

# Fertility Enhancing Potential of *Mucuna Pruriens* Seeds in Female Sprague-Dawley Rats. Ojo Temitope Noah<sup>1</sup>, Gbotolorun Stella Chinwe<sup>1\*</sup>, Oremosu Ademola Ayodele<sup>1</sup>)

(Put \* above the corresponding author and give telephone number, fax number and email ID in the footer)  
1Department of Anatomy, Faculty of Basic Medical Sciences, college of Medicine of the University of Lagos.

## ABSTRACT (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)

**Aims:** To determine the effect of oral administration of methanolic seed extract of *Mucuna pruriens* (*M. pruriens*) on oestrous cycle, ovulation, reproductive hormones and oxidative stress in the ovary of cyclic Sprague-Dawley (S-D) rats.

**Design:** Prospective animal study related to *M. pruriens* in reproductive area. .

**Place and Duration:** Animal Facility of the Department of Anatomy, Faculty of Basic Medical Sciences, College of Medicine of the University of Lagos, Nigeria between the months of June 2012 and August, 2012.

**Methodology:** Forty female S-D rats with regular 4 days cycle averagely weighing 145 g were used. Methanolic extract of *M. pruriens* was given orally at 50, 100 and 200 mg/kg body weight. Oestrous cycle was monitored daily. At the end of the experiment animals were sacrificed by cervical dislocation. Oocytes were counted, blood and ovaries were assayed for hormonal and biochemical studies respectively.

**Results:** Oestrous cycle remained unchanged in the treatment groups. Catalase and superoxide dismutase levels were increased slightly compared to control. A dose dependent increase in FSH and LH (p < 0.05 at 200 mg/kg) levels were observed with an increase in the number of oocytes released at ovulation compared to control.

**Conclusion:** *M. pruriens* seed extract has the potential to enhance fertility by increasing serum levels of FSH and LH which in turn increases the number of oocytes released at ovulation possibly through its antioxidant properties.

**Keywords:** [*Mucuna pruriens*, ovulation, oxidative stress markers, FSH, LH]

\* Tel.: +2348038098631

E-mail address: scgbotl@yahoo.com

## 1. INTRODUCTION (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)

Human health is of prime importance to a country's development and progress. Herbal preparation and medications have been in use for the treatment of diseases and various ailments since ancient times in many parts of the world. In developed countries, despite newer formulations of effective conventional drugs, the treatment of diseases and other ailments with herbal remedies is still very popular (1). In developing countries, the use of herbal remedies as alternatives to

\* Tel.: +xx xx 265xxxxx; fax: +xx aa 462xxxxx.

E-mail address: xyz@abc.com.

28 modern medicine is on the increase. In Nigeria, many indigenous  
29 plants have been used in herbal medicinal preparations to cure  
30 sicknesses and diseases and to heal injuries (2), (3). *M. Pruriens* is one  
31 such plant; it is a tropical legume known as velvet bean ('*Agbara*'- Igbo,  
32 '*Yerepe*' - Yoruba). It is found in Africa, India and the Caribbean's;  
33 where it is widely known for its uses in various ailments as reported in  
34 literature (4), (5), (6). It is a constituent of more than 200 indigenous  
35 drug formulations (6), (7). Some authors have reported that all the  
36 various parts of the *Mucuna* plant possess valuable medicinal  
37 properties (6), (8), (9). Following the discovery, that *Mucuna* seeds  
38 contain l-dopa which is used in the treatment of parkinson's disease;  
39 its demand even in the international market has increased considerably  
40 (6). This demand has motivated Indian farmers to start commercial  
41 cultivation of the *Mucuna* plant. It has widespread cultivation over most  
42 of the subcontinent and is found in bushes, hedges and dry deciduous  
43 low forests throughout the plains of India (7), (10), (11).

44 *M. Pruriens* has been reported to enhance fertility in male rats (12-18)  
45 however; there is a dearth of literature on the effect of *M. pruriens* on  
46 the function of the female reproductive system. This study was carried  
47 out to evaluate the effect of *M. pruriens* on the reproductive function of  
48 the mature female Sprague-Dawley rats.

