as food by larvae ready to metamorphose. Alternatively, bacteria living in the biofilms could synthesize certain compounds such as low and high molecular weight polysaccharides, low molecular weight peptides and neurotransmitters, diffusible into the environment, which could induce metamorphosis. Water-soluble chemical compounds produced by the biofilms of two bacterial strains *Macrococcus sp.* and *Bacillus* sp. induced larval settlement of the green-lipped mussel, *Perna canaliculus* [38].

4. Parasitic bacteria

 Parasitic bacteria occur in the body of animals and plants and obtain their organic food or release poisonous secretions called toxins. Many of these toxins act specifically on some organisms. Thus, the majority of bacterial pathogens are highly specialized for a limited number of eukaryotic host organisms. Plant pathogenic bacteria (Figure 4) are responsible for some of the most devastating losses of major agricultural crops and vital fruit trees, at a cost price of millions of dollars annually [39]. Ralstonia solanacearum is a soil borne bacterium, capable of inducing disease on more than 250 plant species by invading their roots, colonizing the xylem vessels and causing a lethal wilting known as bacterial wilt disease [40]. Seeds of cashew, cocoa, coffee, pumpkin and tomato are protected from this bacterial phytopathogen because they produce oligo- and poly-saccharides that block the pathogen lectins from binding to xylem cell wall glycans [41].

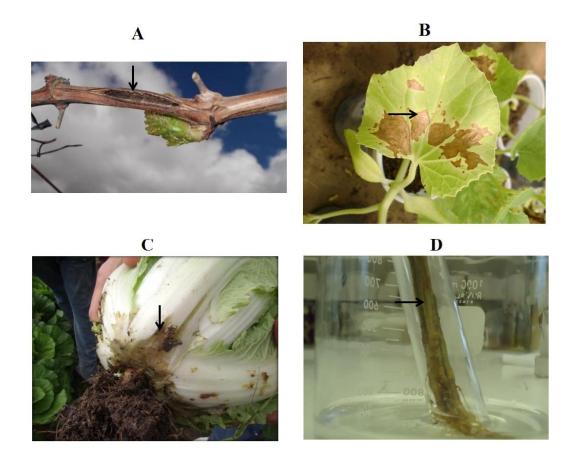


Fig. 4. Diseases caused by phytopathogenic bacteria. Bacterial canker of grapevine caused by *Xanthomonas campestris* pv. *viticola* (A); blotch of melon caused by *Acidovorax citrulli*, showing reddish-brown lesions on leaves (B); rooting of *Brassica pekinensis* by *Pectobacterium carotovorum* subsp. *carotovorum* (C); tomato wilt disease caused by *Ralstonia solanacearum*, showing colonization of xylem vessels (D). Source: photo courtesy of Prof. Dr. Elineide Barbosa de Souza, from the *Laboratório de Fitobacteriologia, Universidade Federal Rural de Pernambuco*, Recife.

Bacterial canker is another disease caused by a phytopathogen that has negative economic impact. Bacterial canker of grapevine caused by *Xanthomonas campestris* pv. *viticola* can manifest in various parts of the plant. In leaves, the symptoms are small, dark and angular leaf spots that may coalesce and dry up, causing necrotic areas and leaf blight. Cankers were often observed on petioles, stems and rachis and were also observed in grapes [42]. Blackleg, a major bacterial disease of potato, is caused by bacterial organism - *Pectobacterium carotovorum* subsp. *carotovorum*. This bacterium can cause rotting of potato tubers (soft rot) during storage. Control of potato blackleg is hampered by the absence of effective tools and strategies and by the dispersing ability of the bacterium, being spread via surface and rain water, by aerosols and also, by insects [43].

Acidovorax citrulli is the bacterial causal agent of bacterial fruit blotch, a devastating disease of melon (*Cucumis melo*) and other plants from the same family. Its destructive potential stems from the fact that, under favorable conditions to bacterial growth, infection spreads rapidly throughout the field [44]. Symptoms of bacterial fruit blotch including water-soaking

and coalescing reddish-brown lesions on cotyledons and reddish-brown lesions on leaves that developed along the venation [45].

