# Hepatic Antioxidant Effect of Paroxetine in Rats Exposed to Chronic Restraint Model.

3

- 4 Short running title:
- 5 Paroxetine and hepatic oxidative markers

- 7 Abstract
- 8 Depressed mood could affect the progression and severity of several diseases e.g. hypertension,
- 9 myocardial infarction, gastritis, peptic ulcer etc... Liver is one of the major organ that could be
- affected by chronic exposure to stress because stress may result in hepatic inflammation in
- particular due to accumulation of reactive oxygen species (ROS). The present study was done to
- investigate the potential antioxidant effect of paroxetine, as a selective serotonin reuptake
- inhibitor (SSRI), to protect against chronic restraint stress-induced oxidative damage in the liver.
- 14 Thirty albino rats were divided into 3 equal groups. Group 1 was control, non-stressed non-
- treated group. Group 2 was exposed to chronic restraint model by placing them in wire mesh
- cages exactly fit to their size for 6 hours daily for 21 days. Group 3 were also exposed to chronic
- 17 restraint model for 21 days while they were administered by paroxetine 1 mg/kg/day ip during
- the restraint period. At the end of the study, liver transaminases (ALT and AST) were
- determined by commercial kits. The hepatic levels of GPx, catalase and TBARS were also
- 20 determined by spectrophotometric methods. Glutathione repletion ability by hepatic cells with
- and without paroxetine treatment was also determined in all tested groups.
- The results showed a significant (p<0.05) increase in serum levels of ALT & AST and liver
- 23 levels of GPx and catalase enzymes while levels of TBARS were significantly (p<0.05) reduced
- in paroxetine-treated group compared with non-stressed non-treated control rats. Glutathione
- repletion ability was also significantly (p<0.05) increased in treated group to a level comparable
- to the control non-stressed non-treated values.

restraint model.

54

Paroxetine could possess a protective effect to liver tissue of chronic restrained rats. This 27 hypothesis may help its use to reduce oxidative stress caused by exposure to chronic stress. 28 **Keywords:** Paroxetine, Chronic restraint model, rats, anti-oxidant, liver. 29 30 Introduction 31 Paroxetine is one of the selective serotonin reuptake inhibitors (SSRIs). It is commonly used in 32 the treatment of patients with depressive disorders since the late 1990s [1]. It is generally 33 preferred over tricyclic antidepressants (TCAs) and monoamine oxidase inhibitors (MAOIs) due 34 to lesser adverse effects, good compliance, and comparable efficacy with these older drug groups 35 [2]. 36 Chronic stress exposure is associated with neurodegeneration and a marked change in anti-37 oxidant enzymes. These enzymes are known to have an important role in the prophylaxis against 38 damage of any cell in the body by oxidative stress. Generation of free radicals and reactive 39 40 oxygen species (ROS) is an ordinary result of metabolic processes that occur regularly in human body. However, in case of exposure to chronic stress, they are produced in a very high levels that 41 lead to toxic effects on intracellular components such as DNA, proteins and lipids [3]. These 42 toxic effects result in abnormalities in the signaling processes in the body and increase the 43 incidence of apoptosis of cells of vital organs [4]. Liver and other body's systems are exposed to 44 damaging effects by oxidative stress associated with depressed mood and disorders of important 45 mediators of central nervous system [5-7]. 46 Depressed mood in human and exposure of animals to chronic restraint model are associated 47 with an increase in incidence of lipid peroxidation and production of ROS [8,9]. Newer 48 generations of antidepressant drugs as venlafaxine were found to have the ability to combat the 49 oxidative stress with their abilities to normalize or even increase the activities of potent 50 51 antioxidant markers [10,11]. The aim of the present study is to investigate the possible protective effect of paroxetine, as an 52 SSRI used in treatment of depressed mood, on the liver of albino rats exposed to chronic 53

