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Research paper

Title: Phytochemical Screening and Investigation of the central and peripheral Analgesic and Anti-Inflammatory activity of ethanol extract of Hiptage Bengalensis (L) Kurz

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Abstract:

25 **Objective:** *This research was focused on the qualitative and quantitative evaluation of anti-*
26 *inflammatory and analgesic effects of Hiptage Bengalensis in laboratory animals and whether*
27 *these effects were of any statistical significance. Phytochemical analysis of ethanol extract of*
28 *Hiptage Bengalensis has indicated the presence of steroid, carbohydrate, flavonoid, alkaloid,*
29 *tanin, phenol and, mangiferin and terpenoid-compounds (1).*

30 **Materials and Method:** *Carrageenan induced Hind Paw Edema test in Long Evans rat was the*
31 *experiment for anti-inflammatory activity of the ethanol extract of Hiptage Bengalensis while*
32 *Hot Plate test and Acetic Acid induced Writhing method were was carried out to assess its*
33 *analgesic activity in Swiss albino mice. At two different doses of 250 and 500 mg/kg body*
34 *weight, the analgesic test was evaluated on mice and the anti-inflammatory test was evaluated*
35 *on rats by the ethanol extract of the leaf.*

36 **Result:** *The experimental activities for the ethanol extract of Hiptage Bengalensis exhibited*
37 *statistically significant ($p < 0.05$) anti-inflammatory activity in Carrageenan induced Hind Paw*
38 *Edema in Long Evans rat and analgesic activity by Hot Plate and acetic acid induced writhing*
39 *method in Swiss albino mice.*

40 **Conclusion:** *In conclusion, these observations provide evidence and possible mechanisms of*
41 *action for the anti-inflammatory and analgesic properties of leaf of Hiptage Bengalensis*
42 *claimed in Ayurveda medicine. Further studies should be undertaken to correlate the*
43 *pharmacological activities with the chemical constituents of the leaf of Hiptage Bengalensis.*

44

45 **Key words:**

46 Analgesic, Anti-inflammatory, Carrageenan, Acetic Acid, Hiptage Bengalensis, Phytochemical.

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I. Introduction

50
51 Hiptage Bengalensis (L) Kurz belongs to the family Malphigiaceae. It is a large,
52 woody climber found in India , South East Asia and Philippines .This is a woody
53 climbing shrub with clusters of pink to white and yellow fragrant flowers and 3-
54 winged, helicopter-like fruits. Flowers look like a decorative accessory. The
55 fragrance is very strong and pleasant, resembles fruity perfume. Leaves are
56 narrow and drooping. The leaves and bark are hot, acrid, bitter, insecticidal,
57 vulnerary , astringent, refrigerant, expectorant, cardio tonic, anti-inflammatory and
58 useful in the treatment of biliousness, cough, burning sensation, thirst and
59 inflammation, wounds, ulcers, and rheumatism; it also has the ability to treat skin
60 diseases and leprosy. The plant has strong therapeutic potential thus occasionally
61 cultivated for medicinal purposes in the alternative medicine practice Ayurveda.
62 The leaves of H.benghalensis (L.) Kurz are used in treating skin diseases in
63 Burma and the bark is used to heal wounds in Indonesia. In India, H.benghalensis
64 (L.) Kurz is widely used to treat cough, asthma, leprosy and also to quench thirst.
65 According to some researches the therapeutic actions of this plant may be due to
66 the presence of mangiferin, which is known to be anti-inflammatory,
67 hepatoprotective, antioxidant, and antimicrobial. So far no information is available
68 for the analgesic and anti-inflammatory activity of the ethanol extract of so, the
69 present study has been undertaken to evaluate the analgesic and anti-
70 inflammatory activity of the ethanol extract of H.Benghalensis using hot plate
71 method in Swiss albino mice and Carrageenan-induced Hind Paw Edema
72 methods in long evans rat respectively.

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2. Materials and Methods:

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Hot Plate (Model – 35100, UGO BASILE, ITALY), Balance, Refrigerator, Beakers,

79

Petri dishes & glass wrought, Safety rat handling gloves, Mortar & pestle.,

80

Hypodermic , Syringes, Holder & test tube, Hot water Bath , Plethysmometer,

81

Acetic Acid , Carrageenan

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2.1. Medicinal plants (extracts)

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Extract were examined in two concentrations of 500mg/kg and 250mg/kg body

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weight of animal

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2.2. Control & Positive Control

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2.2.1. Analgesic activity

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1. Control – Distilled water

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2. Positive control – Diclofenac sodium

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Administered dose – 50mg/kg body weight animal

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2.2.2. Anti-inflammatory activity

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1. Control –Distilled water

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2. Positive control – Diclofenac sodium

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Administered dose – 50mg/kg body weight animal

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2.3. Experimental animal

95

Swiss albino mice (male and female), weighing 20-30g bred in International

96

Centre for Diarrheal Diseases and Research, Bangladesh(ICDDR,B) and grown in

97

the Animal House of the Department of Pharmacy, North South University (NSU).