Among pathogenic bacteria to humans, there is *Staphylococcus aureus*, which is a coccus (spherical) microorganism usually with irregular distribution in clusters like bunches of grapes that is responsible for many infections in humans such as endocarditis, acute hematogenous osteomyelitis, meningitis or pulmonary infection. *E. coli*, which is a rod-shaped (a bacillus) organism, is part of the normal flora but can opportunistically cause diseases (such as urinary tract infection, diarrhea, meningitis and septicemia). *P. aeruginosa*, a mobile aerobic bacillus organism widely distributed in nature, is found in small groups of normal intestinal flora and on human skin. *P. aeruginosa* rarely causes disease in a healthy immune system, but in individuals with compromised immune systems, this bacterium infects the respiratory tract, urinary tract, burns, and also causes other blood infections [4].

5. Antibacterial lectins

Antimicrobial peptides and polypeptides are promising candidates for use as raw materials for producing new antibiotics. The carbohydrate recognizing proteins known as lectins are noteworthy since they are able to interact with carbohydrates on bacterial cellular walls [46-48]. Lectins can agglutinate cells and precipitate polysaccharides, glycoproteins or glycolipids, without structural modifications [49]. The presence of lectins in a sample can be evaluated in a microtiter plate through incubation with erythrocytes. The linkage between lectins and glycoconjugates from erythrocyte surface maintains the cells agglutinated and suspended in solution.

Gomes. [50] isolated an antimicrobial chitin-binding-lectin from the leaves of the medicinal plant *Schinus terebinthifolius*. The authors reported that this lectin showed antibacterial activity against *E. coli* (MIC, minimal inhibitory concentration, of 28.5 μg/ml; MBC, minimal bactericidal concentration, of 115 μg/ml), *K. pneumoniae* (MIC of 3.59 μg/ml; MBC of 115 μg/ml), *Proteus mirabilis* (MIC of 3.59 μg/ml; MBC of 14.37 μg/ml), *P. aeruginosa* (MIC of 1.79 μg/ml; MBC of 14.37 μg/ml), *Salmonella enteritidis* (MIC of 0.45 μg/ml; MBC of 115 μg/ml), and *S. aureus* (MIC of 1.79 μg/ml; MBC of 7.18 μg/ml).

Lectins can also promote agglutination of bacterial cells [47]. Figure 5 shows the agglutination of *S. aureus* promoted by *S. terebinthifolius* leaf lectin (SteLL) in assay tubes. The agglutination occurs through linkage between the carbohydrate binding sites of lectin and glycoconjugates from bacterial surface as schematized in Figure 5A. The bacterial agglutination was inhibited in the presence of the carbohydrate inhibitor of SteLL (N-acetyl-glucosamine) as shown in Figure 5B. The inhibition of bacterial agglutination by lectins after incubation with free carbohydrates or glycoconjugates ensures that the binding of lectins to bacteria involves the carbohydrate-binding sites.

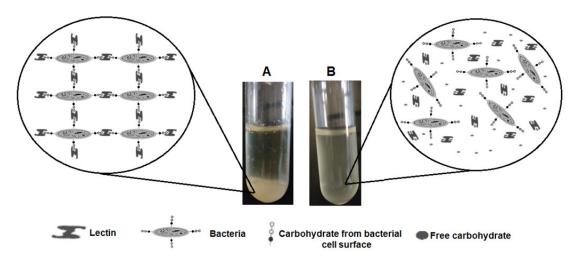


Fig. 5: (A) Agglutination of *Staphylococcus aureus* by incubation with the *Schinus terebinthifolius* leaf lectin (SteLL). (B) Inhibition of agglutination in the presence of the carbohydrate inhibitor of SteLL (N-acetyl-glucosamine). The circles show schematizations of the phenomena occurring in the assay tubes. Source: authors