- Material and methods
- 57 Animals
- Albino rats (150-200 g body weight) were purchased from the animal house (Bilharzial Center of
- 59 Research, Ain Shams University, Cairo, Egypt), housed in controlled environmental conditions.
- They were left for one week for acclimatization. They were housed in cages with a natural light-
- dark cycle and fed on a standard pellet diet and water ad libitum.
- 62 Chemicals
- Paroxetine HCl (Glaxo SmithKline, U.K.) was purchased as an odorless, off-white powder
- 64 whose molecular weight is 329.4, N-(l-naphthyl)ethylenediamine dihydrochloride (ICN
- 65 Biomedicals Inc., Egypt) and diethyl maleate (Merck Biochemicals, Egypt). Serum alanine &
- 66 aspartate transaminases (ALT, AST) spectrum diagnostic kits were obtained from
- 67 Biodiagnostics, Cairo, Egypt. All other chemicals were purchased from Sigma chemicals co.
- 68 Animal Grouping
- Albino rats were divided into 3 groups (N= 10 rats in each group). They were classified as
- 70 follows:
- 71 Group 1: non-stressed non-treated control group which received normal diet
- Group 2: non-treated group but exposed to chronic restraint model and administered only by
- saline ip as a solvent of paroxetine
- Group 3. Paroxetine-treated group: was administered paroxetine dissolved in saline (1 mg/kg
- b.wt./day ip) for 21 days. This dose was selected according to many studies [12, 21, 22]
- 76 Ethics
- All procedures were in accordance with the National Institute of Health's Guide for the Care and
- Use of Laboratory Animals, as well as the guidelines of the Animal Welfare Act (www.nih.gov).
- 79 Justification of Sample size

80	Number of rats in each group = 10 was determined by sample size determination using program
81	of Microlab, version 2005 so as the $\alpha$ value is < 0.05 and $\beta$ value is > 80%. The 10 rats/group is
82	the minimal number sufficient to produce these statistical values determined by the pilot study
83	done before the full experimental research. Values will be statistically analysed using Prism
84	version 3.0 to calculate ANOVA between different studied groups and to determine which
85	group is responsible for the significant changes.
86	
87	Chronic restraint stress procedure
88	Each rat of both groups 2 &3 was placed in a wire mesh restrainer 6 hours daily for 21 days. At
89	the end of the restraint period, rats were moved to their cages.
90	At the end of the experimental period, the animals were weighted, anesthetized with urethane (1
91	g/Kg). Blood samples were collected from the abdominal aorta as follows: the rat was fixed on a
92	wooden plate and the abdominal cavity was opened, then the abdominal aorta was explored at its
93	bifurcation after gentle traction of the viscera using soft tissue. Blood was centrifuged at 5000
94	rpm for 10 minutes for serum separation, and kept at -80°C until measurements of bot ALT &
95	AST.
96	Body weight of all tested rats were constant all over the study. Serum corticosterone was
97	significantly (p<0.05) increased in stressed non-treated rats of group 2 [ $22.5\pm2.3~\mu g/mL$ , mean
98	$\pm SD$ ] compared to control (group 1) and paroxetine-treated group (3) [ $0.9\pm0.01$ & $1.2\pm0.03$
99	$\mu$ g/mL respectively, mean $\pm$ SD].
100	Biochemical Measurements
101	Serum levels of alanine and aspartate transaminases (ALT &AST)
102	They were measured using biochemistry automatic analyzer (Hitachi 7600)
103	
104	Determination of Glutathione Peroxidase [GPx] Enzyme Activity

105	Glutathione peroxidase (GPx) activity in the liver homogenates was measured by the method
106	described by Rotruck, et al [13].
107	Determination of Catalase Enzyme Activity Catalase (CAT) activity in the liver homogenates
108	was assayed colorimetrically as described by Sinha, et al [14] at 620 nm.
109	
110	Hepatic lipid peroxidation Hepatic lipid peroxidation was assayed by measurement of the
111	concentration of TBARS in nmoles/ milligram protein of hepatic tissues according to the method
112	described by Fraga et al.[15]
113	The absorbance of the precipitate of hepatic tissue was measured spectrophotometrically at 532
114	nm using 1,1,5,5-tetraethoxypropane as standard.
115	
116	Determination of Glutathione Repletion Ability by Hepatic Cells of Tested Rats
117	Diethyl maleate (DEM.), a carbonyl compound that rapidly depletes cytosolic glutathione was
118	used to compare glutathione-replenishing abilities of hepatic cells of all tested groups according
119	to Saville [16]. The concentration of glutathione in different samples were expressed in nmoles
120	per milligram of protein.
121	
122	Protein determination
123	The protein content of the liver homogenates was measured spectrophotometrically by Bradford
124	method [17]. Activities of both catalase and glutathione peroxidase enzymes were expressed in
125	Unit/mg tissue protein.
126	
127	
128	Data Analysis

