

Original Research Article

Effect of Sinusoidal Excitation on Fluid Flow across a Cu-Mica Microchannel

ABSTRACT

Microfluidic devices integrated with on-chip control circuitry have been widely used in various biological and chemical synthesis applications. The objective of this paper is to investigate the effect of gravity, temperature, pulse width vibrations (PWM), and sinusoidal excitations on the flow of methanol, ethanol, and chloroform through an indigenously fabricated Cu-Mica microchannel for automatic identification of fluids. The results of the investigations showed that the rate of flow of microfluids increases with an increase in the angle of elevation. Chloroform shows maximum speed and the acceleration is maximum around the elevation angles 60° - 70° . Ethanol shows minimum speed. Both ethanol and methanol show maximum acceleration around 80° - 90° of elevation angles. The velocity of the fluids is also a function of temperature. Chloroform shows maximum velocity at 50°C and minimum at the room temperature. The acceleration is fluid dependent.

For PWM vibrations, chloroform takes comparatively lesser time to flow across the given microchannel. The velocity of the fluids is not a monotonic function of the PWM frequency. For sinusoidal excitations, ethanol exhibits maximum velocity around the frequency 1.5 KHz. The minimum velocity is shown at 4.5 KHz. For methanol, maximum velocity observed is around 2.5 KHz and minimum at 3.5 KHz. Chloroform shows no visible effect of excitation in its flow velocity. As velocity profile for a given set of influencing factors is fluid dependent, microchannel based sensors may be developed for automatic identification of liquids.

Keywords: Microchannel, microfluids, PWM vibration, sinusoidal Excitation, velocity profile.

1. INTRODUCTION

Microfluidics posses broad range of applications in different fields due to low cost integration with on-chip systems, low power utilization, and higher sensitivity [1]. The electrical interrogation of micro-devices has led to the extensive exploration in usage of integrated on-chip systems, especially, microfluidics. Microelectronics devices integrated with microfluidics has various sensing applications like fluid sensor, flow control in liquids [2] [3]. These sensors have a number of applications in flow cytometry [4], wind estimations [5], gas chromatography [6], gas monitoring [7], wall-shear stress [8], and viscosity measurements [9]. These techniques include microfluidic device development such as valves, pumps, and microfluidic channels that result into lab-on chip microfluidics devices or micro machining analysis systems while integrated together [10-12].

Microfluidic devices integrated with on-chip control circuitry may be used in various biological or chemical synthesis applications for manipulating fluidic flow and precisely controlling its motion [13]. The miniaturized microchannels has been also used for detection [14] [15], purification, fractionating [16-18], single cell sorting of DNA [14-19], and linear analysis of stretched DNA molecules [20] [21]. The selection criteria for choice of materials and suitable technique to fabricate microfluidics device depends on certain parameters such as capillary effects, surface to volume ratio, electro-osmotic flows, geometrical cross section of the channel [21]. Various fabrication techniques such as sacrificial layer etching, e-beam lithography, nano imprint lithography [22-25], photolithography, laser ablation, hot embossing, CVD set up, and micromachining [26] has already been employed for microchannel fabrication by various researchers.

The objective of this paper is to investigate the effect of gravity, temperature, PWM vibrations, and sinusoidal excitations on the flow of selective fluids, such as, methanol, ethanol, and chloroform through

an indigenously fabricated Cu-Mica microchannel for automatic identification of fluids. This work also explains the basics of the factors responsible for the flow of microfluids through microchannels. The details of pattern etching and basics of flow injection analysis are discussed in the following section. The methodology is presented in Section 3 followed by the results and conclusion in the subsequent sections.

2. MICROFLUIDIC FLOW PARAMETERS

The various factors responsible for the flow of microfluids are briefed as follows.