98

Long Evans rats (male and female), weighing 100-170g of either sex, bred in NSU

99

and ICDDR,B and grown in the animal house of the Department of Pharmacy

100 NSU. All the animals were acclimatized one week prior to the experiments .The
101 animals were housed under standard laboratory conditions (relative humidity 55-
102 65%, room temperature 25.0± 20C, and 12 hours light dark cycle). The animals
103 were fed with standard diet from ICDDR, B and had free access to filtered water
104 (2).

105 **2.4. Plant Extraction method**

106 **2.4.1. Collection**

107 The plant sample of Hiptage Bengalensis was collected from Ayurvedic
108 Institution 'Back to Nature' during 18.06.2012 in the form of leaf shavings. The
109 leaf of the plant was collected and washed with water several times.

110 **2.4.2. Drying and grinding**

111 The collected plant leaf was washed with water, separated from undesirable
112 materials or plant parts, partially dried by fan aeration and then fully dried in the
113 oven at below 40°C for 2 days. The fully dried leaves was then grinded to a
114 powdered form and stored in there refrigerator at +4°C for a few days.

115 **2.4.3. Cold extraction (Ethanol extraction)**

116 103gm of powered material was taken in a clean, flat bottomed glass container
117 and soaked in 500 ml of 80%ethanol, sealed and kept for a period of 2 days with
118 occasional shaking and stirring. It was then filtered first by cotton material and
119 twice through whatman filter paper to obtain a finer filtrate. The filtrate (Ethanol
120 extract) obtained was evaporated by Rotary evaporator (Eyela n 1000, Tokyo
121 Rikaki kai co.ltd, Rotary vacuum, Japan) at 4 to 5 rpm and at 65°C temperature.
122 The separated filtrate was found to be a precipitate of dark green color and the
123 gummy concentrate was designated as the crude ethanol extract of the leaves of

124 *Hiptage Benghalensis*. It was then dried in the freeze drier and preserved at +4°C
125 for two weeks.

128 **2.5. Phytochemical Analysis**

129 **2.5.1. Study Design**

130 Qualitative phytochemical tests for the identification of alkaloids, flavonoids,
131 steroids, gum and carbohydrates, reducing sugar, saponins, tannin and
132 terpenoids were carried out for the plant extract by the method described by
133 **Harborne and Sazada** (3,4) .The freshly prepared extract of *Hiptage*
134 *Benghalensis* was qualitatively tested for the presence of chemical constituents.
135 Phytochemical screening of the extract was performed using the following
136 reagents and chemicals: Alkaloids with Wagner reagent, flavonoids with the use
137 of conc HCl, tannins with 0.1% ferric chloride, and saponins with ability to produce
138 suds. Gum was tested using Molish reagents and concentrated sulfuric acid,
139 steroids with sulfuric acid, reducing sugar with the use α -naphthol and sulfuric acid
140 and terpenoids with chloroform and conc. HCl.

142 **2.6. Analgesic activity of *Hiptage Benghalensis***

143 **2.6.1. Study design (For both Hot-Plate and Writhing)**

144 Experimental animals were randomly selected and divided into four groups
145 denoted as group-I, group-II, group-III, group-IV consisting of 6 mice in each
146 group individual weighing was done to adjust individual doses. Here, distilled
147 water was given to group-I, 50 mg/kg Diclofenac sodium for group II, 250 mg/kg

148 for group III and 500mg/kg for group IV of the crude extract of *Hiptage*
149 *Benghalensis*.

151 **2.6.2. Mice Screening**

152 Young Swiss-albino mice aged 4-5 weeks, average weight 25-30 gm. were used
153 for this study. They were kept in standard environmental condition for one week
154 in the animal house of the Department of Pharmacy, North south University,
155 Bangladesh for adaptation after their purchase. The animals were provided with
156 standard laboratory food and tap water ad libitum and maintained at natural day
157 night cycle. Mice screening was performed before Hot plate test. In that
158 experiment mice with significant response action (Licking, Shaking and Jumping)
159 and response time (at the range of 0-20 seconds) were selected.

