

Original Research Article

The effects of short-term repeated oral administration of potassium cyanide on some haematological indices and internal organs morphology of rabbits

ABSTRACT

This study investigated the effects of short-term repeated oral administration of sub-toxic dose of potassium cyanide on the haematological indices and the structure of the thyroid, liver, adrenal, and spleen of rabbits. A total of 16 rabbits, weighing 1.2 ± 0.2 kg were randomly divided into two groups. Group 1 was the control, and the animals were treated with 10 mL/kg body weight of distilled water *per os*. Group 2 was treated with 0.3mg/kg potassium cyanide (KCN) in distilled water *per os*. Results revealed atrophy and distended thyroid follicles with flattened epithelial cells only in the cyanide treated group. The liver revealed severe periportal lymphocytic infiltration only in the cyanide treated animals, coupled with focal areas of hepatocellular coagulative necrosis, and cholangitis. The spleen revealed mild congestion of the red pulp in both treated and control groups, while hemosiderosis was seen only in the cyanide treated group. There was no visible lesion in the adrenal gland. The values of parameters evaluated in the KCN- treated animals were as follows: Packed Cell Volume (PCV) ($33.25 \pm 2.4\%$), Red blood Cell Count (RBC) ($6.93 \pm 0.7 \times 10^6/\mu\text{L}$), TWBC (Total White Blood cell Count) ($9.4 \pm 1.0 \times 10^3/\mu\text{L}$), Haemoglobin Concentration (HC) (14.7 ± 1.9 g/dL), Aspartate Transaminase (AST) (29.8 ± 5.7 IU/mL), Alanine amino transaminase (ALT) (12.8 ± 1.8 IU/mL) and Alkaline Phosphatase (ALP) (48.0 ± 5.7 IU/mL). Those of controls were PCV ($31.0 \pm 0.94\%$), RBC ($5.45 \pm 0.3 \times 10^6/\mu\text{L}$), TWBC ($6.8 \pm 0.43 \times 10^3/\mu\text{L}$), HC (11.07 ± 0.94 g/dL), AST (16.33 ± 0.3 IU/mL), ALT (8.33 ± 1.0 IU/mL), ALP (23.7 ± 2.8 IU/mL). There was no significant difference ($p < 0.05$) between the haematological indices of the treated and the control group. AST and ALP of the treated group was significantly higher ($p < 0.05$) than that of the control.

Key words: Potassium cyanide, liver, thyroid, sub-toxic dose, hematological indices.

1. INTRODUCTION

Cyanogenic glycosides are substances present in many plants that can produce highly toxic hydrogen cyanide (HCN) and the contents of this substance can be as high as 100 – 800 mg/kg of the plant material (Conn, 1978). Enzymatic conversion enhanced when plant cells are damaged or stressed, of the glycosides is as it occurs when the plant is chewed, crushed, droughted, wilted, or frozen. A myriad of plant species are known to contain cyanogenic glycosides with the potential to produce HCN poisoning. Some of these plants are grown as food sources for humans and animals, for example, sorghum (*Sorghum* spp.), corn (*Zea mays*), clovers (*Trifolium* spp.), and manihot or cassava (*Manihotesculenta*). Although cyanide most commonly occurs as hydrogen cyanide, and in salt forms, such as sodium and potassium cyanide, it also occurs naturally in cassava (*manihot esculenta* Cranz) as linamarin, a cyanogenic glycoside (Kamalu, 1995). Cassava roots are a major source of calories for over 500 million people in the tropics, and the leaves are also used as vegetable in soups (FAO, 2002; Maduagwu and Umoh, 1982). This increasing dependence of both man and animals on cassava and maize-based foods has made further study into the possible adverse effects of cyanide necessary. A relationship has also been suggested between pancreatic diabetes and prolonged exposure to the cassava (McMillian and Geevarghese, 1997). While there is substantial information on the effect of cyanide generally (Faust, 1994), not much is known about the specific effects of sub-chronic or repeated short term oral administration of cyanide in rabbits, especially its effects on the haematology and some structures of the internal organs. The objective of this study is therefore to evaluate the effects of short-term repeated oral administration of potassium cyanide on the structure of the thyroid, liver, adrenal, spleen as well as the liver markers (AST, ALP and ALT) and the haematological indices (RBC, WBC and Hb) of rabbits.