Oliveira. [51] isolated an antibacterial lectin from Eugenia uniflora seeds (EuniSL), which inhibited the growth of S. aureus, P. aeruginosa and Klebsiella spp with MIC of 1.5 μg/ml. Additionally, EuniSL also inhibited the growth of Bacillus subtilis, Streptococcus spp and E. coli, although less efficiently (MIC of 16.5 µg/ml). The authors also showed that EuniSL was able to agglutinate S. aureus, Streptococcus spp, Klebsiella spp and P. aeruginosa. MuHL, a chitin-binding-lectin isolated from Myracrodruon urundeuva heartwood, was able to inhibit the growth and agglutinate the gram-positive bacterium S. aureus (MIC of 0.58 μq/ml; MAC, minimal agglutinating concentration, of 2.34 μg/ml), Enterococcus (Streptococcus) faecalis (MIC of 2.34 μg/ml; MAC of 4.68), B. subtilis (MIC of 2.34 μg/ml; MAC of 4.68 μg/ml), and Corynebacterium callunae (MIC of 1.17 μg/ml; MAC of 4.68 μg/ml), as well as the gramnegative bacteria E. coli (MIC of 9.37 μg/ml; MAC of 9.37 μg/ml), K. pneumoniae (MIC of 9.37 μg/ml; MAC of 9.37 μg/ml) and P. aeruginosa (MIC of 4.68 μg/ml; MAC of 9.37 μg/ml) [52]. M. urundeuva heartwood is very resistant to the deteriorative biological agents and the authors pointed out that antibacterial activity of MuHL may be involved in this resistance. A lectin isolated from the leaf of *Phthirusa pyrifolia* also showed antibacterial potentials being active against gram-positive bacteria such as Staphylococcus epidermidis, Enterococcus (Streptococcus) faecalis and B. subtilis and the gram-negative bacterium, K. pneumoniae with the MIC values ranging from 250 μg/ml to >2000 μg/ml [53]. WSMoL, a water-soluble lectin purified from seeds of Moringa oleifera, reduced the growth of S. aureus and E. coli and was also active against ambient lake water bacteria [54].

Antimicrobial lectins have also been isolated from animals. Nunes. [55] purified a lectin from *Bothrops leucurus* snake venom, which inhibited the growth of the gram-positive bacteria S. *aureus*, E. *faecalis* and B. *subtilis* with MIC of 31.25, 62.25 and 125 μ g/ml, respectively. Table 1 lists antibacterial lectins and species against which they are active.

Table 1. Antimicrobial activity of lectins.

294

295 296

297

298 299 300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316 317

318

319

320

321

322

323

Lectin <mark>s</mark>	Source <mark>s</mark>	Antibacterial activiti <mark>es</mark>	
Lectins fro	m plants		
EuniSL	Seeds of E. uniflora	S. aureus. P. aeruginosa and Klebsiella sp	

MuHL	Heartwood of <i>M.</i> urundeuva	S. aureus, E. faecalis, B. subtilis, C. callunae, E. coli, K. pneumoniae and P. aeruginosa
PpyLL	Leaves of P. pyrifolia	S. epidermidis, E. faecalis, B. subtilis and K. pneumonia
WSMoL	Seeds of M. oleifera	S. aureus, E. coli and bacteria from ambient lake water
SteLL	Leaves of <i>S. terebinthifolius</i>	E. coli, K. pneumonia, P. mirabilis, P. aeruginosa, S. enteritidis and S. aureus

Lectin from animal

BIL	B.	leucurus	snake	S. aureus, E. faecalis and B. subtilis
	vend	om		

References: [50-52, 54, 55].

Moura. [56] overviewed the processes involved in biofilm formation and in biocorrosion of pipes used for oil transportation, which occurs due to fixation of bacteria that release metabolites and form biofilms thus inducing or accelerating corrosion. The authors highlighted five groups of bacteria (EPS-producing bacteria, acid-producing bacteria, sulfur-oxidizing bacteria, iron-precipitating bacteria and sulfate-reducing bacteria) as promoters of biocorrosion. In addition, authors pointed out the use of biocides, protective coatings (antifouling) and corrosion inhibitors as the main methods applied by industries to prevent corrosive bacteria from spreading. It was then suggested that plant compounds, including lectins should be used for controlling biocorrosion

6. Bacteria as biological control agents

Pesticides are used to control organisms that are considered harmful [57]. However, the main problem with the use of chemical pesticides is the development of resistance, resulting in reduced efficiency of the product and increased environmental risk. Pesticides are one of the causes of water pollution, and some pesticides are persistent organic pollutants which contribute to soil contamination [58].