161 **2.6.3. Hot plate test method**

162 The hot-plate test employed for measurement of analgesic activity which was
163 previously described by *Lanthers et al., (1992)* and modified by *Mahomed and*
164 *Ojewole., (2004)*. A comparison of Hot plate test was made between positive
165 controls (Diclofenac Sodium), control and test sample given orally 30 minutes
166 after hot plate induction. Positive analgesic activity was shown when sample
167 animal gave longer number of stimuli than the control, or the sample. The
168 temperature of the metal surface of the hot plate was maintained at $55 \pm 0.2^{\circ}\text{C}$.
169 Latency to a discomfort reaction (licking, shaking or jumping) was determined
170 before and after drug administration. The cut-off time was fixed at 15s to avoid the
171 damage to the animal paw. The latency was recorded at 0, 30, 60, 120, 180, 240
172 min following oral administration of the agents. The prolongation of the sample

173 latency time compared with that of control was used for statistical comparison.

174 Each mouse was placed in the beaker (on the hot plate) in order to obtain its
175 response to electrical heat induced nociceptive pain stimulus. The time for each
176 mouse to lick its paws or jump out of the beaker was taken (reaction time). Each
177 mouse served as its own control (5,6,) before treatment, its reaction time was
178 taken once. The mean of these values on determination constituted initial reaction
179 time before treatment of the mouse. Each of the test mice were thereafter treated
180 with either distilled water, diclofenac sodium (50mg/kg of body wt.) and ethanol
181 extract at the doses of H.Benghalensis 250 mg/kg and 500 mg/kg body wt. orally.
182 Thirty min after treatment, the reaction time of each group mice were again
183 evaluated five times individually in one hour interval on this occasion. Percent
184 analgesic score was calculated as:

$$185 \quad (1)(PAS) = \frac{T_b - T_a}{T_b} \times 100$$

186 Where, T_b = Reaction time (in second) before drug administration

187 T_a = Reaction time (in seconds) after drug administration

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189 **2.6.4. Acetic acid induced writhing test in mice**

190 The analgesic activity of the samples was evaluated using acetic acid induced
191 writhing method in mice . In this method, acetic acid is administered intra-
192 peritoneally to the experimental animals to create pain sensation. As a positive
193 control, any standard NSAID drug can be used. In the present study Diclofenac
194 sodium was used to serve the purpose. The plant extract was administered orally
195 in two different doses (250 and 500 mg/kg body weight) to the Swiss Albino mice
196 after an overnight fast. Test samples and vehicle were administered orally 30
197 minutes prior to intra-peritoneal administration of 0.7% v/v acetic acid solution

198 (0.1ml/10g) but Diclofenac sodium was administered 15 minutes prior to acetic
199 acid injection. Then the animals were placed on an observation table. Each
200 mouse of all groups were observed individually for counting the number of
201 writhing they made in 15 minutes commencing just 5 minutes after the intra-
202 peritoneal administration of acetic acid solution. Full writhing was not always
203 accomplished by the animal, because sometimes the animals started to give
204 writhing but they did not complete it. This incomplete writhing was considered as
205 half-writhing. Accordingly, two half-writhing were taken as one full writhing. The
206 number of writhes in each treated group was compared to that of a control group
207 while Diclofenac sodium (50 mg/kg) was used as a reference substance (positive
208 control).

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210 **2.7. Anti-inflammatory Effect of *Hiptage Benghalensis***

211 Preparation of inflammatory agent Carrageenan was used as inflammatory agent
212 in this experiment. It was obtained from Jahangirnagar University. Carrageenan
213 powder was suspended in 5 ml saline to make 0.1% suspension and kept in water
214 bath for proper homogenization. The tube was kept in hot water ($50\pm 2^{\circ}\text{C}$)
215 containing beaker to prevent transformation into a jelly like compound.

216

217 **2.7.1. Carrageenan-induced Rat Hind Paw Edema test**

218 The ethanol extract of *Hiptage Benghalensis* on carrageenan induced
219 inflammation in rat paw was investigated by following the method of Winter et al
220 (1962) with minor modifications. Rats were randomly divided into four groups,
221 each consisting of six animals, of which group I was kept as control giving only
222 water .Group II was given carrageenan as inflammatory agent. Group III and