129	Results are expressed as mean $\pm$ SD [Standard Deviation]. Statistical analysis was performed by
130	analysis of variance followed by Tukey's post hoc using GraphPad Prism version 3.00 for
131	Windows 97 (Graph Pad Software, San Diego, CA, USA). Differences with p< 0.05 were
132	considered to be statistically significant.
133	

134	Results
135	I. Effect of treatment with paroxetine on the serum level of alanine and aspartate
136	aminotransferase (ALT & AST) enzymes
137	Chronic restraint of rats of group (2) significantly ( $p < 0.05$ ) reduces the levels of serum
138	ALT & AST . While, paroxetine administration, in a dose of 1 mg/kg/day ip for 21 days,
139	significantly (p<0.05) restores them to levels comparable to control non-stressed, non-
140	treated group (1) (Fig.1)
141	
142	Figure 1
143	
144	Figure (1): Effect of treatment with paroxetine on the serum level of alanine and aspartate
145	aminotraminase (ALT & AST) enzymes of rats compared to both groups 1 &2 .
146	* Significant ( p< 0.05) reduction in levels of both enzymes compared to control non-
147	stressed group 1 and paroxetine-treated stressed group 3.
148	

II. Table (1): Mean  $\pm$  SD changes in the activities of liver CAT, GPx in rats of all tested groups expressed as unit/mg tissue protein of liver homogenates.

			Stressed
			paroxetine-
	Control group	Stressed non-	treated group
		treated group	
Glutathione			
peroxidase	$12.33\pm1.5$	0.55± 0.01*	10.96± 0.65**
Unit <sup>b</sup> /mg tissue			
protein			
Catalase Unit <sup>a</sup> /mg	65.43± 3.2	3.12± 0.42*	57.65± 4.5**
tissue protein			

151

149

150

- 152 Table (1)
- <sup>a</sup> = Moles of hydrogen peroxide consumed per minute.
- 154  $^{b} = \mu g$  of glutathione consumed per minute.
- \*\*P < 0.05, significant reduction in activity of both enzymes in stressed non-treated group versus
- 156 control group.
- \*\*P, 0.05, significant increase in activity of both enzymes by paroxetine administration versus
- stressed non-treated group.

159

160

161

162

164

165

- A significant (p < 0.05) decrease in the activities of these enzymes was demonstrated in stressed non-treated rats. Paroxetine-treated group showed that their activities were significantly (p < 0.05) increased compared to stressed non-treated group and reached levels comparable to that
- recorded with control group (1).

This suggests that paroxetine owns a possible antioxidant action in case of chronic restrained albino rats with a significant (p < 0.05) decrease in their liver transaminases.

167	III. Effect of 21 days administration of paroxetine on thiobarbituric acid-reactive substance
168	(TBARS) in nmol/mg tissue protein of the liver tissue homogenates of all tested albino rats
169	
170	Figure (2)
171	
172	Figure 2: Effect of 21 days administration of paroxetine on thiobarbituric acid-reactive
173	substance (TBARS) in nmol/mg tissue protein of the liver tissue homogenates of the tested
174	rats.
175	Results are expressed as mean $\pm$ SD (n = 10 albino rats/group).
176	A significant (*p < 0.05) decrease in TBARS levels of paroxetine-treated group (3) in
177	comparison to the stressed non treated group (2).
178	• * $p$ <0.05= significant increase in TBARS levels in group (2) compared to the contro
179	non-stressed non-treated rats group (1)
180	• ** $p<0.05$ = significant decrease in TBARS levels in paroxetine-treated group (3
181	compared to the stressed non-treated rats group (2)
182	
183	
184	

IV.Table (2): Effect of treatment with paroxetine on Glutathione repletion ability of hepatic cells expressed nmoles of glutathione per milligram of protein

The table shows changes in glutathione levels expressed as nmol/ mg protein before and 4 hrs. after exposure to 0.5 mM diethyl maleate (DEM). Data were obtained from hepatic cells isolated from either control, stressed non-treated or stressed+paroxetine-treated group.

Groups	Change in glutathione level in hepatic cells					
	Before exposure to	4 hours after exposure	% change in			
	DEM	to DEM	glutathione level			
Control	54.5 ±2.4	55.4 ±3.2	+1.65%			
Stressed non-treated	53.5± 3.4	2.1 ±0.03*	-96.07%			
Stressed paroxetine- treated	$51.2 \pm 3.1$	55.7 ±3.7**	+8.79%			

There is significant (\*p<0.05) reduction in glutathione levels in stressed non-treated rats compared to control group. However, treatment of chronic restrained rats with paroxetine for 21 days significantly (\*\*p<0.05) is likely to protect hepatic cells against acute depletion of glutathione in comparison to group 2 suggesting that ip paroxetine administration would enhance glulathione replenishing ability in hepatic cells of chronic restrained rats.

