

The simultaneous determination of some water-soluble vitamins in gum of *Acacia nilotica* by high performance liquid chromatography

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**Abstract**

A rapid, simple and precise method by HPLC (high performance liquid chromatography) has been developed for simultaneous determination of water-soluble vitamins as thiamine(B<sub>1</sub>), nicotinamide(B<sub>3</sub>), panthotenic(B<sub>5</sub>), pyridoxine(B<sub>6</sub>) and biotin(B<sub>8</sub>) in gum of *Acacia nilotica* using enzymatic hydrolysis. The method uses a C<sub>18</sub> column (4.6×150 mm, 5μm). Mobile phase such as methanol 0.1M, sodium dihydrogen phosphate (pH = 2.5), (10:90 v/v) is found most suitable for rapid separation and identification of this water – soluble vitamins. Good linearity was observed between the concentration of analytes and peak area (r = 0.9999). Each vitamin was quantitatively determined at its maximum wavelength. Recovery percentages ranged from 97% to 99%.

*Keywords: Water – soluble vitamins; Gum, Acacia nilotica; HPLC.*

**1. Introduction**

Acacia gums have a complex and branched structure, which makes them have good adhesive and cohesive properties. These properties are useful in pharmaceutical preparations. They are used as dental and other adhesives and as bulk laxatives. These hydrophilic polymers are useful as tablet binders, emulsifiers, suspending agents, gelling agents, stabilizers, thickeners, protective colloids and suspending agents keeping tablets [1]. They can also be

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used as tablet disintegrants [2]. Their adhesive property could be used in the apparatus of colostomies and also in fixing dental prosthesis [3].

For internal use, they help in the preparation of medicines to soothe coughs, diarrhea, dysentery and hemorrhages; for external use, they calm inflammations, so the presence of vitamins in *Acacia* gums is very important since vitamins are essential for human health. [4]

As far as we know, other researchers have not reported the presence or absence of vitamins B<sub>1</sub>, B<sub>3</sub>, B<sub>5</sub>, B<sub>6</sub>, B<sub>8</sub> in *Acacia* gum, particularly that of *Acacia nilotica*.

These vitamins are very important for the production of energy (B<sub>1</sub>), normal growth and development (B<sub>3</sub>), the regulation of neurotransmitters (messengers of nerve impulses) (B<sub>5</sub>), physical balance and regulation of blood sugar (B<sub>6</sub>) and the processing of several products such as glucose and fatty acids (B<sub>8</sub>). [5]

Due to the nutritional importance of these vitamins, microbiological assay and several analytical methodologies have been developed for the determination of these substances in food, pharmaceutical supplements and biological fluids [6-10]. There are many analytical methods for performing the assay of vitamins in food, pharmaceutical and physiological specimens such as spectrophotometry [6,11-13] spectrophotofluorimetry [7], voltammetry [8], the gas chromatography [15-17] and high performance liquid chromatography [18-28].

Normally, it is necessary to determine more than one vitamin; the analytical method must be able to determine multiple components in complex samples, which can lead to interference in chemical analysis.

The aim of this study is to develop a rapid and reliable technique for the simultaneous determination of five water- soluble vitamins (B<sub>1</sub>, B<sub>3</sub>, B<sub>5</sub>, B<sub>6</sub>, and B<sub>8</sub>) in gum of *Acacia nilotica* by HPLC using the enzymatic hydrolysis.

## 58 2. Experimental

### 59 2.1. Reagents and chemicals

60 Methanol was of HPLC grade. Other chemicals as sodium dihydrogen  
61 phosphate ( $\text{NaH}_2\text{PO}_4$ ), sodium acetate, sulfuric acid ( $\text{H}_2\text{SO}_4$ ) and acetic acid  
62 ( $\text{C}_2\text{H}_4\text{O}_2$ ) (Sigma) were of reagent grade. Purified water was obtained from a  
63 Millipore Milli-Q system.

64 Standards of thiamine, nicotinamide, pantothenic, pyridoxine and biotin were  
65 purchased from Sigma.

66 Taka-diastase enzyme from *Aspergillus oryzae* powder, slightly beige was  
67 obtained from Sigma. All chemicals and reagents used are of HPLC and were  
68 used without further purification. Also, all solutions were filtered through a  
69  $0.45\ \mu\text{m}$  membrane (Millipore), protected from light and stored at  $4^\circ\text{C}$ .