Bacteria have been studied as safer and more eco-friendly alternatives for the control of postharvest decays caused by fungi. The bacteria used for this purpose usually act as antagonistic microorganisms probably through competition for nutrients and space as well as by production of antibiotics, direct parasitism, and possibly induced resistance in the harvested commodity. *B. subtilis* has been used as an antagonist for phytopathogenic fungi that attack fruits [59].

Investigations carried out by Yoshiyama and Kimura [60] showed that seven bacterial strains belonging to *Bacillus* genus, isolated from the digestive tract of the Japanese honeybee (*Apis cerana japonica*), inhibited the development of the gram-positive bacterium *Paenibacillus larvae*, the causal agent of American foulbrood. This disease is contagious and affects the larval and pupal stages of honeybees. The authors suggested that these *Bacillus* strains can be used for control of this disease by acting as antagonists of *P. larvae*. An alternative is the use of *Bacillus thuringiensis*, a bacterium that produces toxins with hemolytic and cytolytic activities. This versatile pathogen is capable of infecting protozoans, nematodes, flatworms, mites and insects [61]. *B. thuringiensis* is characterized by the production of crystals composed of proteins known as deltaendotoxins that are toxic to insect pests [62].

B. thuringiensis (Bt), before 1976, was used exclusively for the control of insect pests in agriculture. The discovery of a pathogenic strain against Diptera, called Bt israelensis (Bti) initiated the use of this bacterium in the control of the vector disease.

Insects such as *Aedes* aegypti (of the Culicidae family) - vector of dengue and yellow feverand *Simulium* spp. (of Simuliidae family), transmitters of filariasis, are included in the Diptera order. The use of bacteria for biological control of insect larvae from Culicidae and Simuliidae family has been highlighted by having more kinds of formulation (granules, powder or liquid), genetic stability, not toxic to humans, besides being more advantageous considering social and environmental costs of using non-selective insecticides in aquatic ecosystems [63].

A strategy used in Brazil as part of the National Program of Dengue Control is the biological control with *Bacillus thuringiensis* serovar *israelensis* (Bti). The endotoxin Cry1AC, produced during Bti sporulation, is digested by enzymes of larvae midgut releasing larvicidal toxins; tablet containing spore and crystals (15%, w/w) of *B. thuringiensis* was able to cause 100 % mortality of larvae and was suggested for use in programs to control dengue vector [64]. Cry1AC has an *N*-acetylgalactosamine-specific lectin domain that binds glycoconjugates at insect midgut [65, 66]. Another example of larvicidal protein produced by bacterial strains is that from *Bacillus sphaericus*, which was lethal to the 3rd instar larvae of *Culex pipiens*, the vector of the West Nile fever and the Rift Valley fever [67].

7. CONCLUSION

The purpose of this review was to re-evaluate the biological importance of bacteria to the environments, how they are used as biological control agents and the importance of lectins in controlling pathogenic bacteria that affect animals (including humans) and plants. Moreover, this review re-emphasizes biotechnological applications of bacteria in many areas of human interest. This review was motivated by the lack of adequate knowledge about the ecology of bacteria and use of plant lectins as antimicrobial agents.

ACKNOWLEDGMENTS

The authors express their gratitude to the *Conselho Nacional de Desenvolvimento Científico e Tecnológico* (CNPq) for research grants and fellowships (L.C.B.B. Coelho and P.M.G. Paiva), *Coordenação de Aperfeiçoamento de Pessoal de Nível Superior* (CAPES), *Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco* (FACEPE) and the *Ministério da Ciência, Tecnologia e Inovação* (MCTI) for research grants. F.S. Gomes would like to thank FACEPE for doctoral scholarship; E.V. Pontual acknowledges CAPES and FACEPE for post-doctoral fellowship. Authors are grateful to Dr. Rosiely Felix Bezerra (Figure 1); also to Prof. Dr. Elineide Barbosa de Souza, from the Laboratório de Fitobacteriologia, Universidade Federal Rural de Pernambuco, Recife, for some pictures of plant diseases.

COMPETING INTEREST

Authors have declared that no competing interests exist.