196	* $p$ <0.05= significant reduction in hepatic glutathione replenishing ability in group (2)
197	compared to the control non-stressed non-treated rats group (1)
198	** $p < 0.05$ = significant increase in hepatic glutathione replenishing ability in paroxetine-treated
199	group (3) compared to the stressed non-treated rats group (2)
200	
201	
202	
203	

#### Discussion 204 Exposure of albino rats, in the present study, to chronic restraint model significantly decreases 205 serum levels of liver transaminases (ALT &AST) and reduces activities of CAT and GPx anti-206 oxidant enzymes in hepatic homogenates of these stressed rats. It also decreases hepatic 207 replenishing ability of glutathione. Ip Administration of paroxetine for 21 days reverses all these 208 changes to almost the levels of control non stressed group. 209 Intraperitoneal (ip) administration of paroxetine for 28 days for anhedonic rats, either due to 210 acute dexamethasone administration or due to exposure to chronic mild stress (CMS) model of 211 anhedonia, reversed the anhedonic effect. These results provide a strong evidence of 212 antidepressant effect of paroxetine as already well-known. Its antidepressant effect against 213 anhedonia-induced by glucocorticoids could be related to suppression of oxidative stress that is 214 associated with dexamethasone administration [18]. 215 Corticotropin-releasing factor (CRF) plays a central role in the stress response by regulating the 216 hypothalamic-pituitary-adrenal (HPA) axis. In response to stress, CRF initiates a cascade of 217 events that causes the release of glucocorticoids from the adrenal cortex. As a result of the great 218 number of physiological and behavioral effects exerted by glucocorticoids, several mechanisms 219 have evolved to control HPA axis activation and integrate the stress response. Glucocorticoid 220 221 feedback inhibition plays a prominent role in regulating the magnitude and duration of glucocorticoid release. In addition to glucocorticoid feedback, the HPA axis is regulated at the 222 level of the hypothalamus by a diverse group of afferent projections from limbic, midbrain, and 223 brain stem nuclei. The stress response is also mediated in part by brain stem noradrenergic 224 225 neurons, sympathetic andrenomedullary circuits, and parasympathetic systems. The HPA axis has an integrated role in the adaptive responses to stress via neuronal and endocrine systems to 226 227 maintain body's homeostasis in a stand-by condition to combat any stressful stimuli [19]. There is a reported relationship between depression and immune system. Results of previous 228 229 studies revealed that depressed mood is often associated with an activation of the immune system functions with a marked secretion of inflammatory cytokines [20]. This increase in 230 immune functions was thought to be related to damaging actions of exposure to oxidative stress. 231 As it was found that activation of immune system results in stimulation of marked synthesis of 232 ROS, instability of anti-oxidant enzymes and down-regulation of their synthesis [8]. Proper 233

234	control of the functions of immune system leads to a positive impact on the synthesis and
235	activities of the antioxidant enzyme mitochondrial manganese superoxide dimutase (SOD2)
236	[21].
237	Chronic paroxetine administration to stressed adult rats produces protective effects against
238	apoptosis and oxidative stress in their hippocampus [22]. These findings could be applied to the
239	results of the present study in relation to the possible hepatoprotective effect of paroxetine in
240	chronic restrained rats by its preservation of antioxidant enzymes and a significant reduction in
241	hepatic TBARS, as a marker of lipid peroxidation.
242	Male Sprague-Dawley rats were exposed to chronic unpredicted stress with administration of
243	Paroxetine (1.8 mg/kg once daily) by an intragastric gavage. The study recorded any change in
244	rat behaviors, activites of some anti-oxidant enzymes: superoxide dismutase and catalase and
245	lipid peroxidation in the form of malondialdehyde level in the serum in addition to the
246	expression of serotonin transporter (5-HTT) in the hippocampus and norepinephrine transporter
247	(NET) in the pons. Results demonstrated that pparoxetine produced a significant reduction in
248	depression-like behaviors and in the malondialdehyde level. Additionally, it increased the
249	activities of superoxide dismutase and catalase anti-oxidant enzymes with an increase in 5-HTT
250	and NET expression. The study concluded that the antidepressive effect of paroxetine could be
251	related partly to overcoming oxidative stress disorders and increasing the expression of 5-HTT
252	and NET that elevated mood of depressed rats [23].
253	Clinically, chronic administration of paroxetine would protect against apoptosis, oxidative stress
254	and improve cytoskeletal functions, intracellular signaling and serotonergic and
255	catecholaminergic neurotransmission. Paroxetine preserves antioxidant enzymes and plays a
256	vital role in the protection of cells from oxidative stress and hence, it will protect intracellular
257	components such as DNA, proteins and lipids and will prevent disrupting signaling pathways
258	that render each cell in the body susceptible to apoptotic or necrotic cell death. It would also
259	protect the central nervous system from the damaging effect of oxidative stress that accompany
260	exposure of human being to depressed mood, stressful conditions or systemic diseases as
261	diabetes and its cardiovascular complications [24].