70 The mobile phase of the HPLC system was comprised of pure methanol and  
71 sodium dihydrogenphosphate  $\text{NaH}_2\text{PO}_4$  (10:90 v/v).

### 72 2.2. Chromatographic conditions

73 The HPLC system (Agilent) was equipped with a pump type technology  
74 Agilent 1200 series, a vacuum degassing unit model G1322A, a UV-VIS  
75 spectrometer to 8 wavelengths, a fluorescence detector (G1321 Agilent 1200  
76 Series), an analytical  $\text{C}_{18}$  column (Agilent) ( $4.6 \times 150\text{mm}$ ,  $5\mu\text{m}$ ). During the  
77 analysis the column was equilibrated at  $30^\circ\text{C}$  and a manual injector uses an  
78 injection valve sample seven lane Rheodyne 7725i. The chromatographic peaks  
79 were recorded and elaborated automatically by employing a computerized  
80 program 'Agilent ChemStation'.

81 The analyzes were performed by gradient elution of wavelength at room  
82 temperature, at a flow rate of  $1\ \text{mL} / \text{min}$ . The total execution time required is  
83 less than 20 min.

84 The program of wavelength changes during elution time for five vitamins  
85 determination in gum of *Acacia nilotica* shown in Table2.

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### 2.3. Standard solutions

The vitamin stock solution: 100 mg / L were prepared by dissolving 10 mg of each standard in 100 mL of methanol in dark volumetric flasks. These solutions are stable hang at least one month when stored in the dark at 4°C. Working solutions were prepared from stock solutions by appropriate dilution with methanol and protected from light. The following Table1 illustrates the calibration of the analytical method:

Table1. Concentration of the standards used for plotting the calibration curve of five vitamins (B<sub>1</sub>, B<sub>3</sub>, B<sub>5</sub>, B<sub>6</sub>, B<sub>8</sub>)

Vitamins	Concentrations (mg/L)
Thiamine B <sub>1</sub>	1.0, 5.0, 10.0, 15.0, 20.0, 30.0
Nicotinamide B <sub>3</sub>	1.0, 5.0, 10.0, 15.0, 20.0, 30.0
Pantothenic B <sub>5</sub>	1.0, 5.0, 10.0, 15.0, 20.0, 30.0
Pyridoxine B <sub>6</sub>	1.0, 5.0, 10.0, 15.0, 20.0, 30.0
Biotin B <sub>8</sub>	1.0, 2.0 ,5.0 ,10.0 ,15.0, 20.0, 30.0

### 2.4 Sample preparation

1g of gum *Acacia nilotica* which is a fine powder was accurately weighed in a 250 mL erlenmeyer flask, 10mL of sulfuric acid (1N) was added. The mixture was thoroughly shaken, after the pH was adjusted to 4.5 with sodium acetate (2.5M), then 500mg of the enzyme Taka-diastrase stirring was added .The solution was incubated at 37°C and protected from light all night. The following content was filtered on 0.45µm filter. Finally, 20 µL of the extract was injected into the HPLC system for analysis.

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### 3. Results and discussion

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The determination of vitamins in gum of *Acacia nilotica* is a complex analytical problem for several reasons: gum of *Acacia nilotica* is a very complex matrix, vitamins that are micro constituents and vitamins are easily destroyed by strong acids or alkalis, which is why we find that the enzymatic hydrolysis is a good solution for these problems.

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First, scan analysis of standard vitamins was performed to check the optimum conditions for the detection. Wavelengths were changed according to the elution time of each vitamin, as is shown in Table 2.

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Table 2. Program of wavelength changes during elution time for five water-soluble determinations in gum *Acacia nilotica*.