223 group IV were given the test sample at the dose of 250 and 500 mg/kg body
224 weight respectively. Half an hour after oral administration of the test materials,
225 0.1ml 0.1% carrageenan suspension was injected subcutaneously in left hind paw
226 of each animal leading to the formation of edema *in situ* (localized inflammation).
227 The volume of paw edema was measured at 1, 2, 3, 6, and 8 hours using water
228 plethysmometer after administration of carrageenan. The right hind paw served as
229 a reference non inflamed paw for comparison (7,8) the average percent increase
230 in paw volume with time was calculated and compared against the control group.
231 Percent inhibition was calculated using the formula:

232 **(2) % Inhibition of paw edema = [1 - (Vt / Vc)] X 100**

233 Where Vc and Vt represent average paw volume of control and treated animal
234 respectively.

236 **2.7.2. Statistical analysis**

237 All the results were expressed as Mean ± Standard deviation (SD). Data was
238 analyzed using one-way ANOVA followed by Dunnett's t-test. P values <0.05
239 were considered as statistically significant.

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3.Result

3.1. Phytochemical screening

Phytochemical screening of the ethanol extract of *H.Benghalensis* leaf and stem revealed the presence of various bioactive components such as tannins, flavonoids, saponins, gums, steroids, alkaloids, reducing sugar and terpenoids (1)
The result of phytochemical test has been summarized in the table below-

Table 1: Result of Phytochemical Screening of Plant Extract

<i>Hiptage Benghalensis</i> Extract	<i>Leaf & Stem</i>						
	<i>Tannins</i>	<i>Saponins</i>	<i>Flavinoids</i>	<i>Gums& Carbohydrates</i>	<i>Alkaloids</i>	<i>Reducing Sugars</i>	<i>Terpenoids</i>
<i>80% ethanol</i>	+++	++	++	+++	+++	+++	+++

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Treatment	0 Hr.	1 Hr.	2Hr	3 Hr.	6 Hr.	8 Hr.
Control	.71±.055	.88±.077	1.18±.007	1.46±.063	1.55±.066	1.62±.065
Standard	.65±.039	.85±.058	.99±.036	1.24±.046	1.02±.028***	.79±.020***
(250mg/kg) Hiptage	.67±.057	.99±.101	1.19±.077	1.28±.054	1.09±.053***	.92±.032***
(500mg/kg) Hiptage	.68±.031	1.02±.081	1.21±.056	1.29±.131	1.08±.038***	.89±.027***

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3.2. Anti-inflammatory Activity

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Table 2: Anti-inflammatory effect of ethanol extract of *Hiptage Bengalensis* on carrageenan induced rat paw inflammation

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Table 3: Percent inhibition of the standard and two different concentrations of the extract compared with their respective means at 0 hour

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Treatment	%inhibition 1 hour	%inhibition 2 hour	%inhibition 3 hour	%inhibition 6 hour	%inhibition 8 hour
Standard	29.97	51.68	89.91	56.27	20.49
<i>Hiptage</i> (250mg/kg)	48.61	77.08	91.07	63.54	36.61
<i>Hiptage</i> (500mg/kg)	48.74	77.19	88.88	57.89	29.53

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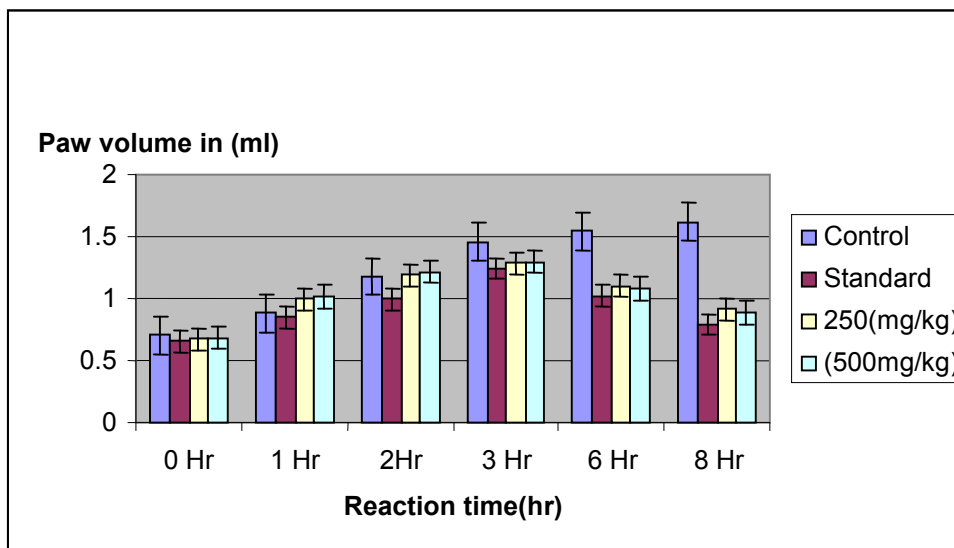