2.1 Reynolds number

This is a dimensionless parameter and used to determine the type of flow pattern. The value of Reynolds number depends upon the geometry of the channel and is given by

$$R_e = \frac{LV_{avg}\rho}{\mu}$$

where $h = \frac{2\gamma\cos\theta}{gr\rho}$ is the channel length, A the crosssectional area of the channel, P the wetted perimeter of the channel, μ the viscosity, ρ the fluid density, and V_{avg} the average velocity of flow [27]. R_e depends on material properties (density, viscosity), boundary conditions, and critical velocity. Reynolds number less than 25 is common in microfluidics. The flow may be laminar flow, turbulent, or transient depending upon the value of Reynolds number as shown in Table 1.

TABLE 1: Nature of fluid flow on basis of Reynolds number

Range of R_e	Nature of fluid flow
$R_e < 2300$	Laminar Flow
$2300 < R_e < 4000$	Transient Flow
$R_e > 4000$	Turbulent Flow

Laminar flow is a streamline flow in which smooth sliding of adjacent layers resembling a set of parallel layers takes place. The field of velocity vectors is constant with time. The flow rates are relatively low and the Reynolds number value of such flow is less than 2300 [28]. The laminar flow at higher velocities is referred as turbulent flow. Reynolds number value for such flow is usually above 4000. The curling of field lines takes place leading to mixing of the adjacent layers in such a way that there occurs unpredictable development of the velocity vector field. The flow pattern is observed to be increasingly turbulent towards the higher velocities [27-29]. Transient flow is a periodic flow and termed as third flow regime in micro fluidics. This includes the surface waves and the acoustic waves. The Reynolds number value varies between the 2300 and 4000. Since micro channels are small in dimensions, Reynolds number R_e is much less than 100 and often less than 1 in microfluidics which means the flow of microfluids is laminar without occurrence of any turbulence. However the fluid flow transition from laminar to turbulent can also occur due to its sensitivity to flow disturbances and channel imperfections. The extreme case of laminar flow is the stokes flow which involves the creeping motion of fluid through channels at Reynolds number lesser than 1. This is due to greater effects of viscous forces acting relative to the inertial forces at low Reynolds number values. The microfluidic flow regime includes various types of flow such as a bubbly flow, slug /Taylor flow, churn flow, slug /annular flow and annular flow.

2.2 Viscosity

Viscosity of a fluid is the internal resistance to its flow. The property of posing a friction to the fluid flow is termed as the viscosity. It is necessary to determine the flow speed, since the fluids with low viscosity are faster in flow as compared to highly viscous fluids [30].

2.3 Height of liquid

The amount of material drawn due to the capillary action is termed as height of liquid and is designated as h . Capillary action forms the basis of microfluidics since capillary pressure of a fluid flowing across a microchannel depends upon the capillary action in which the adhesive intermolecular forces at liquid – substrate interface become more stronger than the cohesive intermolecular forces inside the liquid. Thus whenever fluid interacts with the microchannel i.e. capillary interface, the surface tension induced causes the fluid flow advancing across the microchannel [31]. Mathematically, height of liquid is expressed as

$$h = \frac{2\gamma \cos \theta}{gr\rho}$$

where γ represents surface tension of fluid, θ is contact angle, r is column radius, g denotes the gravitational force, and ρ is the density [32-36]. Fluid flow in capillaries of crosssectional dimensions above 1mm forms a new fluidic system called milli-fluidic system. In milli-fluidic system the transition to turbulence when Reynolds number value reaches 1, can never be neglected. This is a limitation of milli-fluidic system as compared to other fluidic systems or even microfluidic systems [31].

3. METHODOLOGY

The methodology for investigating the effect of sinusoidal response of fluidic flow across Cu-Mica microchannels is divided into two steps.

3.1 Microchannel fabrication

Y-shaped microchannels having length 3.6 cm were designed in Coral draw and transferred on a Cu-Mica board using screen printing. The channel width was fixed at 1 mm. Figure 1 shows the Snapshots of Cu-Mica microchannels.