2. MATERIAL AND METHOD

2.1 Experimental animals

Pre-pubertal rabbits, weighing 1.2 ± 0.2 kg were purchased locally. They were acclimatized for 3 weeks and were ascertained to be in good health. The rabbits were housed in standard cages in a room with daily temperature range between 20°C and 28°C. All animal s had access to feeds (both freshly cut grass and Vital® feeds Ltd, Nigeria) and water *ad libitum*, and were exposed to a 12-hour light-dark cycle. The laboratory animals were handled in accordance with the good laboratory practice regulation as contained in the Helsinki Declaration of 1975, as revised in 2000 and 2008.

Potassium Cyanide (KCN) was procured from BDH Chemicals, UK.

2.2 Experimental Design

A total of 16 rabbits, weighing 1.2 ± 0.2 kg were randomly selected into two groups. Group 1 was the control, and the animals were treated with 10 mls/kg body weight of distilled water. Group 2 was treated with 0.3mg/kg body weight of potassium cyanide (KCN) reconstituted in distilled water. Both the distilled water and the KCN were administered daily through the oral route using an improvised oro-gastric canula. Animal weights were regularly taken in order to effect any necessary adjustment(s) to the dose of KCN administered. The animals were treated for 30 days, at the end of which the animals were mildly euthanized using chloroform chamber anaesthesia. Blood samples were collected into EDTA bottles. The organs thyroid, liver, adrenal, and spleen were collected for histological examination as described by Bancroft and Stevens (1977). Other parameters evaluated were erythrocyte count (EC) and total white blood cell counts (TWBC), which were assayed using the method of Schalm et al., 1975; haemoglobin concentration (Hb) which was assayed using the method of monophosphate method as described by Klein et al (1960), serum ALT was estimated colorimetrically by the 2, 4-dinitrophenylhydrazine (DNPH) method of Reitman and Frankel (1957) as described by Bergmeyer (1974) while serum AST was estimated by the Reitman and Frankel (1957) colorimetric method using a QCA test kit (Quimica Clinica Applicada, Spain).

3. RESULTS

The results on the haematological indices and the three liver enzymes assayed are presented in the Table 1. The values for control groups were as follows: PCV ($31.0 \pm 0.94\%$), RBC ($5.45 \pm 0.3 \times 10^6/\mu\text{L}$), TWBC ($6.8 \pm 0.43 \times 10^3/\mu\text{L}$), HC ($11.07 \pm 0.94\text{g/dL}$), AST ($16.33 \pm 0.3 \text{ IU/mL}$), ALT ($8.33 \pm 1.0 \text{ IU/mL}$), ALP ($23.7 \pm 2.8 \text{ IU/mL}$) and those of KCN-treated animals were as follows: PCV ($33.25 \pm 2.4\%$), RBC ($6.93 \pm 0.7 \times 10^6/\mu\text{L}$), TWBC ($9.4 \pm 1.0 \times 10^3/\mu\text{L}$), HC ($14.7 \pm 1.9 \text{ g/dL}$), AST ($29.8 \pm 5.7 \text{ IU/mL}$), ALT ($12.8 \pm 1.8 \text{ IU/mL}$) and ALP ($48 \pm 5.7 \text{ IU/mL}$). The results showed that there was no significant difference ($p < 0.05$) between the haematological indices of the treated and the control group. AST and ALP of the treated group was significantly higher ($p < 0.05$) than that of the control.

The results of the effect of cyanide treatment on the structures of the liver, thyroid, adrenal and spleen of the rabbits are presented in figs 1 to 3. The results revealed atrophy and distended thyroid follicles with flattened epithelial cells in the cyanide treated group (Figs. 1A & 1B). The liver revealed severe periportal lymphocytic infiltration in the cyanide treated animals, coupled with focal areas of hepatocellular coagulative necrosis, and cholangitis. (Figs. 2A & 2B), The spleen revealed mild congestion of the red pulp in both treated and controls,

while hemosiderosis was seen only in the cyanide treated group (Figs. 3A & 3B). There was no visible lesion in the adrenal gland.

