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Study of the hypoglycemic effect of *Tamarindus indica* Linn. seeds on non-diabetic and diabetic model rats.

Abbreviation:

- STZ = Streptozotocin
- BW = Body weight
- GOD-POD = Glucose-Oxidase and Peroxidase
- GI = Gastro intestine
- SPSS = Statistical Package for Social Sciences
- SEM = Standard error of mean
- SD = Standard Deviation
- OGTT = Oral glucose tolerance test
- DM = Diabetes Mellitus
- i.p = intraperitoneal

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53 **ABSTRACT**

54 Aim: The present study was undertaken to evaluate the antidiabetic effects of the *Tamarindus indica*
55 Linn seed in normal (non-diabetic), type-I and type-II model rats and to investigate their effect on
56 gastrointestinal motility and intestinal glucose absorption.

57 Methodology: *T. indica* seed powder was used at a dose of 1.25g/kg bw/10 ml water. Male Long-Evans
58 rats (160-210g body weight) were used for the experiment. Experiments were done in non-diabetic and
59 streptozotocin-induced diabetic model rats with a single feeding in different prandial states and blood
60 was collected. An intestinal perfusion technique was used to study the effects of *T. indica* seed powder
61 on intestinal glucose absorption in normal and type-II model rats. Gut motility was evaluated using
62 barium sulfate milk. Glucose was measured by GOD-POD method.

63 Result: The screening results showed that *T. indica* seed powder had no effect on fasting or
64 postprandial serum glucose level of normal and type-I diabetic rat. The seed powder also showed no
65 hypoglycemic effect in the fasting state and no antihyperglycemic effect in type-II model rats when fed
66 simultaneously with oral glucose load, but it exhibited significant antihyperglycemic effect when the seed

67 powder was fed 30 minutes prior to the glucose load at 105 minutes ($p < 0.03$). Glibenclamide
68 significantly lowered postprandial serum glucose levels of non-diabetic and type-II diabetic model rats
69 ($p < 0.02-0.001$). *T. indica* exerted inhibition on glucose absorption in type-II rats during the whole
70 perfusion period when compared with control. On the other hand, *T. indica* seed powder significantly
71 inhibited the gastrointestinal motility in Type-II rats.

72 Conclusion: The present data suggest that *T. indica* possesses antihyperglycemic properties in type-II
73 rats which are at least partly due to its inhibitory effect on intestinal glucose absorption. This effect
74 cannot be attributed to the acceleration of intestinal transit.

75

76 **KEY WORDS:** Anti-hyperglycemic, *Tamarindus indica*, streptozotocin, type-I diabetes, type-II
77 diabetes, gastro intestine.

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83 1. INTRODUCTION

84 Diabetes mellitus is a disease due to abnormality of carbohydrate metabolism and it is mainly linked
85 with low blood insulin level or insensitivity of target organs to insulin. It is the most prevalent chronic
86 disease in the world affecting a large population and its prevalence is about 6.8% [1].

87

88 Hyperglycemia and hyperlipidemia are two important characters of diabetes mellitus, an endocrine
89 disorder based disease. In modern medicine, no satisfactory effective therapy is still available to cure
90 diabetes mellitus [2]. Though pharmaceutical drugs like sulfonylureas and biguanides are used for the
91 treatment of diabetes but these are either too expensive or have undesirable side effects or
92 contraindications [3, 4]. In recent years, there has been renewed interest in plant medicine [5, 6, 7] for
93 the treatment against different diseases as herbal drugs are generally out of toxic effect [8, 9] reported
94 from research work conducted on experimental model animal. Although in human, whether there is
95 any toxic effect are not investigated but anti-diabetic effect of various plants having "folk medicine
96 reputation" have been screened [10].

97

98 *Tamarindus indica* Linn. (*family-* Caesalpinaceae [Fabaceae]), locally known as Tetul tree, is found
99 throughout the South Asian region and some portions of Africa. It is a large **ample**, evergreen tree, 12-
100 18 m high with round bushy crown and comparatively smaller bole. Phytochemical investigation
101 revealed the presence of many active constituents, such as phenolic compounds, cardiac glycosides,
102 L-(-)mallic acid, tartaric acid, the mucilage and pectin, arabinose, xylose, galactose, glucose, and
103 uronic acid [11, 12]. The fruit pulp contains large quantities (16-18%) of tartaric, citric, malic and
104 acetic acids, potassium tartrate, invert sugar, gum and pectin. It also contains traces of oxalic acid.
105 Seed testa contains a fixed oil. Seeds cotyledons contain albuminoids, fat and carbohydrates [13, 14].
106 Traditional healers claim that the seed of the plant possess antidiabetic properties. Scientific reports
107 also support the hypoglycemic activity of this plant [15, 16, 17, 18]. However, no published report
108 supports the underlying mechanism of the hypoglycemic effect of *T. indica* seed powder **in** normal,
109 STZ induced type I and type II diabetic model rats.

110

111 The purpose of this work was to explore the possible hypoglycemic / anti-hyperglycemic activity of *T.*
112 *indica* seed extract **in** type I and type II diabetic rats as well as to investigate the possible mode of
113 action beyond this activity.

114

115 **2. MATERIALS AND METHODS**

116 **2.1. Plant materials and preparation of test sample**

117 *Tamarindus indica* Linn. seeds were collected from Kushtia, a district of Bangladesh. The plant was
118 identified by the Bangladesh National Herbarium (voucher specimen no. DACB-38203). The seeds
119 were collected from the fresh fruit pulp and were washed carefully and dried for two days at 40°C in
120 an oven. Then the seeds were crushed in an electric grinder to make fine powder. Afterwards the
121 powder was stored immediately in the refrigerator at -20°C and **kept** in the same temperature up to
122 end of the experiment.