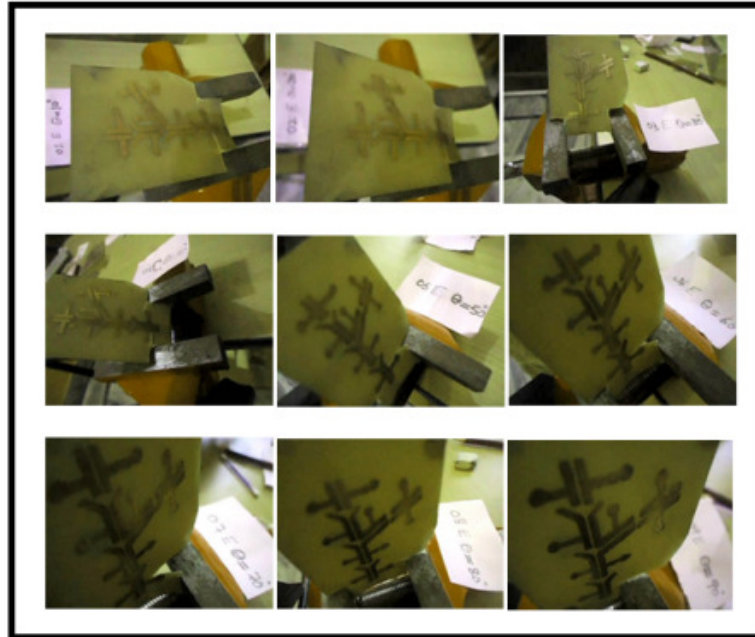


Fig. 1: Snapshots of Cu-Mica microchannel at different elevations.

The channel patterned Cu-Mica board was subjected to etching to obtain microchannels with copper walling. The etchant used for was pale yellow solution of ferric chloride (FeCl_2) and water in the ratio of 3:1. For accelerating the etching few drops of HCl were poured into the solution. The patterns of the microchannels to be fabricated were transferred on a Cu-Mica board coated with swab coating after cleaning with acetone. The patterned Cu-Mica board was immersed in the etchant solution. For enhancing

the rate of etching, the tray containing the etchant was gently moved in left- right direction followed by gentle movements in up/ down directions. The process was continued till the removal of unwanted copper from exposed areas of Cu-Mica board starts appearing. The complete etching took about 15 minutes. After etching, it was washed under running water first and then swab coated using acetone to ensure dirt free channel fabrication.

3.2 Analysis

Four different experiments were carried to investigate the effect of gravity, temperature, PWM vibrations, and sinusoidal excitations on the fluidic flow across Cu-Mica microchannels. During first experiment, the effect of different elevations of the microchannels ranging from 10° to 90° in the steps of 10° on the microfluidic flow has been investigated. Snapshots of different elevations of microchannels varying from 10° to 90° had already been shown in Fig. 1. In the second experiment, the effect of temperature on fluidic flow across Cu-Micamicrochannels has been investigated. For conducting the experiments with the flow of ethanol, methanol and chloroform, the angle of elevation of the microchannels was fixed at 40° . The flow of the liquids at different temperatures ranging from room temperature to 50°C was recorded using a digital movie camera placed in front of the microchannels. The temperature was maintained by using an IR lamp based setup as shown in Fig.2.

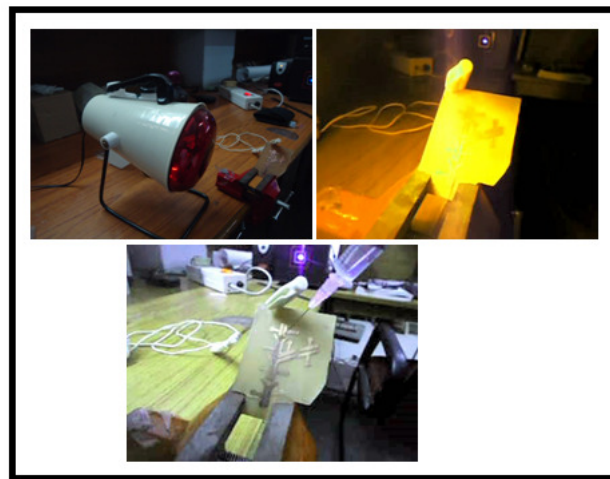


Fig. 2: Temperature control using IR lamp set up.