## 50 2. MATERIAL AND METHODS / EXPERIMENTAL DETAILS / METHODOLOGY 51 (ARIAL, BOLD, 11 FONT, LEFT ALIGNED, CAPS)

### 52 53 2.1 Plant source

54 The *M. pruriens* plant with mature seeds was harvested from Mowe area of Lagos, Nigeria.  
55 Both plant and seeds were identified and authenticated by Professor J.D. Olowokudejo of  
56 the Department of Botany of the University of Lagos. Voucher specimen with accession  
57 number LUH 4922 was deposited in the herbarium of the Department of Botany.

#### 58 2.1.1 Seed extraction

59 The extraction was carried out in the Pharmacognosy Department of the Faculty of  
60 Pharmacy, University of Lagos. Briefly, seeds were obtained from the pods, air-dried and  
61 grounded into fine powder using the mortar and the pestle. 450 g of fine powder was mixed  
62 with alcohol and placed in the Soxhlet apparatus. The mixture was heated at 60 °C and the  
63 extract was obtained by distillation. The powder obtained (107.6 g, 23.9% yield) was stored  
64 at room temperature of 25 °C before use. All dilutions of the extract were made in distilled  
65 water.

#### 66 2.2 Animals

67 Forty mature female S-D rats of two months old weighing 140 - 150 g obtained from the  
68 Animal House of the College of Medicine, University of Lagos, Nigeria were used in this  
69 study. They were housed five animals per cage at the Animal Facility of the Department of  
70 Anatomy, College of Medicine of the University of Lagos, Nigeria. The animals had free  
71 access to water and rat chow purchased from Pfizer Nigeria Limited and was maintained at  
72 12-h light/12-h dark cycle and at temperatures between 25 to 28 °C. The animals were

73 allowed to acclimatize for two weeks before the commencement of the experiment.  
74 Throughout the duration of the experiment, the animals were observed for adverse effects  
75 such as fur loss, diarrhea, bleeding, ataxia, morbidity and mortality resulting from  
76 administration of the extract. All procedures involving animals in this study conformed to the  
77 guiding principles for research involving animals as recommended by the Declaration of  
78 Helsinki and the Guiding Principles in the Care and Use of Animals (19) and were approved  
79 by the Departmental Committee on the use and care of animals and tissue collection.

80

81

82

### 83 2.3 Determination of the oestrous cycle

84 Oestrous cycle was monitored for 24 days. Oestrous cyclicity was determined daily between  
85 8 a.m. and 9:30 a.m. using the vaginal smear method. Vaginal secretion was collected with a  
86 plastic pipette filled with 10  $\mu$ L of normal saline (NaCl 0.9%). The vagina was flushed two or  
87 three times with the pipette and the vaginal fluid was placed on a glass slide. A different slide  
88 was used for each animal. The unstained secretion was observed under a light microscope.  
89 Only animals with a 4-day oestrous cycle were selected for this study.

#### 90 2.3.1 Oestrous Cyclicity Study

91 Twenty rats divided into 4 groups of 5 rats in each were used – A, B, C and D; they were  
92 given daily dosages of *M. pruriens* orally using an oro-gastric tube for 24 days at: 50 mg, 100  
93 mg and 200 mg/kg body weights respectively while group D animals received distilled water  
94 and served as control. The dosages were determined by a previous study (15). Animals  
95 were sacrificed by cervical dislocation. Laparotomy was performed; ovaries were removed,  
96 trimmed of fat and stored at -80 °C for biochemical analysis.

97

#### 98 2.3.2 Ovulation study

99 Twenty animals were used for this study. The animals received a single oral dose of *M.*  
100 *pruriens* at 9 a.m. on the day of proestrus using an oro-gastric tube. The animals in this  
101 group were divided into 4 groups (E, F, G and H). *M. pruriens* was given at a dose of 50, 100  
102 and 200 mg/kg bodyweights to groups E, F and G respectively while group H served as  
103 control and received distilled water. The rats were sacrificed by cervical dislocation the next  
104 day (estrus) at 10 a.m. A ventral laparotomy was performed and the oviduct was dissected  
105 out, placed on glass slides with a drop of saline and covered with cover-slips. This was  
106 squeezed with both sides being gently rocked and each ovum found in the distended  
107 ampulla was counted under a light microscope (20). Control animals received equivalent  
108 volume of distilled water.