REFERENCES

- [1] Philippot L, Andersson SG, Battin TJ, Prosser JI, Schimel JP, Whitman WB, et al. The ecological coherence of high bacterial taxonomic ranks. Nat Rev Microbiol. 2010; 8(7): 523-529.
- [2] Hynes HM, Germida JJ. Impact of clear cutting on soil microbial communities and bioavailable nutrients in the LFH and Ae horizons of Boreal Plain forest soils. Forest Ecol Manag. 2013; 306: 88-95.
- [3] Babalola OO. Beneficial bacteria of agricultural importance. Biotechnol lett. 2010; 32(11): 1559-1570.

[4] Trabulsi LR, Alterthum F. Microbiologia, 4th ed. São Paulo: Atheneu; 2004. Portuguese.

[5] Siddiqui ZA, Nesha R, Singh N, Alam S. Interactions of Plant-Parasitic Nematodes and Plant-Pathogenic Bacteria. In: Maheshwari DK editor. Bacteria in Agrobiology: Plant Probiotics. Springer Berlin Heidelberg. 251-267; 2012.

[6] Rogers GB, Hoffman LR, Carroll MP, Bruce KD. Interpreting infective microbiota: the importance of an ecological perspective. Trends Microbiol. 2013; 21(6): 271-276.

[7] El-Baky RMA, Ahmed HR, Gad GFM. Prevalence and conjugal transfer of vancomycin resistance among clinical isolates of *Staphylococcus aureus*. Adv Res. 2014; 2(1): 12-23

[8] Yeaman, M R and Yount, N Y. Mechanisms of antimicrobial peptide action and resistance. Pharmacol Rev. 2003; 55(1): 27-55.

[9] Deyn GB, Quirk H, Bardgett RD. Plant species richness, identity and productivity differentially influence key groups of microbes in grassland soils of contrasting fertility. Biology lett. 2011; 7(1): 75-78.

[10] Smith FE. The Treatment of Metalliferous Industrial Effluents in Japan. In Proceedings of Sudbury, 95: 915-924; 1995.

[11] He S, Guo Z, Zhang Y, Zhang S, Wang J, Gu N. Biosynthesis of gold nanoparticles using the bacteria *Rhodopseudomonas capsulata*. Mater Lett. 2007; 61(18): 3984-3987.

[12] Linford N, Linford P, Platzman E. Dating environmental change using magnetic bacteria in archaeological soils from the upper Thames Valley, UK. J Archaeol Sci. 2005; 32(7): 1037-1043.

[13] Daly MJ. Engineering radiation-resistant bacteria for environmental biotechnology. Curr Opin Biotech. 2000; 11(3): 280-285.

[14] Fredrickson JK, Gorby YA. Environmental processes mediated by iron-reducing bacteria. Curr Opin Biotech. 1996; 7(3): 287-294.

[15] Garcia C, Moreno DA, Ballester A, Blazquez ML, Gonzalez F. Bioremediation of an industrial acid mine water by metal-tolerant sulphate-reducing bacteria. Miner Eng. 2001; 14(9): 997-1008.

[16] Lee JH, Mitchell RJ, Kim BC, Cullen DC, Gu MB. A cell array biosensor for environmental toxicity analysis. Biosens Bioelectron. 2005; 21(3): 500-507.

[17] Girotti S, Ferri EN, Fumo MG, Maiolini, E. Monitoring of environmental pollutants by bioluminescent bacteria. Anal Chim Acta. 2008; 608(1): 2-29.

[18] Ding Z, Hao A, Wang Z. Water-in-gasoline microemulsions stabilized by polyglycerol esters. Fuel. 2007; 86(4): 597-602.

[19] Chen X, Xiu Z, Wang J, Zhang D, Xu P. Stoichiometric analysis and experimental investigation of glycerol bioconversion to 1, 3-propanediol by *Klebsiella pneumoniae* under microaerobic conditions. Enzyme Microb Tech. 2003; 33(4): 386-394.

471 [20] Metsoviti M, Paramithiotis S, Drosinos EH, Galiotou-Panayotou M, Nychas GJE, Zeng 472 AP et al. Screening of bacterial strains capable of converting biodiesel-derived raw glycerol 473 into 1, 3-propanediol, 2, 3-butanediol and ethanol. Eng Life Sci. 2012; 12(1): 57-68.