Figure 1: Anti-inflammatory activity of Hiptage by Paw edema method

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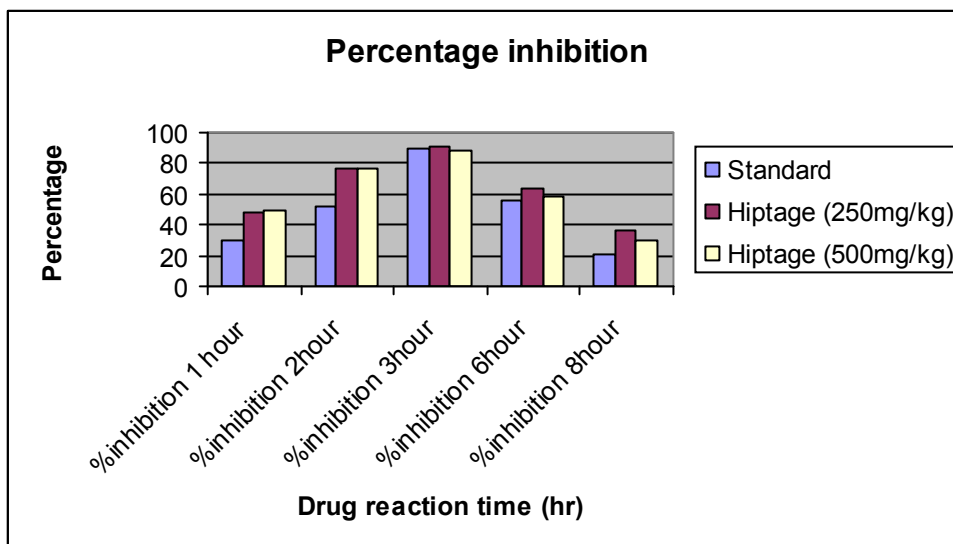


Figure 2: Percentage Inhibition of H.Benghalensis

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3.2.1. Effect of plant extract on Carrageenan-induced Hind Paw Edema

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The ethanol extract of H.Benghalensis exhibited statistically significant ($p < 0.05$) anti-inflammatory activity in Carrageenan-induced Hind Paw Edema of rat. This was determined by analyzing data using one way ANOVA followed by Dunnett's test. In control animals, the sub plantar injection of carrageenan produced a local edema that increased progressively to reach a maximal intensity four hours after

the injection of the phlogistic agent. Ethanol extract of *H.Benghalensis* showed a significant dose depended reduction at both 250 and 500mg/kg body weight. However significant inhibition of edema was found to be 63.54% and 57.89% at

Treatment	0 min	30 min	60 min	120 min	180 min	240 Hour
Control	10.70±.847	9.66±.937	8.00±.814	6.58±.641	5.52±.549	5.00±.443
Standard X	9.14± .524	11.02±1.00	12.60±.945	14.16±1.076***	15.96±.676***	12.48±.698***
Drug 250 mg/kg	7.68±.851	9.28±1.09	10.32±1.12**	11.28±1.07**	12.54±.912***	10.18±.747***
Drug 500 mg/kg ^h	7.65±.312	9.22±.285	10.34±.273	11.72±.233**	12.68±.177***	10.19±.163***

our of study at a dose of 250 and 500mg/kg body weight respectively. Further significant inhibition was to be 36.61% and 29.53% at eight hour of study at a dose of 250 and 500mg/kg body weight respectively.

3.3. Analgesic activity

Table 4: Analgesic effect of the ethanol extract of *H.Benghalensis* using the hot – plate method. Statistical evaluation of the results shown in table

Values in the results are expressed as mean ± SEM., ^a significantly different in comparison with control at P<0.05

Table 5: Percent inhibition of the standard and two different concentrations of the

extract compared with their respective means at 0 hour

Treatment group	% Inhibition				
	½ Hour	1 Hour	2 Hours	3 Hours	4 Hours
Standard	20.56	37.00	54.90	74.61	36.54
Hiptage 250 mg/kg	20.83	34.37	46.87	63.28	32.55
Hiptage 500 mg/kg	20.52	35.16	53.20	67.75	33.20