Vitamins	Time (min)	Wavelengths (nm)
Biotin (B <sub>8</sub> )	0.0 - 2.5	204
Nicotinamide (B <sub>3</sub> )	2.6 - 3.8	261
Thiamine (B <sub>1</sub> )	3.9 - 4.5	234
Pyridoxine (B <sub>6</sub> )	4.6 - 5.0	275
Pantothenic acid (B <sub>5</sub> )	5.1 - 7.0	210

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The mobile phase was composed of methanol and sodium dihydrogen phosphate NaH<sub>2</sub>PO<sub>4</sub> (10:90) v/v) for the determination of vitamins B<sub>1</sub>, B<sub>3</sub>, B<sub>5</sub>, B<sub>6</sub>, and B<sub>8</sub> in gum *Acacia nilotica*. A study of pH and the proportion of methanol and NaH<sub>2</sub>PO<sub>4</sub> were necessary to improve the resolution in the gum of *Acacia nilotica* formulae. When the proportion of methanol is 20%, vitamins are eluted in less than 5 min, but there is an overlap peak of certain vitamins.

123 The pH of the mobile phase is extremely important for the separation of  
124 vitamins in order to overcome this problem, a decrease in the proportion of  
125 methanol by 10 %, which has the effect of providing a higher resolution but  
126 against party, the time of analysis.

127 A choice of pH = 2.5 implies that most vitamins are of molecular form since  
128 the pH is less than the peaks of all vitamins (B<sub>1</sub>, B<sub>3</sub>, B<sub>5</sub>, B<sub>6</sub>, B<sub>8</sub>). Figure 1 shows  
129 the chromatogram of vitamins B<sub>1</sub>, B<sub>3</sub>, B<sub>8</sub> in gum of *Acacia nilotica*.

130 We note from the figure that gum of *Acacia nilotica* contains a wide range of  
131 vitamin B<sub>8</sub>. The peak of vitamin B<sub>8</sub> was detected at a retention time of about  
132 2.2 to 2.4 min (Fig.1, 2), with minor variations on a daily basis due to  
133 temperature fluctuations in the laboratory [29]. No other peaks were observed  
134 at 204 nm.

135 All calculations prove that vitamin B<sub>8</sub> is in the order of 12,000 ppm. A part  
136 from vitamin B<sub>8</sub>, there are vitamins B<sub>1</sub> and B<sub>3</sub> but with low levels. We can  
137 conclude that the method gives a good resolution of vitamin B<sub>8</sub>.

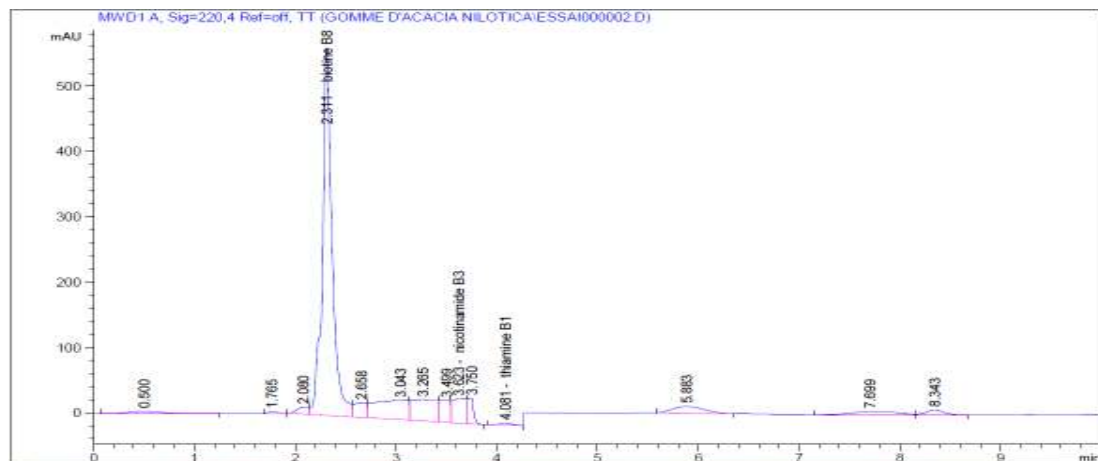
### 138 **3.1. Characteristic of the HPLC method**

139 The proposed method allows the resolution of various forms of vitamin B  
140 especially B<sub>8</sub> in gum of *Acacia nilotica* by HPLC with UV detection.