In the third experiment, the effect of PWM vibrations on fluidic flow across Cu-Mica microchannels has been investigated. In this experiment the angle of elevation of the microchannels was fixed at 25° . The instructions regarding the pulse durations were sent to the microcontroller system using a PC via serial programming interface. The frequency of the pulses was varied from 100 Hz to 5KHz with a step of 5KHz. PWM output of microcontroller was applied to the piezoelectric vibrator through a driver circuit. The vibrations were coupled to the microfluidic system using a rigidly bounded piezoelectric crystal. These steps are shown in Fig. 3.

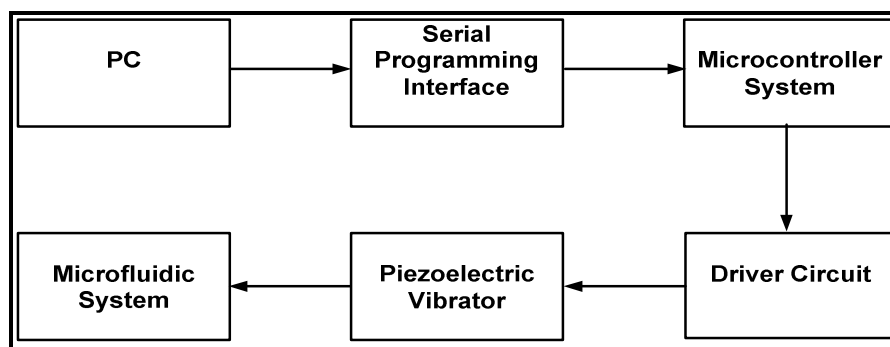


Fig. 3: Block diagram of the set up used for proposed investigations.

In the fourth experiment, the effect of sinusoidal excitations on the fluidic flow across Cu-Mica microchannels has been investigated. The elevation of the microchannels was fixed at 40° using a buncher. The sinusoidal excitations in the range of 100Hz to 5KHz with a step of 500Hz were generated using a PC equipped with high quality sound card. The output of the sound card was applied to an audio amplifier and the amplified signal was coupled to the microchannel by using a speaker as shown in Fig. 4.



Fig. 4: Set-up for evaluating the effect of sinusoidal excitations.

The audio sine wave at 100 Hz frequency was input and corresponding vibrations were produced. Microfluids were injected using metal syringe into microchannel supplied with sine wave vibrations and the flow of fluid was recorded using a digital movie camera.



Fig. 5: Set-up for recording and analysis.

The flow pattern of other two fluids was also recorded in the same way. Then the sine wave generated at 500 Hz was applied as input to produce vibrations and flow pattern of each fluid was recorded at 500 Hz. This was repeated for different frequencies varying from 500 Hz to 5 KHz for each fluid and recorded for further proposed investigations as shown in Fig. 5. For injecting the fluids in the microchannels, micro syringe was used called flow injection analysis (FIA) technique. It is a robust chemical analysis technique used for analysis of continuous fluid flows. FIA is preferred over conventional chemical analysis techniques due to its high reproducibility and sensitivity performance throughput capability that provides precise and fast analytical results with high degree of automation at low costs. It is also advantageous since it requires small amount of reagents for high resolution analysis [33] compared to conventional Flow analysis systems.

4. Results and Discussions

The results of the first experiment showed that speed of the microfluids increases with an increase in the angle of elevation. Chloroform shows maximum speed and the acceleration is maximum around the elevation angles 60° - 70° . Ethanol shows minimum flow speed. Both ethanol and methanol show maximum acceleration around 80° - 90° of elevation angles.

TABLE II: Time and velocity computation of data recorded during analysis.