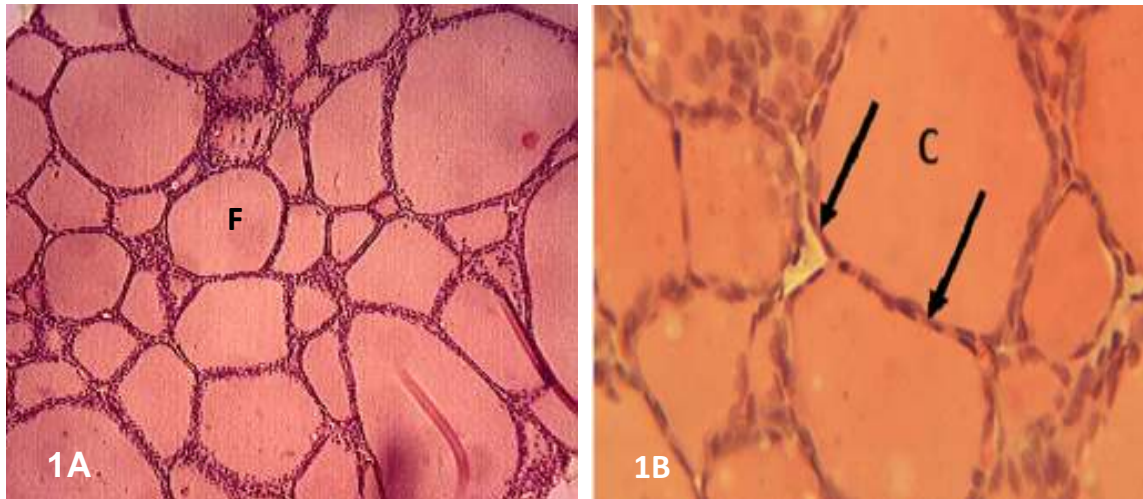


FIG.1A: Thyroid of animal in group 1 (10mL/kg distilled water) showing no visible pathological lesion, F=follicle distended with colloid, while in **FIG 1B:** shows thyroid parenchyma of animal in treated group 2 (0.3mg/kg KCN) with atrophy of thyropid epithelium with distended follicles and flattened epithelial cells (arrows), C= colloid. H &E stain , x40; x200)

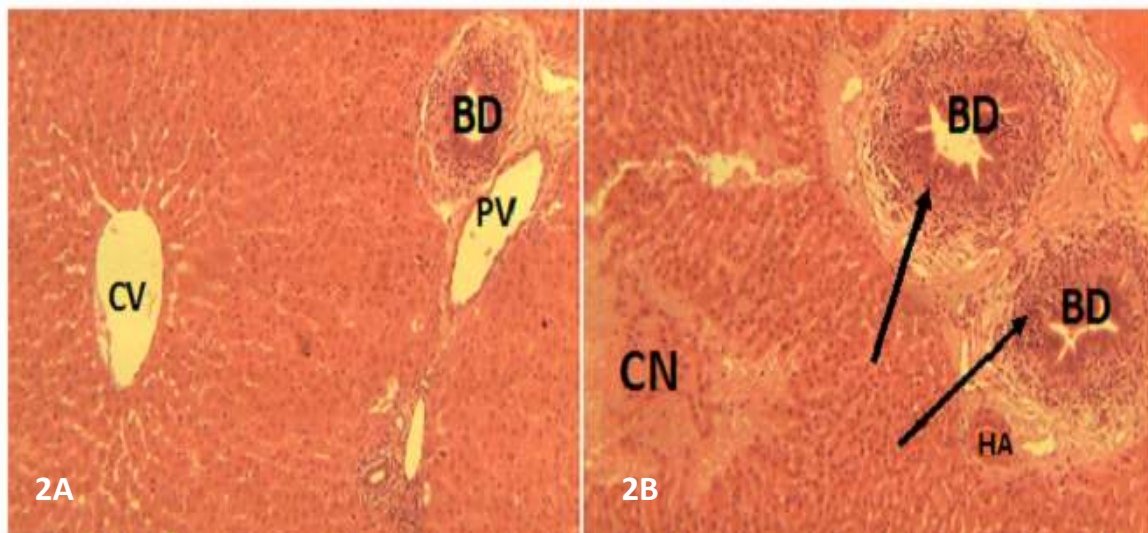


FIG. 2A: Histology of the liver of group 1 (10mL/kg distilled water) showing mild periportal lymphocytic infiltration around bile duct (BD), PV=portal vein, CV=central vein. **FIG.2B:** The histology of the liver in treated group 2 (0.3mg/kg KCN), showing the liver with hepatocellular coagulative necrosis (N) and periportal lymphocytic infiltration with marked cholangitis (arrows), HA=hepatic artery. H & E stain x40.