262	In conclusion, the results of the present study would provide a suggestion that paroxetine
263	possesses an anti-oxidant action that helps in protection of livers of chronic restrained rats
264	Disclosure
265	
266	The author reports no conflicts of interest in this work.
267	
268	
269	
270	
271	
272	

#### 273 References

- 1. Mant A. Rendle, V. A., Hall, W. D., Mitchell, P. B., Montgomery, W. S., McManus, P.
- 275 R., and Hickie, I. B. (2004). Making new choices about antidepressants in Australia: the
- long view 1975-2002. Med. J. Aust. 181, S21–S24.
- 2. Fava, M., and Kendler, K. S. (2000). Major depressive disorder. Neuron 28, 335–341.
- 278 doi: 10.1016/S0896- 6273(00)00112-4
- 3. Ellis, E. M. (2007). Reactive car-bonyls and oxidative stress: poten-tial for therapeutic
- intervention. Pharmacol. Ther. 115, 13–24. doi: 10.1016/j.pharmthera.2007.03.015
- 4. Halliwell, B. (2001). Role of free radicals in the neurodegen- erative diseases: therapeutic
- implications for antioxidant treat- ment. Drugs Aging 18, 685–716. doi:
- 283 10.2165/00002512-200118090- 00004
- 5. Guicciardi, M. E., Deussing, J., Miyoshi, H., Bronk, S. F., Svingen, P. A., Peters, C., et
- al. (2000). Cathepsin B contributes to TNF- $\alpha$  mediated hepatocyte apoptosis by
- promoting mitochondrial release of cytochrome c. J. Clin. Invest. 106, 1127–1137. doi:
- 287 10.1172/JCI9914
- 6. Gould, T. D., and Manji, H. K. (2002). Signaling networks in the pathophysiology and
- treatment of mood disorders. J. Psychosom. Res. 53, 687–697. doi: 10.1016/S0022-
- 290 3999(02)00426-9
- 7. Maser, E. (2006). Neuroprotective role for carbonyl reductase? Biochem. Biophys. Res.
- 292 Commun. 340, 1019–1022. doi: 10.1016/j.bbrc.2005.12.113
- 8. Bilici, M., Efe, H., Koroglu, M. A., Uydu, H. A., Bekaroglu, M., and Deger, O. (2001).
- Antioxidant enzymes and lipid peroxidation in major depression: alterations by
- antidepressant treatment. J. Affect. Disord. 64, 43–51. doi: 10.1016/S0165-
- 296 0327(00)00199-3
- 9. Fontella, F. U., Siqueira, I. R., Vasconcellos, A.P.S., Tabajara, A.S., Netto, C. A., and
- Dalmaz, C. (2005). Repeated restraint stress induces oxidative damage in rat hippocam-
- pus. Neurochem. Res. 31, 105–111. doi: 10.1007/s11064-004-9691-6
- 300 10. Eren I, Naziroğlu M, Demirdaş A, Celik O, Uğuz AC, Altunbaşak A, Ozmen I, Uz
- 301 E.(2007). Venlafaxine modulates depression-induced oxidative stress in brain and
- medulla of rat. Neurochem Res. 32(3):497-505