Frequency (Hz)	Time (s)			Speed (cm/s)		
	Ethanol	Methanol	Chloroform	Ethanol	Methanol	Chloroform
100	7.2	5.7	1.0	0.333	0.421	2.40
500	4.6	3.2	1.0	0.522	0.750	2.40
1000	4.2	4.9	1.0	0.571	0.489	2.40
1500	3.8	5.4	1.0	0.631	0.444	2.40
2000	4.2	3.7	1.0	0.571	0.648	2.40
2500	7.0	3.6	1.0	0.343	0.666	2.40
3000	5.1	10.1	1.0	0.471	0.238	2.40
3500	5.0	13.1	1.0	0.480	0.183	2.40
4000	4.5	12.6	1.0	0.533	0.190	2.40
4500	9.2	11.4	1.0	0.261	0.211	2.40
5000	6.5	7.5	1.0	0.369	0.320	2.40

The results of second experiment show that flow velocity of the liquids is a function of temperature. The chloroform shows maximum velocity at 50°C and minimum at the room temperature. The observations showed that the acceleration is fluid dependent. The results of the third experiment are plotted in Fig. 6 as velocity profile for the three liquids. The plot shows that the chloroform takes comparatively lesser time to flow across the given microchannel. From the figure, it is observed that chloroform takes comparatively lesser time to flow across the given microchannel. In general, it may be expected that the velocity of the fluids is not a monotonic function of the frequency. The velocity profile with respect to frequency shows maxima around 1 kHz and 3 kHz. The height of maxima goes on increasing with the frequency of vibration.

It is interesting to note that there are some frequencies for which ethanol and methanol do not show any movement irrespective of the amplitude of the vibration. Further, the investigations showed that Methanol could not cross the channel even on the application of high frequency vibrations. On the other hand Ethanol started flowing above 2 KHz and the velocity increased with increasing frequency. Chloroform was observed to be the fastest moving fluid and its flow rate increases with increase in frequency. The maximum velocity was observed at 5 KHz and minimum velocity at 100 Hz. The results of the fourth experiment are tabulated in Table II and plotted as velocity profile as a function of frequency

in Fig. 7. From the plots it may be observed that maximum velocity was observed for ethanol around the frequency of the excitation of 1500 Hz. The minimum velocity is shown at 4500 Hz. Some peaks in the velocity are observed around 1 KHz, 1.5 KHz, 2 KHz. For methanol maximum velocity observed was around 2.5 KHz and minimum at 3.5 KHz. The peak velocities are observed around 500 Hz, 2 KHz and 2.5 KHz. Chloroform showed no visible effect of vibration in its velocity profile and remained constant at all frequencies of vibrations.