474

482 483

484

488

495

501 502

503

504 505

506

507

511 512

513

514

515

518

- 475 [21] Rossi DM, da Costa JB, de Souza EA, Peralba MDCR, Ayub MAZ. Bioconversion of 476 residual glycerol from biodiesel synthesis into 1, 3-propanediol and ethanol by isolated 477 bacteria from environmental consortia. Renew Energ. 2012; 39(1): 223-227. 478
- [22] Orr CH, James A, Leifert C, Cooper JM, Cummings SP. Diversity and activity of free-living nitrogen-fixing bacteria and total bacteria in organic and conventionally managed soils. Appl Environ Microb. 2011; 77(3): 911-919.
 - [23] Glick BR. Phytoremediation: synergistic use of plants and bacteria to clean up the environment. Biotechnol Adv. 2003; 21(5): 383-393.
- 485 486 [24] Sokolova MG, Akimova GP, Vaishlya OB. Effect of phytohormones synthesized by 487 rhizosphere bacteria on plants. Appl Biochem Microb. 2011; 47(3): 274-278.
- 489 [25] Schalk IJ, Hannauer M, Braud A. New roles for bacterial siderophores in metal transport 490 and tolerance. Environ Microbiol. 2011; 13(11): 2844-2854. 491
- 492 [26] Guo H, Luo S, Chen L, Xiao X, Xi Q, Wei W et al. Bioremediation of heavy metals by growing hyperaccumulaor endophytic bacterium *Bacillus* sp. L14. Bioresource Technol. 494 2010; 101(22): 8599-8605.
- 496 [27] Ma W, Zalec K, Glick BR. Effects of the bioluminescence-labeling of the soil bacterium 497 *Kluyvera ascorbata* SUD165/26. FEMS Microbiol Ecol. 2001; 35: 137–44. 498
- 499 [28] Cho I, Blaser MJ. The human microbiome: at the interface of health and disease. Nat Sou Rev Genet. 2012; 13(4): 260-270.
 - [29] Verschuere L, Rombaut G, Sorgeloos P, Verstraete W. Probiotic bacteria as biological control agents in aquaculture. Microbiol Mol Biol R. 2000; 64(4): 655-671.
 - [30] Dalié DKD, Deschamps AM, Richard-Forget F. Lactic acid bacteria—Potential for control of mould growth and mycotoxins: A review. Food Control. 2010; 21(4): 370-380.
- 508 [31] Trias R, Bañeras L, Montesinos E, Badosa, E. Lactic acid bacteria from fresh fruit and vegetables as biocontrol agents of phytopathogenic bacteria and fungi. Int Microbiol. 2010; 11(4): 231-236.
 - [32] Riquelme, C Araya R, Vergara N, Rojas A, Guaita M, Candia M. Potential probiotic strains in the culture of the Chilean scallop *Argopecten purpuratus* (Lamarck, 1819). Aquaculture. 1997; 154(1): 17-26.
- 516 [33] Rouse S, Harnett D, Vaughan A, van Sinderen D. Lactic acid bacteria with potential to eliminate fungal spoilage in foods. J Appl Microbiol. 2008; 104:915–923.
- 519 [34] Yang L, Maeda H, Yoshikawa T, Zhou GQ. Algicidal effect of bacterial isolates of Pedobacter sp. against cyanobacterium Microcystis aeruginosa. Water Sci Eng. 2012; 5(4): 375-382.

523 [35] Betcher MA, Fung JM, Han AW, O'Connor R, Seronay R, Concepcion GP et al. 524 Microbial distribution and abundance in the digestive system of five shipworm species 525 (Bivalvia: Teredinidae). PLoS One. 2012; 7 (9): e45309