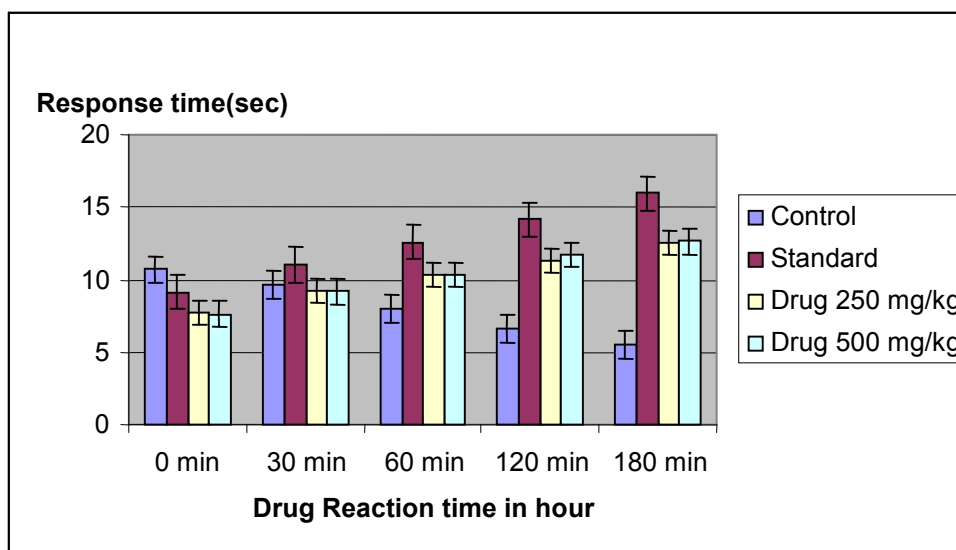
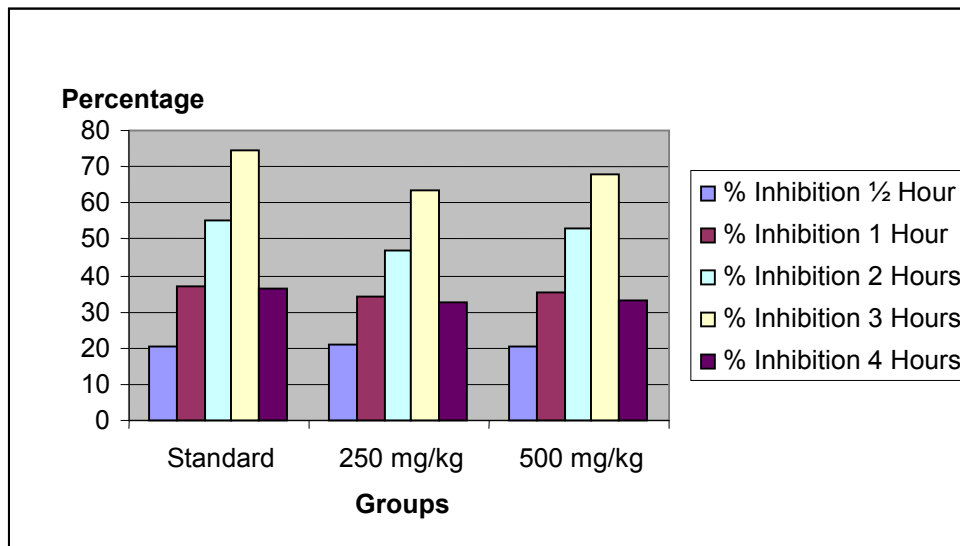


Figure 3: Analgesic activity of Hiptage by Hotplate method



± SEM	% Inhibition
00±4.11825	

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Figure 4: % Inhibition of Hiptage

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3.3.1. Effect of plant extract on Hot-Plate test

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The ethanol extract of *H. Bengalensis* exhibited statistically significant ($p > 0.05$) analgesic effect in hot plate test of white albino mice. This was determined by analyzing data using one way ANOVA followed by Dunnett's post hoc test. However, the data shows that the dose dependent effect reached 67.75% at 180 minutes and 63.28% at the 180 minutes at the doses of 500 and 250 mg/kg-body weight respectively.

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3.3.2. Effect of *Hiptage Bengalensis* extract in acetic acid induced writhing

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Table 6: Statistical evaluation of the results shown in table:

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392	Standard	14	18	16	17	13	15.6000±.92736***	50.00%
393	Hiptage 250 mg/kg	19	19	21	21	18	21.4000±1.96469** *	37.66%
394								
395	Hiptage 500 mg/kg	16	18	7	11	18	14.0000±2.16795** *	55.69%
396								
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400 Table 6 shows the effects of the extracts of *H.Benghalensis* on acetic acid-

401 induced writhing in mice. Both doses of the plant extract showed significant

402 reduction ($p<0.05$) of writhing induced by the acetic acid after oral administration

403 in a dose dependent manner. After oral administration of two different doses- 250

404 and 500 mg/kg body weight, the percent inhibition was 37.66% & 55.69%

405 *respectively*.