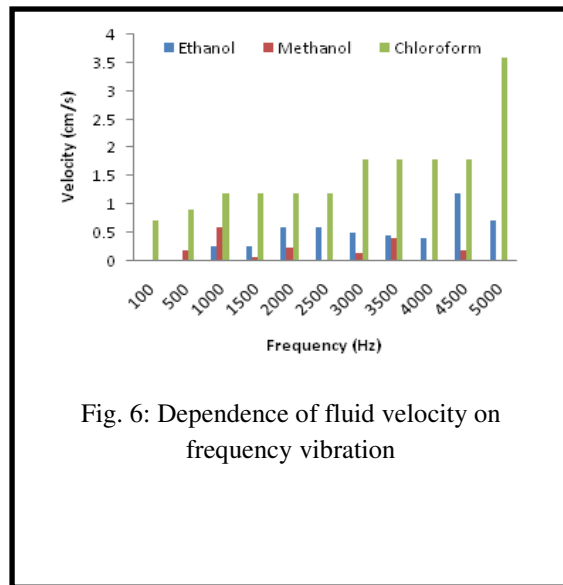


Fig. 6: Dependence of fluid velocity on frequency vibration

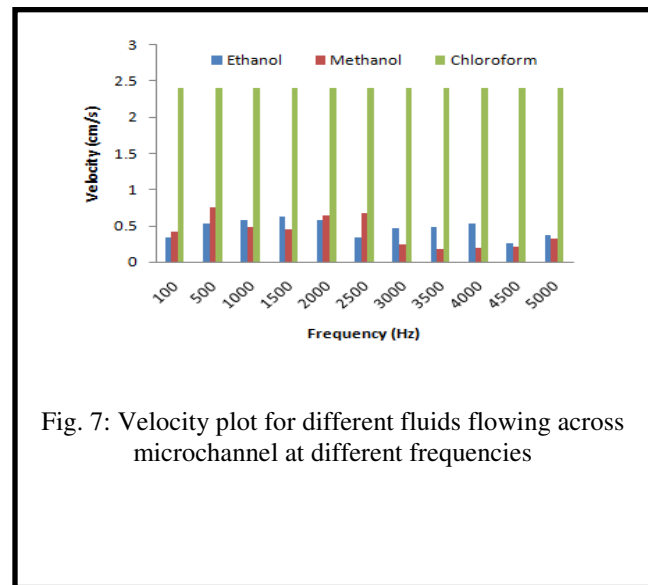


Fig. 7: Velocity plot for different fluids flowing across microchannel at different frequencies

5. CONCLUSION

Investigations were carried out to study the effect of four different factors namely gravity, temperature, PWM vibrations and sinusoidal excitations on fluid flow across fabricated Cu- Mica microchannels. The results of the first experiment showed that speed of the microfluids increases with an increase in the angle of elevation. Chloroform shows maximum speed and the acceleration is maximum around the elevation angles 60° - 70° . Ethanol shows minimum flow speed. Both ethanol and methanol show maximum acceleration around 80° - 90° of elevation angles. The results of second experiment show that flow velocity of the liquids is a function of temperature. The chloroform shows maximum velocity at 50°C and minimum at the room temperature. The observations show that the acceleration is fluid dependent. The results of the third experiment show that the chloroform takes comparatively lesser time to flow across the given microchannel. In general, it may be expected that the velocity of the fluids is not a monotonic function of the frequency. The velocity file with respect to frequency shows maxima around 1 kHz and 3 kHz. The height of maxima goes on increasing with the frequency of vibration. The results of the fourth experiment showed that ethanol exhibits maximum velocity around the frequency of the excitation of 1.5 KHz. The minimum velocity is shown at 4.5 KHz. For methanol maximum velocity observed is around 2.5 KHz and minimum at 3.5 KHz. Chloroform showed no visible effect of vibration in its velocity profile and remained constant at all frequencies of vibrations. In conclusion, microchannel based sensors may be developed for automatic identification of liquids.

References

- [1] J. T. W. Kuo, L. Yu, and E. Meng, "Micromachined Thermal Flow Sensors—A Review," *Micromachines*, vol. 3, no. 4, pp. 550–573, Jul. 2012.