141 A reliable chromatographic assay requires an acceptable resolution, reasonable  
142 retention times and good peak symmetry. Accordingly, in preliminary studies  
143 optimal chromatographic conditions were investigated in gradient elution  
144 system with varying wavelengths. The advantage of gradient elution is that the  
145 bandwidth can be nearly constant at both early and tardative analytes.  
146 Therefore an elution system of five wavelengths has been developed with a  
147 beneficial effect on the sensitivity of biotin.

148 Representative chromatograms with other chromatographic parameters are  
149 shown in Figure 1, 2 and Table 3.

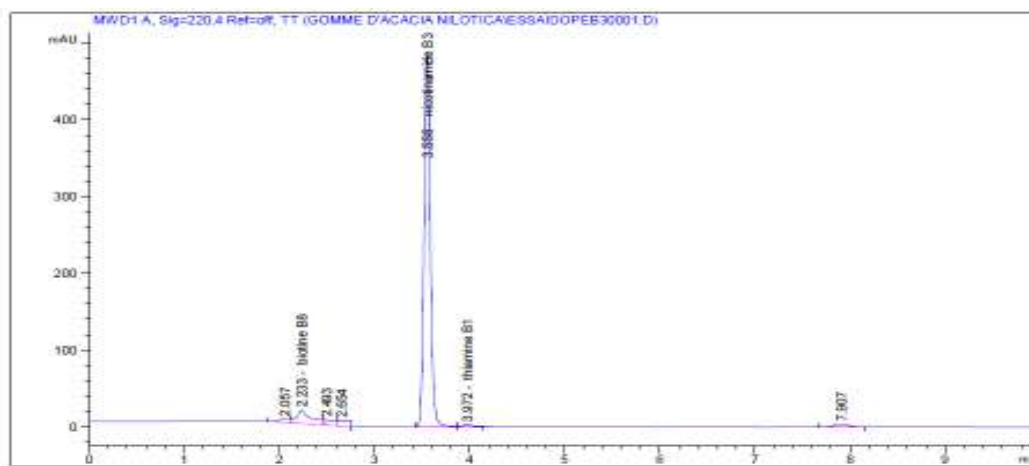
Amount (mg/L)



Retention time (min)

Figure 1: Typical chromatograms of vitamins.

Amount (mg/L)



Retention time (min)

Figure 2: Typical chromatograms of vitamins.

Table 3. High performance liquid chromatographic parameters of the *Acacia gum* separation using the gradient elution system  $t_0 = t_m = 0.56$  min.

Vitamins	Chromatographic parameters			
	T	$t_R$ (min)	$R_S$	$k'$
Biotin B <sub>8</sub>	2.306	3.117	1.0	2.412 (B3-B8)
Nicotinamide B <sub>3</sub>	3.512	5.27	1.5	2.087 (B1-B3)
Thiamine B <sub>1</sub>	4.242	6.575	1.9	3.872 (B1-B8)

$t_R$ : retention time reduced;  $k'$ : Retention factor;  $R_S$ : Resolution factor; T: The asymmetric peak

In practice, care must be taken to keep values  $k'$  inferior to 10 for a period of reasonable analysis, values between 2 and 5 are the correct values.

### 3.1.1 Linearity

Six working solutions were prepared for each analyte whose range is between 1 and 30 mg /L for B<sub>1</sub>, B<sub>3</sub>, B<sub>5</sub>, B<sub>6</sub> and seven solutions between 1 and 30 mg/L for B<sub>8</sub>. The analysis was performed in triplicate to determine the linearity of the assay. The regression lines were calculated by the method of least squares of the areas of the peaks relative to the analyte.

The equations corresponding to the five regression analytes were

$$B_1: y = 25.82754x + 6.26753$$

$$B_3: y = 33.21959x - 1.42661$$

$$B_5: y = 7.15590x + 5.776 \cdot 10^{-1}$$

$$B_6: y = 13.70389x + 9.16444 \cdot 10^{-1}$$

$$B_8: y = 11.89793x + 2.37982$$

x: Amount et y: Area

They were consistently linear in the already mentioned range for all compounds.



182 The linearity was checked by analysis of variance of the regression (Table 4).  
 183 A value of  $r$  above 0.9949 for all vitamins, ( $P < 0.001$ ) except for thiamine with  
 184  $r = 0.9781$ . The coefficient of determination ( $r^2$ ) is more than 95.66% for  
 185 thiamine and 99.66% higher than for others. Six determinations of the same  
 186 sample were performed to assess the accuracy of the method.