109

### 110 2.4 Biochemical analysis

111 The right ovaries were homogenized using a Potter–Elvehjem homogenizer. A 20% (1/5 w/v)  
112 homogenate of the tissue was prepared in 50 mM Tris-HCl buffer (pH 7.4) containing 1.15%  
113 potassium chloride and centrifuged at 10,000 rpm at 4 °C for 10 min.

114 Superoxide dismutase (SOD) was assayed utilizing the technique of (21). A single unit of  
115 enzyme was expressed as 50% inhibition of Nitroblue tetrazo-lium (NBT)  
116 reduction/min/mg/protein.

117 Catalase (CAT) was assayed colorimetrically at 620 nm and expressed as  $\mu$ moles of H<sub>2</sub>O<sub>2</sub>  
118 Consumed/min/mg/protein as described by (22).

119

## 120 2.5 Hormonal assay studies

121 Blood was obtained from the angular vein of the eye of the S-D rats at 6 p.m. in the evening  
122 of proestrus and collected into heparinised bottles. Each blood sample was spun at 2,500  
123 rpm for 10 minutes in an angle-head desktop centrifuge at temperatures of 25 °C. Serum  
124 samples were assayed in batches with control sera at both physiological and patho-logical  
125 levels by Standard Quantitative Enzyme- Linked Immunosorbent Assay (ELISA) technique  
126 with Microwell kits from Syntro Bioresearch Inc., California, USA.

127

## 128 2.6 Statistical analysis

129 Results were analyzed and expressed as Mean  $\pm$  SD and were subjected to one-way  
130 ANOVA with Newman-Kenls post hoc test version 5.0 for windows. Statistical significance  
131 was considered at P = .05.

## 132 3. RESULTS AND DISCUSSION

133

134 All the treated rats showed normal behaviour throughout the study. No signs of adverse  
135 effects were observed; no fur loss, diarrhoea, bleeding, ataxia, morbidity nor mortality.

### 136 3.1 Oestrous Cycle

137 Analysis of the oestrous cycle revealed that oral administration of 50 mg, 100 mg and 200  
138 mg/kg body weight of methanolic seed extract of *M. pruriens* did not produce any  
139 irregularity/derangement in the cycle pattern. Also, length of cycle remained unchanged in all  
140 the treated rats; animals showed a regular four days cycle as shown (Table 1). Estrogens  
141 and progesterone are important in the normal functioning of the female reproductive system.  
142 They are responsible for the development and maturation of reproductive organs and also  
143 provide the proper environment required for the transport of gametes and nidation. The  
144 balance in hormonal interplay between estrogens and progesterone is responsible for a  
145 normal regular cycle (23), (24). This study revealed that the oral administration of *M.*  
146 *pruriens* seed extract did not alter the oestrous cycle in all the treated animals throughout the  
147 treatment period of 24 days. The treated animals maintained a normal cycle pattern and  
148 cycle length that was comparable with the control animals. Although authors did not  
149 determine the levels of progesterone and estrogens in this study however, we can deduce  
150 from our findings that *M. pruriens* did not produce any negative effect either directly on the  
151 pituitary or indirectly on the hypothalamus to disrupt the intricate balance in hormonal  
152 interplay between progesterone and estrogens levels that is necessary to maintain a normal  
153 cycle.

154

155 Table 1: Effect of the oral administration of *M. pruriens* for 24 days on the length of the  
156 oestrous cycle in S-D rats.

157 Treatment groups                      Length of oestrous cycle in days

158 Control                                      4.0 ± 0.00

159 50 mg/kg                                    4.0 ± 0.10

160 100 mg/kg                                  4.0 ± 0.20

161 200 mg/kg                                  4.0 ± 0.40

162 *n* = 5. Values are expressed as mean ± standard deviation

163 3.2            Antioxidant status of CAT and SOD

164 The extract exhibited a dose dependent increase in CAT and SOD activities in the treatment  
165 groups compared to the control group however, this increase was not statistically significant  
166 (Table 2).