- 239 [2] J. Friend and L. Yeo, "Fabrication of microfluidic devices using polydimethylsiloxane," *Biomicrofluidics*,
240 vol. 4, no. 2, p. 026502, 2010.
- 241 [3] K. Park, H.-J. Suk, D. Akin, and R. Bashir, "Dielectrophoresis-based cell manipulation using electrodes on a
242 reusable printed circuit board," *Lab Chip*, vol. 9, no. 15, p. 2224, 2009.
- 243 [4] W. K. Wu, C. K. Liang, and J. Z. Huang, "MEMS-based flow cytometry: microfluidics-based cell
244 identification system by fluorescent imaging," *The 26th Annual International Conference of the IEEE
245 Engineering in Medicine and Biology Society*.
- 246 [5] K. A. A. Makinwa and J. H. Huijsing, "A wind sensor with an integrated low-offset instrumentation
247 amplifier," *ICECS 2001. 8th IEEE International Conference on Electronics, Circuits and Systems*.
- 248 [6] B. C. Kaanta, H. Chen, G. Lambertus, W. H. Steinecker, O. Zhdanev, and X. Zhang, "High Sensitivity
249 Micro-Thermal Conductivity Detector for Gas Chromatography," *IEEE 22nd International Conference on
250 Micro Electro Mechanical Systems*, 2009.
- 251 [7] Bolin Yu, Zhiyin Gan, Shu Cao, Jingping Xu, and Sheng Liu, "A micro channel integrated gas flow sensor for
252 high sensitivity," *11th Intersociety Conference on Thermal and Thermomechanical Phenomena in Electronic
253 Systems*, 2008.
- 254 [8] L. Löfdahl and M. Gad-el-Hak, "MEMS-based pressure and shear stress sensors for turbulent flows,"
255 *Measurement Science and Technology*, vol. 10, no. 8, pp. 665–686, Aug. 1999.
- 256 [9] P.-Y. Ju, C.-H. Tsai, L.-M. Fu, and C.-H. Lin, "Microfluidic flow meter and viscometer utilizing flow-induced
257 vibration on an optic fiber cantilever," *16th International Solid-State Sensors, Actuators and Microsystems
258 Conference*. 2011.
- 259 [10] A. F. P. van Putten and S. Middelhoeck, "Integrated silicon anemometer," *Electronics Letters*, vol. 10, no. 21,
260 p. 425, 1974.
- 261 [11] N. Nguyen, "Micromachined flow sensors—a review," *Flow Measurement and Instrumentation*, vol. 8, no.
262 1, pp. 7–16, Mar. 1997.
- 263 [12] Y.-H. Wang, C.-P. Chen, and C.-M. Chang, "MEMS based gas flow sensors," *MicrofluidNanofluidics*, vol. 6,
264 pp. 333–346, 2006.
- 265 [13] T. S. Hug, N. F. de Rooij, and U. Staufer, "Fabrication and electroosmotic flow measurements in micro- and
266 nanofluidic channels," *MicrofluidNanofluid*, vol. 2, no. 2, pp. 117–124, Mar. 2006.
- 267 [14] Z. Petrášek, M. Krishnan, I. Mönch, and P. Schwill, "Simultaneous two-photon fluorescence correlation
268 spectroscopy and lifetime imaging of dye molecules in submicrometer fluidic structures," *Microscopy
269 Research and Technique*, vol. 70, no. 5, pp. 459–466, May 2007.
- 270 [15] S. W. Turner, "Monolithic nanofluid sieving structures for DNA manipulation," *J. Vac. Sci. Technol. B*, vol.
271 16, no. 6, p. 3835, Nov. 1998.
- 272 [16] C.-F. Chou, O. Bakajin, S. W. P. Turner, T. A. J. Duke, S. S. Chan, E. C. Cox, H. G. Craighead, and R. H.
273 Austin, "Sorting by diffusion: An asymmetric obstacle course for continuous molecular separation,"
274 *Proceedings of the National Academy of Sciences*, vol. 96, no. 24, pp. 13762–13765, Nov. 1999.
- 275 [17] M. Cabodi, S. W. P. Turner, and H. G. Craighead, "Entropic Recoil Separation of Long DNA Molecules,"
276 *Analytical Chemistry*, vol. 74, no. 20, pp. 5169–5174, Oct. 2002.
- 277 [18] M. Baba, T. Sano, N. Iguchi, K. Iida, T. Sakamoto, and H. Kawaura, "DNA size separation using artificially
278 nanostructured matrix," *Appl. Phys. Lett.*, vol. 83, no. 7, p. 1468, 2003.
- 279 [19] H. Cao, J. O. Tegenfeldt, R. H. Austin, and S. Y. Chou, "Gradient nanostructures for interfacing microfluidics
280 and nanofluidics," *Appl. Phys. Lett.*, vol. 81, no. 16, p. 3058, 2002.
- 281 [20] L. H. Thamdrup, A. Klukowska, and A. Kristensen, "Stretching DNA in polymer nanochannels fabricated by
282 thermal imprint in PMMA," *Nanotechnology*, vol. 19, no. 12, p. 125301, Mar. 2008.
- 283 [21] J. O. Tegenfeldt, C. Prinz, R. L. Huang, R. H. Austin, S. Y. Chou, E. C. Cox, J. C. Sturm, and H. Cao,
284 "Micro- and nanofluidics for DNA analysis," *Analytical and Bioanalytical Chemistry*, vol. 378, no. 7, pp.
285 1678–1692, Apr. 2004.
- 286 [22] L. J. Guo, X. Cheng, and C.-F. Chou, "Fabrication of Size-Controllable Nanofluidic Channels by
287 Nanoimprinting and Its Application for DNA Stretching," *NanoLett.*, vol. 4, no. 1, pp. 69–73, Jan. 2004.
- 288 [23] Z. Yu, L. Chen, W. Wu, H. Ge, and S. Y. Chou, "Fabrication of nanoscale gratings with reduced line edge
289 roughness using nanoimprint lithography," *J. Vac. Sci. Technol. B*, vol. 21, no. 5, p. 2089, 2003.
- 290 [24] M. J. O'Brien, P. Bisong, L. K. Ista, E. M. Rabinovich, A. L. Garcia, S. S. Sibbett, G. P. Lopez, and S. R. J.
291 Brueck, "Fabrication of an integrated nanofluidic chip using interferometric lithography," *J. Vac. Sci.
292 Technol. B*, vol. 21, no. 6, p. 2941, 2003.
- 293 [25] C. Lee, E. H. Yang, N. V. Myung, and T. George, "A Nanochannel Fabrication Technique without
294 Nanolithography," *NanoLett.*, vol. 3, no. 10, pp. 1339–1340, Oct. 2003.