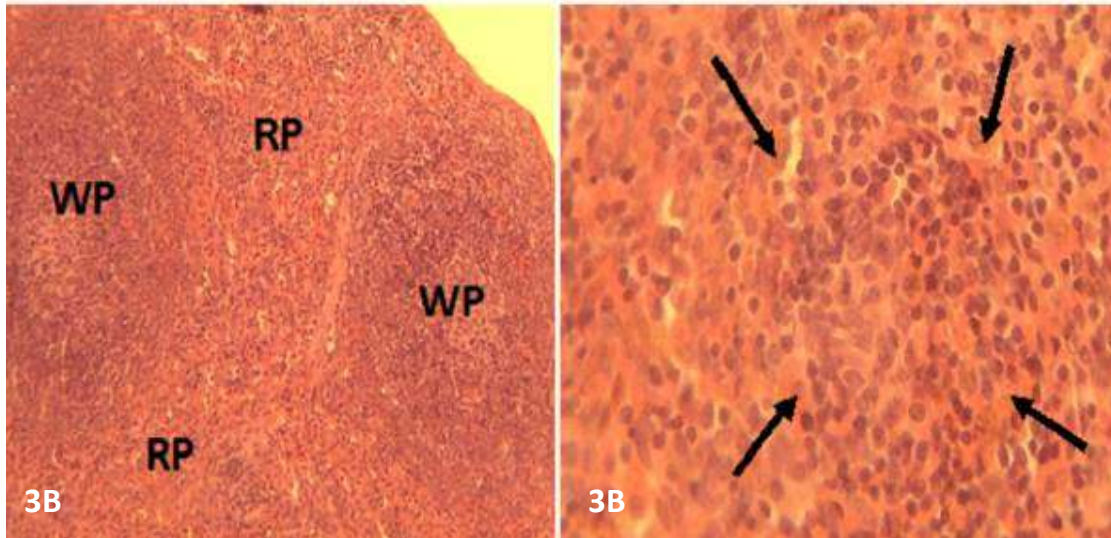


FIG.3A: Histologic changes of the spleen of group one (10mL/kg distilled water) showing mild congestion of red pulp (RP) and **3B:** showing that in group 2 (0.3mg/kg KCN) showing severe congestion of red pulp with hemosiderosis (arrows), white pulp (WP). H & E (X40).

Table 1: Mean hematological indices and three liver enzyme levels of rabbits administered short-term repeated sub-lethal dose of potassium cyanide (KCN)

parameters	Groups	
	1	2
PCV (%)	31.0 ± 0.94	33.25 ± 2.4
RBC (X 10 ⁶ /μL)	5.45 ± 0.3	6.93 ± 0.7
TWBC (×10 ³ /μL)	6.8 ± 0.43	9.4 ± 1.0
HC (g/dL)	11.07 ± 0.94	14.7 ± 1.9
AST (IU/mL)	16.33 ± 0.3	29.8 ± 5.7*
ALT (IU/mL)	8.33 ± 1.0	12.8 ± 1.8
ALP (IU/mL)	23.7 ± 2.8	48.0 ± 5.7*

* Significance at $p \leq 0.05$.

Group 1 was administered 10 mL/kg body weight, bw distilled water.

Group 2 was administered 0.3 mg/kg bw KCN.

4. DISCUSSION

The hepatic effects observed in this study which includes mild-severe periportal infiltration of lymphocytes, cholangitis and focal areas of hepatocellular coagulative necrosis indicates the toxic effects of cyanide even at low doses. This likely explains the increase in the levels of the serum enzymes assayed especially AST and ALP. Focal necrosis, congestion, fatty degeneration, hydropic degeneration, and severe cytoplasmic vacuolization of hepatocytes have been reported as hepatotoxic effects of KCN in both man and animal (Okolie and Osagie, 1999, 2000; Kamalu, 1993; Sousa et al., 2002; Soto-Blanco and Gorniak, 2003). Severe