526

530

533

537

540

545 546

547

548

549

553 554

555

556

557 558

559

560

561 562

563

564

565 566

567

568

569

- 527 [36] Prieur D, Mevel G, Nicolas JL, Plusquellec A, Vigneulle M. Interactions between bivalve 528 molluscs and bacteria in the marine environment. Oceanogr. Mar. Biol. Annu. Rev. 1990; 28: 529 277-352.
- [37] Belkin S, Wirsen CO, Jannasch HW. A new sulfur-reducing, extremely thermophilic eubacterium from a submarine thermal vent. Appl Environ Microb. 1986; 51(6): 1180-1185.
- [38] Alfaro AC, Young T, Ganesan AM. Regulatory effects of mussel (*Aulacomya maoriana* Iredale 1915) larval settlement by neuroactive compounds, amino acids and bacterial biofilms. Aquaculture. 2011; 322: 158-168.
- 538 [39] Nadarasah G, Stavrinides J. Insects as alternative hosts for phytopathogenic bacteria. 539 FEMS Microbiol Rev. 2011; 35(3): 555-575.
- [40] N'Guessan CA, Brisse S, Roux-Nio ACL, Poussier S, Koné D, Wicker, E. Development of variable number of tandem repeats typing schemes for *Ralstonia solanacearum*, the agent of bacterial wilt, banana Moko disease and potato brown rot. J Microbiol Meth. 2013; 92: 366-374.
 - [41] Rachmaninov O, Zinger-Yosovich KD, Gilboa-Garber N. Preventing *Ralstonia solanacearum* adhesion with glycans from cashew, cocoa, coffee, pumpkin, and tomato seed extract. Can J Microbiol. 2012; 58(7): 856-862.
- 550 [42] Nascimento AR, Michereff SJ, Mariano RDL, Gomes, AM. Elaboração e validação de 551 escala diagramática para cancro bacteriano da videira. Sum Phytopathol. 2005; 31(1): 59-552 64. Portuguese
 - [43] Czajkowski R, Grabe GJ, Van der Wolf JM. Distribution of Dickeya spp. and Pectobacterium carotovorum subsp. carotovorum in naturally infected seed potatoes. Eur J Plant Pathol. 2009; 125(2): 263-275.
 - [44] Walcott RR, Gitaitis, RD. Detection of Acidovorax avenae subsp. citrulli in watermelon seed using immunomagnetic separation and the polymerase chain reaction. Plant Dis, 2000; 84(4): 470-474.
 - [45] Walcott RR, Fessehaie A, Castro AC. Differences in pathogenicity between two genetically distinct groups of *Acidovorax avenae* subsp. *citrulli* on cucurbit hosts. J Phytopathol. 2004; 152(5): 277-285.
 - [46] Gaidamashvili M, Van Staden J. Interaction of lectin-like proteins of South African medicinal plants with *Staphylococcus aureus* and *Bacillus subtilis*. J Ethnopharmacol. 2002; 80(2): 131-135.
- [47] Ratanapo S, Ngamjunyaporn W, Chulavatnatol M. Interaction of a mulberry leaf lectin with a phytopathogenic bacterium, *P. syringae pv mori*. Plant Sci. 2001; 160(4): 739-744.
- 573 [48] Tasumi S, Yang WJ, Usami T, Tsutsui S, Ohira T, Kawazoe I et al. Characteristics and primary structure of a galectin in the skin mucus of the Japanese eel *Anguilla japonica*. Dev Comp Immunol. 2004; *28*(4): 325-335.

[49] Paiva PMG, Pontual EV, Napoleão TH, Coelho LCBB. Effects of plant lectins and trypsin inhibitors on development, morphology and biochemistry of insect larvae. In: Pourali, H.; Raad, V.N (Eds.). Larvae: Morphology, Biology and Life Cycle. Nova Science Publishers, Inc., New York, 37–55; 2012.

[50] Gomes FS, Procópio TF, Napoleão TH, Coelho LCBB, Paiva PMG. Antimicrobial lectin from *Schinus terebinthifolius* leaf. J Appl Microbiol. 2013; 114(3): 672-679.

[51] Oliveira MDL, Andrade CAS, Santos-Magalhães NS, Coelho LCBB, Teixeira JA, Carneiro-da-Cunha MG et al. Purification of a lectin from *Eugenia uniflora* L. seeds and its potential antibacterial activity. Lett Appl Microbiol. 2008; 46(3): 371-376.