167 Literature is rife with studies reporting that *M. pruriens* has an excellent scavenging ability  
168 that mops up excessive production of reactive oxygen species (ROS) and free radicals (17),  
169 (25), (26), (27), (28). ROS plays both a physiological as well as a pathological role in the  
170 female reproductive tract. Numerous animal and human studies have demonstrated the  
171 presence of ROS in the female reproductive tract such as in the ovaries (29), (30), (31), the  
172 fallopian tubes (32) and in embryos (33). ROS is involved in the modulation of an entire  
173 spectrum of physiological reproductive functions such as oocyte maturation, ovarian  
174 steroidogenesis, corpus luteal function and luteolysis (29), (31), (34). On the other hand, the  
175 pathological effects are exerted by various mechanisms including lipid damage, inhibition of  
176 protein synthesis, and depletion of ATP (35). ROS have been implicated in more than 100  
177 diseases (36), (37), (38). The superoxide radical is formed when electrons leak from the  
178 electron transport chain (39). SOD decomposes superoxide anion into hydrogen peroxide  
179 and oxygen at very high rates. Superoxide radical is involved in diverse physiological and  
180 pathophysiological processes (40). CAT catalyses the decomposition of hydrogen peroxide  
181 to water and oxygen. High concentration of hydrogen peroxide is deleterious to cells such as  
182 DNA, proteins, and lipids, leading to mutagenesis and cell death (41).

183 The slight increase in the activities of SOD and CAT observed in the ovary in this study is an  
184 indication of the antioxidant properties inherent in *M. pruriens*. The upregulation in these  
185 markers of oxidative stress is in response to ROS and free radicals. As earlier stated, both  
186 animal and human studies have demonstrated the presence of ROS in the ovary.  
187 **Phytochemical** analysis has shown that *M. pruriens* seeds contain flavonoids (27), (42) and  
188 tannins (6). Flavonoids and tannins are phenolic compounds, and plant phenolics are a  
189 major group of compounds that act as primary antioxidants or free radical scavengers (43).  
190 In addition, *M. pruriens* seeds are a rich source of L-Dopa and its metabolites. In vitro  
191 antioxidant assays have supported the antioxidant property of L-Dopa (44). Dopamine, a  
192 product of L-Dopa metabolism, has also been found to possess strong anti-oxidant capacity  
193 and free radical scavenging activity (45), (46). Antioxidants prevent oxidative stress caused  
194 by free radicals which damage cells and vital biomolecules. They terminate chain reactions  
195 triggered by free radicals by removing free radical intermediates and inhibit other oxidation

196 reactions (47). The antioxidant capacity of the extracts may be attributed to the presence of  
197 L-Dopa and its metabolite, dopamine and also, the identified phytochemicals.

198

199 Table 2: Effect of the oral administration of *M. pruriens* on the enzymatic antioxidant  
200 activities of CAT and SOD in the ovary of S-D rats.

201	Treatment groups	SOD (min/mg protein)	CAT (Mmol/min/mg protein)
202	Control	1.50 ± 0.30	60.17 ± 16.50
203	50 mg/kg	1.65 ± 0.37	60.83 ± 16.57
204	100 mg/kg	1.67 ± 0.56	61.14 ± 15.30
205	200 mg/kg	1.88 ± 0.90	62.67 ± 17.40

206 **n = 5. Values are expressed as mean ± standard deviation.**

207

208 3.3 Serum concentrations of Follicle Stimulating (FSH) Hormone and Luteinizing Hormone  
209 (LH)

210 A dose dependent increase in serum concentrations of FSH and LH was observed. This  
211 increase was significant for LH at 200 mg/kg (Table 3). Our study showed a dose dependent  
212 increase in the levels of FSH and LH compared to the control. Increase in LH was significant  
213 at 200 mg/kg body weight of the extract. Treatment with *M. pruriens* significantly improved  
214 blood levels of dopamine, adrenaline and noradrenaline in infertile males (13). L-Dopa and  
215 its metabolite dopamine have been reported to stimulate the hypothalamus and forebrain to  
216 secrete gonadotropin-releasing hormone (GnRH) (13), (17), (48). This ultimately will activate  
217 the anterior lobe of the pituitary gland to secrete FSH and LH. The report of this study is in  
218 agreement with studies carried out by elegant researchers in other parts of the world on both  
219 animal and human males in which FSH and LH levels increased significantly following the  
220 administration of *M. pruriens*. (13), (14), (16), (26).