cytoplasmic vacuolization of hepatocytes was observed in male rats that ingested 3.6 mg/kg/day of KCN in drinking water for 15 days, however hepatic lesion were minimal at 0.36 - 1.2 mg/kg/day, and absent at 0.12 mg/kg/day (Sousa et al., 2002). The periportal inflammatory response observed in this study however appears to be due to factors other than cyanide, as similar picture was seen in control animals though to a lesser extent. In Nigeria, a popular cassava meal (gari), which may be consumed at least once a day in many homes, is reported to release 128µmol of cyanide per 150g of diet. This value is relatively small when compared to a minimum of 5.76 mmol daily cyanide ingestion in the present protocol, which consequently would have been expected to produce more toxic effects. However, the fact that there is continuous ingestion of this and related food products that contain cyanogenic glycosides in both man and animals calls for worry especially to consumers of products that contain cyanogenic materials even at low doses. The fact that there was no significant difference in the haematological profile between the control and the treated groups suggest that cyanide poisoning may not lead to anaemia.

CONCLUSION

In conclusion, the study demonstrated that repeated administration of sub-toxic dose of cyanide may not have caused anaemia but could lead to liver and thyroid damage in the rabbits.

ETHICAL APPROVAL The authors hereby declare that "Principles of laboratory animal care" (NIH publication No. 85-23, revised 1985) were followed. All experiments have been examined and followed the appropriate guidelines of Ethics and Research committee of University of Nigeria (2005 Revision).

REFERENCES

- Bancroft JD, Stevens A. Theory and Practice of Histological Techniques. Churchill Livingstone, Edinburgh, 1977: 16–64.
- Conn E. Cyanogenesis, the production of cyanide, by plants. In: R.F Keeler, K.R. Van Kampen and Lf James, Editors, Effects of Poisons Plants on Livestock, Academic Press, San Diego. 1978: 301 – 310.
- Food and Agricultural Organisation (FAO). Partnership formed to improve cassava, staple food for 600 million people. Rome. FAO report. 2002.
- Faust RA. Toxicity summary for cyanide. Oak Ridge Reservation Environmental Restoration Program (Report), 1994.

- 164 Kachmar JF. Determination of blood haemoglobin by the cyanomethaemoglobin procedure. In: Tietz, N.W. Ed.,
165 Fundamentals of Clinical Chemistry. W. B. Saunders Company, Philadelphia, 1970: 268-269.
- 166 Klein B, Read PA, Babson AL. Rapid method for the quantitative determination of serum alkaline phosphatase.
167 Clinical Chemistry. 1960; 6: 269 – 275.
- 168 Kamalu BP. The adverse effects of long-term cassava (*Manihot esculenta Crantz*) consumption. Inter. J. Food
169 Sci. Nutrition. 1993; 46: 65-93.
- 170 Maduagwu EN, Umoh IB. Detoxification of cassava leaves by simple traditional methods, Toxicol. Letter. 1982;
171 10; 245-48.
- 172 Mcmillian DE, Geevarghese PJ. Dietary cyanide and tropical malnutrition diabetes. Diabetes Care. 1997; 2: 202
173 – 208.
- 174 Okolie NP, Osagie AU. Liver and kidney lesions and associated enzyme changes induced in rabbits by chronic
175 cyanide exposure. Food Chem. Toxicol., 1999; 37: 745-750.
- 176 Poulton JE. Cyanogenic compounds in plants and their toxic effects. In: (R.F.Keeler and A.T Tu, Editors)
177 Handbook of natural toxins, Mercel Dekker, New York.1983, pp117 – 157.
- 178 Reitman S, Frankel S. A colorimetric method for determination of serum glutamic oxaloacetic and glutamic
179 pyruvic transaminases. Amer. J. Clin. Pathology.1957; 28: 56 – 62.
- 180 Schalm OW, Jain NC, Carrol EJ. Veterinary Haematology. Lea & Febiger, 3rd edn. Philadelphia. 1975, pp. 17 -
181 269.
- 182 Soto-blanco B, Gorniak SL. Milk transfer of cyanide and thiocyanate: cyanide exposure by lactation in goats. Vet.
183 Research. 2003; 34: 213-220.
- 184 Sousa AB, Soto-Blanco B, Guerra JL,, Kimura ET, Gorniak SL. Does prolonged oral exposure to cyanide
185 promote hepatotoxicity and nephrotoxicity. Toxocol., 2002;174: 87-95.
- 186