[52] Sá RA, Gomes FS, Napoleão TH, Santos ND, Melo CM, Gusmão NB et al. Antibacterial and antifungal activities of *Myracrodruon urundeuva* heartwood. Wood Sci Technol 2009; *43*(1-2): 85-95.

[53] Costa RMPB, Vaz AFM, Oliva MLV, Coelho LCBB, Correia MTS, Carneiro-da-Cunha MG. A new mistletoe *Phthirusa pyrifolia* leaf lectin with antimicrobial properties. Process Biochem. 2010; 45(4): 526-533.

[54] Ferreira RS, Napoleão TH, Santos AF, Sá RA, Carneiro-da-Cunha MG, Morais MMC et al. Coagulant and antibacterial activities of the water-soluble seed lectin from *Moringa oleifera*. Lett Appl Microbiol. 2011; 53(2): 186-192.

[55] Nunes EDS, de Souza MAA, Vaz AFM, Santana GMS, Gomes FS, Coelho LCBB et al. Purification of a lectin with antibacterial activity from *Bothrops leucurus* snake venom. Comp Biochem Phys B. 2011; 159(1): 57-63.

[56] Moura MC, Pontual EV, Paiva PMG, Coelho LCBB. An Outline to Corrosive Bacteria. In: Microbial pathogens and strategies for combating them: science, technology and education. Formatex Research Centre, Badajos. 2013: (in press).

[57] Hasheesh WS, Mohamed RT. Bioassay of two pesticides on *Bulinus truncatus* snails with emphasis on some biological and histological parameters. Pestic Biochem Phys. 2011; 100(1): 1-6.

[58] Fu JM, Mai BX, Sheng GY, Zhang G, Wang XM, Peng P et al. Persistent organic pollutants in environment of the Pearl River Delta, China: an overview. Chemosphere. 2003; 52: 1411–1422.

[59] Sharma RR, Singh D, Singh R. Biological control of postharvest diseases of fruits and vegetables by microbial antagonists: A review. Biol control. 2009; 50(3): 205-221.

[60] Yoshiyama M, Kimura K. Bacteria in the gut of Japanese honeybee, *Apis cerana japonica*, and their antagonistic effect against *Paenibacillus larvae*, the causal agent of American foulbrood. J Invertebr Pathol. 2009; 102(2): 91-96.

[61] Khan MQ, Abbasi MW, Zaki MJ, Khan SA. Evaluation of *Bacillus thuringiensis* isolates against root-knot nematodes following seed application in okra and mungbean. Pak. J. Bot. 2010; 42(4): 2903-2910.

- [62] Khedher SB, Zouari N, Messaddeq N, Schultz P, Jaoua S. Overproduction of deltaendotoxins by sporeless *Bacillus thuringiensis mutants* obtained by nitrous acid mutagenesis. Curr Microbiol. 2011; 62(1): 38-43.
 - [63] Stevens MM, Hughes PA, Mo J. Evaluation of a commercial *Bacillus thuringiensis* var. *israelensis* formulation for the control of chironomid midge larvae (Diptera: Chironomidae) in establishing rice crops in south-eastern Australia. J Invertebr Pathol. 2013; 112: 9–15
 - [64] Araújo AP, de Melo-Santos MAV, Carlos SDO, Rios EMMM, Regis L. Evaluation of an experimental product based on *Bacillus thuringiensis* sorovar. *Israelensis* against *Aedes aegypti* larvae (Diptera: Culicidae). Biol Control, 2007; 41(3): 339-347.
 - [65] Gill SS, Cowles EA, Pietrantonio PV. The mode of action of *Bacillus thuringiensis* endotoxins. Annu Rev Entomol, 1992; 37(1): 615-634.
- 643 [66] Burton SL, Ellar DJ, Li J, Derbyshire DJ. *N*-acetylgalactosamine on the putative insect 644 receptor aminopeptidase N is recognised by a site on the domain III lectin-like fold of a *Bacillus thuringiensis* insecticidal toxin. J Mol Biol. 1999; 287(5): 1011-1022.
- [67] Rashad FM, Saleh WD, Nasr M, Fathy HM. Identification of mosquito larvicidal bacterial strains isolated from north Sinai in Egypt. AMB Express. 2012; 2(1): 1-15.