- 295 [26] W.-C. Jung, Y.-M.Heo, G.-S.Yoon, K.-H.Shin, S.-H.Chang, G.-H.Kim, and M.-W. Cho, "Micro Machining
296 of Injection Mold Inserts for Fluidic Channel of Polymeric Biochips," *Sensors*, vol. 7, no. 8, pp. 1643–1654,
297 Aug. 2007.
- 298 [27] "Lab-On-A-Chip: Microfluidics on ice," *Nature Methods*, vol. 10, no. 3, pp. 194–194, Feb. 2013.
- 299 [28] S. Arya, S. Khan, A. Vaid, H. Kour, P. Lehana, "Microfluidic Mechanics and Applications: a Review,"
300 *Journal of Nano- and Electronic Physics*, vol. 5, No. 4, pp. 04047-1-04047-12, 2013..
- 301 [29] S. Jayaraj, S. Kang, and Y. K. Suh, "A review on the analysis and experiment of fluid flow and mixing in
302 micro-channels," *Journal of Mechanical Science and Technology*, vol. 21, no. 3, pp. 536–548, Mar. 2007.
- 303 [30] S. Li, M. Li, Y. S. Hui, W. Cao, W. Li, and W. Wen, "A novel method to construct 3D electrodes at the
304 sidewall of microfluidic channel," *MicrofluidNanofluid*, vol. 14, no. 3–4, pp. 499–508, Mar. 2013.
- 305 [31] P. B. Vainshtein and A. L. Yarin, "Multiphase fluid dynamics," *International Journal of Multiphase Flow*,
306 vol. 18, no. 1, pp. 157–158, Jan. 1992.
- 307 [32] "Surface Tension-Confined Confined Microfluidics," Springer Reference, 2011.
- 308 [33] M. Trojanowicz, "Flow Injection Analysis - Instrumentation and Applications."
- 309 [34] "Laminar to Turbulent Flow Transition," *SpringerReference*, 2011.
- 310 [35] P. Tarazona, "Capillarity and Wetting," *Phys. Scr.*, vol. T19B, pp. 369–374, Jan. 1987.
- 311 [36] M. T. Kreutzer, F. Kapteijn, J. A. Moulijn, and J. J. Heiszwolf, "Multiphase monolith reactors: Chemical
312 reaction engineering of segmented flow in microchannels," *Chemical Engineering Science*, vol. 60, no. 22,
313 pp. 5895–5916, Nov. 2005.
- 314