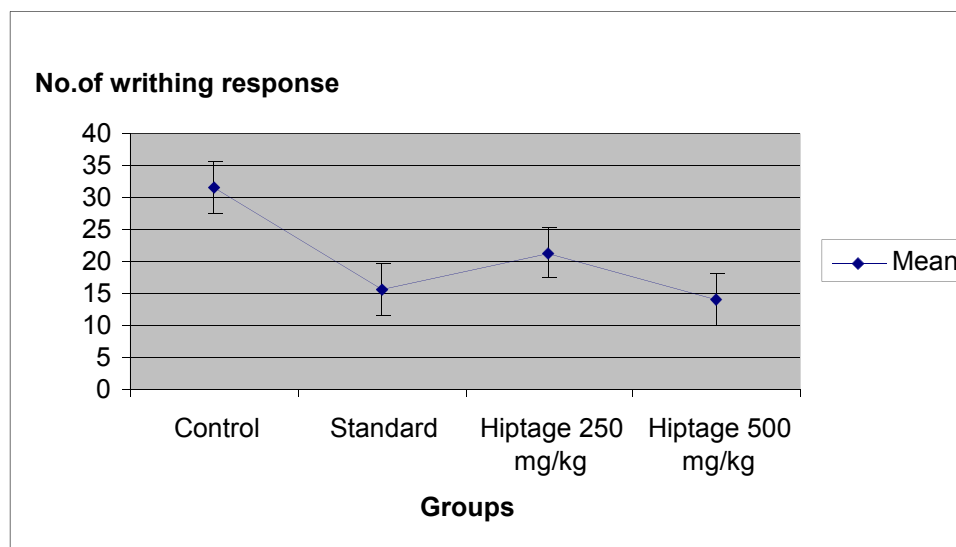
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Figure 7: Analgesic activity by acetic acid method(IP)



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Figure 8: Analgesic activity by acetic acid method(IP)

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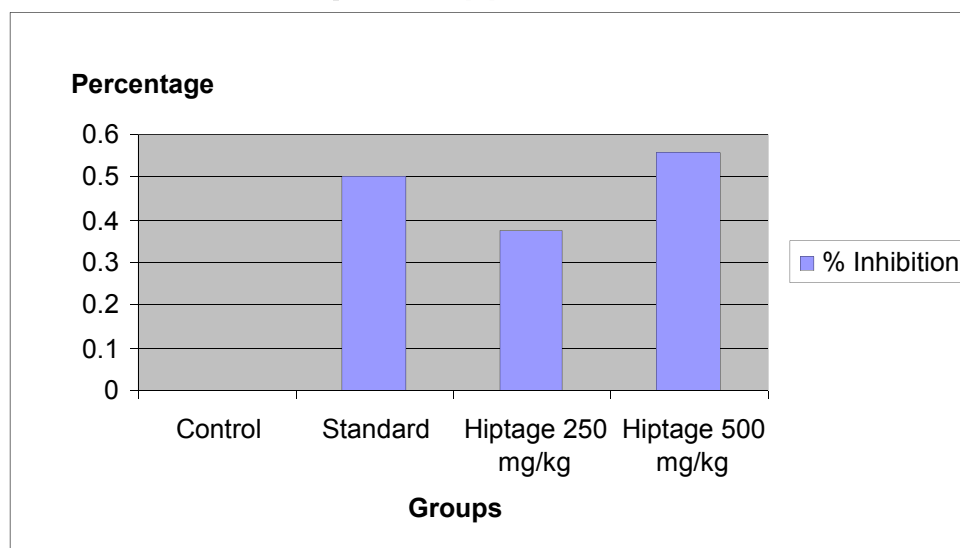


Figure 9: % Inhibition of Hiptage by acetic acid method(IP)

3.3.2. Acute toxicity

Oral administration of graded doses (250 & 500mg/kg) of the ethanol extract of *H. Bengalensis* to rats and mice did not produce any significant changes in behavior, breathing, cutaneous effects, sensory nervous system responses or gastrointestinal effects during the observation period. No mortality was recorded in any group after 24h of administering the extract to the animal.