187 Table 4. Linearity of standard curves of vitamins B<sub>1</sub>, B<sub>3</sub>, B<sub>5</sub>, B<sub>6</sub>, and B<sub>8</sub>

Vitamins	$r$	$r^2$	$F_{\text{exp}}^a$	$DF^b$	$P$
Biotin (B <sub>8</sub> )	0.9949	99.88	21247.5	1.5	( $P < 0.001$ )
Nicotinamide(B <sub>3</sub> )	0.9998	99.96	165492.10	1.5	( $P < 0.001$ )
Thiamine (B <sub>1</sub> )	0.9781	95.66	100026.5	1.5	( $P < 0.001$ )
Pyridoxine (B <sub>6</sub> )	0.9998	99.96	27417.53	1.5	( $P < 0.001$ )
Pantothénic acid (B <sub>5</sub> )	0.9999	99.98	7686.97	1.5	( $P < 0.001$ )

188  $F^a (1.5; 0.001) = 6.61$ .  $F_{\text{tab}}$  and  $F_{\text{exp}}$  are tabulated and experimental  
 189 Snedecor's  $F$  values, respectively in ANOVA analysis,  $DF^b$ , degrees of freedom

### 190 3.1.2 Accuracy and precision

191 Six determinations of the same sample were performed to assess the accuracy  
 192 of the method.

193 The following table illustrates the accuracy of the method for the determination  
 194 of vitamins B<sub>1</sub>, B<sub>3</sub>, B<sub>5</sub>, B<sub>6</sub>, and B<sub>8</sub> in gum of *Acacia nilotica*.

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199 Table 5. Peak area range and concentration, correlation data of the calibration curves  
 200 and quantification limit of determined vitamins in gum of *Acacia nilotica*.

Vitamins	Concentration range (mg/L)	Surface broad peak	Correlation coefficient	Detection limits(mg/L)	Quantification limits (mg/L)
Biotin (B <sub>8</sub> )	1-30	59.83426- 348.10776	0.9949	0.006	0.022
Nicotinamide (B <sub>3</sub> )	1-30	31.45451- 1000.048	0.9998	0.008	0.028
Thiamine (B <sub>1</sub> )	1-30	18.14213- 714.6253	0.9781	0.012	0.042
Pyrodixine (B <sub>6</sub> )	1-30	14.31208- 409.0012	0.9998	0.002	0.007
Pantotenic acid (B <sub>5</sub> )	1-30	8.05888- 214.6708	0.9999	0.001	0.0035

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### 202 3.1.3 Recovery

203 The recovery rate was tested by the standard addition procedure. One level was  
 204 used for each water-soluble vitamin in gum samples (Table 6).Mean recoveries  
 205 obtained were always satisfactory-higher than 99% for biotin, higher than  
 206 nicotinamide 98.8 %,and higher than thiamine.

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Table 6. Study of determining recovery rate by the addition of 500μL of vitamins B<sub>1</sub>, B<sub>3</sub> and B<sub>8</sub>

N <sub>0</sub> test	Biotin			Nicotinamide			Thiamine		
	Found	Recovery	%	Found	Recovery	%	Found	Recovery	%
	value	%	RSD	value	%	RSD	value	%	RSD
	(mg/L)			(mg/L)			(mg/L)		
1	1994.7	99.0	0.23	5.04	98.8	0.91	2.81e <sup>-1</sup>	97.2	1.26

#### 4. Conclusion

In this work, we optimized HPLC conditions for determination of the water – soluble vitamins such as thiamine (B<sub>1</sub>), nicotinamide (B<sub>3</sub>), pantothenic acid (B<sub>5</sub>), pyridoxine (B<sub>6</sub>), biotin (B<sub>8</sub>) in gum of *Acacia nilotica* following sample preparation by enzymatic hydrolys. The chromatographic separation was performed on a C<sub>18</sub> reverse phase, and vitamins are detected at different wavelengths by UV-visible. This method is rapid, simple, and reliable and saves a significant amount of reagent.

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