- 11. Zafir, A., Ara, A., and Banu, N. (2009). In vivo antioxidant status: a putative target of
- antidepressant action. Prog. Neuropsychopharmacol. Biol. Psychiatry 33, 220–228. doi:
- 305 10.1016/j.pnpbp. 2008.11.010
- 12. David D J P, M Bourin, G Jego, C Przybylski, P Jolliet and A M Gardier (2003). Effects
- of acute treatment with paroxetine, citalopram and venlafaxine in vivo on noradrenaline
- and serotonin outflow: a microdialysis study in Swiss mice. Br.J.
- 309 Pharmacol.,140,1128–1136
- 310 13. Rotruck, J. Pope, A. Ganther, H. Swanson, A. Hafeman, D. Hoekstra, W. (1973)
- Biochemical role as a component of Glutathione peroxidase. Science. 79:588–590.
- 312 14. Sinha, K.A. (1972) Colorimetric assay of catalase. Ann Biochem. 47: 389–394
- 313 15. Fraga, C. Leibovitz, B. Tappel, A. (1988) Lipid peroxidation measured as thiobarbituric
- acid reactive substances in tissue slices: characterization and comparison with
- homogenates and microsomes. Free Radic Biol Med.;4:155–161
- 316 16. Saville, B. (1958). A scheme for the colorimetric determination of microgram amounts
- of thiols. The Analyst 83, 670-672.
- 318 17. Bradford, M. (1976) A rapid and sensitive method for the quantitation of microgram
- quantities of protein utilizing the principle of protein-dye binding. Anal Biochem.;
- 320 72:248–254
- 321 18. Casarotto P.C. Andreatini R. (2007) Repeated paroxetine treatment reverses anhedonia
- induced in rats by chronic mild stress or dexamethasone. Eur Neuropsychopharmacol.
- 323 17(11):735-742.
- 19. Smith SM, Vale WW (2006) The role of the hypothalamic-pituitary-adrenal axis in
- neuroendocrine responses to stress. Dialogues Clin Neurosci. 8(4):383-95.
- 20. Maes, M., Scharpè, S., Meltzer, H. Y., Okayli, G., Bosmans, E., D'Hondt, P., et al.
- 327 (1994). Increased neopterin and interferon- gamma secretion and lower availability of L-
- tryptophan in major depression: further evidence for an immune response. Psychiatry
- Res. 54, 143–160. doi: 10.1016/0165-1781(94)90003-5
- 21. Bogunovic, B., Stojakovic, M., Chen, L., and Maric, M. (2008). An unexpected
- functional link between lysosome thiol reductase and mitochondrial manganese
- superoxide dismutase. J. Biol. Chem. 283, 8855–8862. doi: 10.1074/jbc.M708998200

22.	. Karanges E. K	ashem M. Sarke	er R. Ahme	d E	. Ahme	d S. Van N	Vieuwen	huijzen P. K	Cemp
	A. McGregor	L (2013). Hipp	ocampal p	rote	in expre	ession in o	different	ially affecte	d by
	chronic paroxe	etine treatment	in adolesce	nts	and adu	ult rats: A	possib	le mechanisi	m of
	"paradoxical"	antidepressant	responses	in	young	patients.	Front.	Pharmacol	doi:
	10.3389/fphar	00086							

- 23. Qiu HM, Yang JX, Wu XH, Li N, Liu D, Wang LJ, Qin LJ, Zhou QX (2013). Antidepressive effect of paroxetine in a rat model: upregulating expression of serotonin and norepinephrine transporter. Neuroreport. 10;24(10):520-5. doi: 10.1097/WNR.0b013e328362066d.
- 24. Gerö D, Szoleczky P, Suzuki K, Módis K, Oláh G, Coletta C, Szabo C (2013). Cell-based screening identifies paroxetine as an inhibitor of diabetic endothelial dysfunction. Diabetes. 62(3):953-64. doi: 10.2337/db12-0789.

### Figure (1): Effect of treatment with paroxetine on the serum level of alanine and aspartate aminotransferase (ALT & AST) enzymes

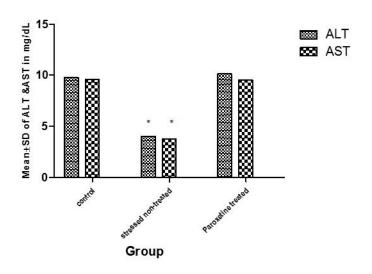


Figure (2): Effect of 21 days administration of paroxetine on thiobarbituric acid-reactive substance (TBARS) in nmol/mg tissue protein of the liver tissue homogenates of all tested albino rats