221

222

223

224 Table 3: Effect of the oral administration of *M. pruriens* on serum concentrations of FSH and  
225 LH at 6.00 p.m. on proestrus.

226	Treatment groups	FSH	LH
227	Control	1.83 ± 0.77	1.13 ± 0.15
228	50 mg/kg	1.87 ± 0.04	1.56 ± 0.81
229	100 mg/kg	1.95 ± 0.63	1.90 ± 0.51

230 200 mg/kg 2.01 ± 0.02 2.03 ± 0.76\*

231 n = 5. Values are expressed as mean ± standard deviation. \* P < 0.05

### 232 3.4 Ovulation and number of ova shed

233 A slight increase in the number of oocytes released in the oviduct was observed in the  
234 treated animals compared to the control (Table 4). LH is critical to ovulation because it is  
235 responsible for all the processes and events that accompany ovulation. The rapid surge of  
236 LH that occurs between 5 to 7 p.m. in the evening of proestrus induces follicular rupture and  
237 ovulation in rats (49). The present study showed a dose dependent increase in LH levels  
238 and a slight increase in oocyte number at 50 and 100 mg/kg body weights. However, at the  
239 highest dosage of 200 mg/kg we recorded a significant increase in the levels of circulating  
240 LH and also a concomitant increase in the number of oocytes released at ovulation  
241 compared to the control. Increase in testosterone levels resulting from increase in circulating  
242 LH levels have been recorded following treatment with *M. pruriens* in both human and  
243 animal studies. This in turn has increased fertility indices such as sperm count, sperm  
244 motility, sperm morphology and libido (12), (16), (17), (26). Therefore, the report of this study  
245 suggests that the increasing levels of circulating LH from the anterior pituitary produced by  
246 the administration of *M. pruriens* was responsible for the increase in the number of oocytes  
247 released at ovulation.

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250

251

252 Table 4: Effect of the oral administration of a single dose of *M. pruriens* on the number of  
253 ova shed in the oviduct in the morning of estrus in S-D Rats.

254 Treatment groups	Number of ova shed in the oviduct
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255 Control	7.5 ± 2.40
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256 50 mg/kg	7.6 ± 1.30
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257 100 mg/kg	7.6 ± 1.50
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258 200 mg/kg	8.1 ± 2.50
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259 n = 5. Values are expressed as mean ± standard deviation.

260

## 261 4. CONCLUSION

262

263 *M. pruriens* enhances fertility in female Sprague-Dawley rats by producing a dose dependent  
264 increase in FSH and LH which in turn increased the number of oocytes released at ovulation  
265 possibly through its rich source of L-Dopa and its metabolite, dopamine. At higher dosages  
266 than that was administered in this study, it is possible that a significant increase in the  
267 number of ova shed may be observed. Thus, the use of *M. pruriens* in the treatment of  
268 female infertility caused by anovulation seems promising.



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273 Biochemical analysis.

274

275 **COMPETING INTERESTS**

276

277 “Authors declare that no competing interests exist”.

278

279 **AUTHORS’ CONTRIBUTIONS**

280

281 First author wrote the first part of the manuscript, managed the analysis of the study and  
282 performed the statistical analysis. Second author designed the study, performed literature  
283 search and wrote the final manuscript in accordance with the guideline of this journal. Third  
284 author wrote the protocol and edited the manuscript for submission.

285

286 “All authors read and approved the final manuscript.”

287

288 **ETHICAL APPROVAL (WHERE EVER APPLICABLE)**

289

290 **“ALL AUTHORS HEREBY DECLARE THAT “PRINCIPLES OF LABORATORY**  
291 **ANIMAL CARE” (NIH PUBLICATION NO. 85-23, REVISED 1985) WERE**  
292 **FOLLOWED, AS WELL AS SPECIFIC NATIONAL LAWS WHERE APPLICABLE.**  
293 **ALL EXPERIMENTS HAVE BEEN EXAMINED AND APPROVED BY THE**  
294 **APPROPRIATE ETHICS COMMITTEE”**

295

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## 412 **DEFINITIONS, ACRONYMS, ABBREVIATIONS**

413 Here is the Definitions section. This is an optional section.

414 **Term:** Definition for the term

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## 416 **APPENDIX**