4. Discussion

As a result of adverse side effects, like gastric lesions, caused by NSAIDs and tolerance and dependence induced by opiates, the use of these drugs as anti-inflammatory and analgesic agents have not been successful in all the cases. Therefore, new anti-inflammatory and analgesic drugs lacking those effects are being searched all over the world as alternatives to NSAIDs and opiates. During this process, the investigation of the efficacy of plant-based drugs used in the traditional medicine have been paid great attention because they are cheap, have

440 little side effects and according to WHO still about 80% of the world population
441 rely mainly on plant-based drugs(9).

442 Carrageenan induced paw edema is most widely use acute inflammatory model
443 for studying anti-inflammatory activity and it includes two phases. First phase
444 occurs within an hour of injection of phlogistic agent and is mediated through
445 release of histamine serotonin and kinin.While the second phase which can be
446 measured around 3 to 4 hours is related to release of
447 prostaglandins.(10)Carrageenan-induced edema involves the synthesis or release
448 of mediators at the injured site. These mediators cause pain and fever (11).
449 Inhibitions of these mediators from reaching the injured site or from bringing out
450 their pharmacological effects normally ameliorate the inflammation and other
451 symptoms. In the present study, it has been shown that the ethanol extract of the
452 *H.Benghalensis* possess a significant anti-edematogenic effect on paw edema
453 induced by carrageenan. Slight inhibition of inflammation is observed in first
454 phase and maximum in second phase, which is mainly due to release of
455 prostaglandins. The possible anti-inflammatory effect may be due ro inhibition of
456 cyclooxygenase enzyme which catalyzes the biosynthesis of prostaglandins and
457 thromboxane from arachidonic acid. These are reports that flavonoids possess
458 anti-inflammatory activity (12,13) and some act as phospholipase inhibitors
459 (14).Such inhibitors are able to decrease the inflammatory response to
460 Carrageenan in rats (15, 16)

461 Effect of ethanol extract of *Hiptage Benghalensis* in hot plate method is shown in
462 the figures. It is one of the most common test for evaluating the analgesic efficacy
463 of drugs/compounds. The paws of mice and rats are very sensitive to heat at
464 temperature which is not damaging to the skin. The responses are shaking,

465 jumping, withdrawal of the paws and licking of the paws. The time until this
466 response is prolonged after administration of centrally acting analgesics.
467 *H.Benghalensis* extract at the dose of 250 and 500 mg/kg showed the significant
468 ($P<0.05$) increase in latency time as compared to control. Positive control
469 Diclofenac Na showed significant ($P<0.05$) analgesic activity at the dose of 10
470 mg/kg. The analgesic activity was expressed as mean increase in latency after
471 drug administration \pm SEM. *H.Benghalensis* exhibited potent analgesic activity at
472 the dose levels of 250 and 500mg/kg. These extracts show analgesic activity at
473 low dose of 250mg/kg even in first hour in test. These results indicate that ethanol
474 extract of *H.Benghalensis* can produce significant analgesic effect.

475 The acetic acid-induced writhing is a sensitive method to evaluate peripherally
476 acting analgesics. Ethanol extract of *H.Benghalensis* possess analgesic effects in
477 the model of acetic acid-induced writhing test. Acetic acid induced writhing in mice
478 finds much attention in the screening of analgesic drugs in acetic acid-induced
479 abdominal writhing, the visceral pain model, and released arachidonic acid via
480 cyclooxygenase and prostaglandin biosynthesis which played a role in the
481 nociceptive mechanism. This model of response is thought to be mediated by
482 peritoneal mast cells acid sensing ion channels and the prostaglandin pathway. In
483 other words, the acetic acid induced writhing has been associated with increased
484 level of PGE₂ and PGF₂ α in peritoneal fluids as well as lipoxygenase products
485 (17). The increase in prostaglandin levels within the peritoneal cavity then
486 enhances inflammatory pain by increasing capillary permeability. Results of the
487 present studies show that ethanol extract of *H.Benghalensis* produced significant
488 analgesic effect which may be due to the inhibition of the synthesis of the
489 arachidonic acid metabolite.

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5. Conclusion

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The present study indicated that the ethanol extract of H. Bengalensis may have potential use in medicine. In our study, the ethanol extract of the plant showed significant dose dependent inhibition of paw edema and significant analgesic effect. Now our next aim is to isolate the leading compounds and to establish their chemical structure as well. Further studies should be undertaken to correlate the pharmacological activities with the chemical constituents of the leaf of H. Bengalensis and uncover specific mechanisms of action so that we may find a viable natural alternative to the traditional NSAIDs. Thus, it is concluded that the ethanol extract of fruit of Hiptage Bengalensis produce significant anti-inflammatory and analgesic activities in dose dependent manner.

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7. Conflict of Interest

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Authors have no Conflict of Interest

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