123

124 **2.2. Experimental animals**

125 The study was conducted on adult male Long-Evans rats (weighing 160-210g) bred at the BIRDEM
126 animal house maintained at a constant room temperature of 22±5°C, 40-70% humidity conditions and
127 the natural day-night cycle with an *ad libitum* access to food except the day of experimental procedure

128 when animals were used after 12hrs fasting. The rats had no access to food during the whole period
129 of blood sampling. The influence of circadian rhythms was avoided by starting all experiments at 8.30
130 a.m.

131

132 **2.3. Induction of diabetes in rats**

133 Diabetes stimulating type I was induced by a single intra-peritoneal injection of streptozotocin (STZ,
134 Upjohn Company, Kalamazoo, MI USA) dissolved in 0.1 M citrate buffer, pH 4.5, at a dose of 65mg/kg
135 body weight to adult rats [19]. On the 7th day rats (fasting blood glucose \geq 18mmol/l) were taken for
136 carrying out the experiments.

137

138 Type II diabetes was induced by a single intra-peritoneal injection of STZ at a dose of 90mg/ kg body
139 weight, in citrate buffer to the 48 hours old pups as described by Bonner *et al.* [19]. Experiments were
140 carried out 3 months after STZ injection an oral glucose (2.5g/kg bw) tolerance test was performed to
141 check the blood glucose levels of the rats. Rats having blood glucose level 8-11 mmol/l at fasting
142 condition were taken to carry out the experiments.

143

144 A total number of 250 rats were used to carry out the experiments, which include normal, type I and
145 type II diabetic model rats. The animals were divided into 3 groups: Gr 1- water control group; Gr 2-
146 glibenclamide treated positive control group; Gr-3 *T. indica* seed powder treated group. Number of
147 rats were 6-8 rats in each group.

148 **2.4. Acute effect on fasting and postprandial glucose level**

149 **2.4.1. Fasting condition**

150 The powder (1.25g/kg bw/10ml) was fed to overnight fasting (12hrs) rats and blood samples were
151 drawn at 0, 60 and 120 minutes [20]. The positive control group received glibenclamide (5mg/kgbw)
152 for normal and type-II rats and insulin (10 μ l/rat) for type-I rats whereas the control group received only
153 water (10ml/kg bw) [20]. Blood samples were collected by amputation of the tail tip under mild ethar
154 anesthesia. The rats were kept unfed throughout the experimental period.

155 **2.4.2. Postprandial condition**

156 The juice (10 ml/kg bw) and powder (1.25 g/kg bw), with or without glucose (2.5g/10ml/kg bw), were
157 fed to overnight fasted (12hrs) rats and blood samples were drawn at 0, 30, and 75 min when fed

158 simultaneously with glucose and at 0, 60 and 105 min when fed 30 min prior to glucose load. Both
159 positive control and water control rats were fed with glucose solution (2.5g/10ml/kg bw) and water
160 (10ml/kg bw) following glucose load [21].

161 **2.5. Effect of *T. indica* seed powder on intestinal glucose absorption**

162 An intestinal perfusion technique [22] was used to study the effects of *T.indica* seed powder on
163 intestinal absorption of glucose in nondiabetic and type 2 diabetic rats fasted for 36 hours and
164 anesthetized with sodium pentobarbital (50 mg/kg). The seed powder were added to a kreb's solution
165 (g/L 1.02 CaCl₂, 7.37 NaCl, 0.20 KCl, 0.065NaH₂PO₄.6H₂O, 0.6 NaHCO₃, pH 7.4), supplemented with
166 glucose (54.0 g/L) and perfused at a perfusion rate of 0.5 mL/min for 30 min through the duodenum.
167 The perfusate was collected from a catheter set at 40 cm. *T. indica* seed powder were added to kreb's
168 solution to a final conc. of 25 mg/mL so that the amount of seed powder in the perfused intestine is
169 equivalent to the dose of 1.25 g/kg. The control group was perfused only with kreb's buffer
170 supplemented with glucose. The results were expressed as percentage of absorbed glucose,
171 calculated from the amount of glucose in solution before and after the perfusion.

172 **2.6. Gastrointestinal (GI) motility test:**

173 Gastrointestinal motility was evaluated by using barium sulfate (BaSO₄) milk method [23].

174 The experiment was carried out by the method previously described by Chatterjee (1993). Distilled
175 water at a dose of 10ml/kg bw was fed with smooth metallic tube to control rats and *T indica* powder
176 (1.25g/kg bw/ 10ml) was fed to treated group. Baso₄ milk was prepared by adding Baso₄ as 10% w/v
177 in 0.5% CMC suspension. The milk was given to rats after 1 hour of administration of the test material.
178 The rats were sacrificed 15 minutes after the administration of the milk. Before sacrificing the rats were
179 anesthetized with di-ethyl ether. Then the abdominal part was opened and the intestinal part from
180 pylorus to the ileocecal junction as a total length of GI was taken into a petridish filled with distilled
181 water for washing. It was then soaked by tissue paper to make it dry and taken into a white paper
182 marked with 100 cm scale for measurement. The total gastrointestinal tract was measured first. Then
183 the length traversed by Baso₄ was measured (white color). This length traversed by Baso₄ was
184 expressed as percent of the total length of gastrointestinal tract and the result of the test group was
185 compared with that of control group (Chatterjee1993).

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188 **2.7. Biochemical analysis:**

189 Serum glucose was measured on the same day by glucose-oxidase method (GOD-POD) using a
190 commercial kit (Boehringer-Mannheim, GmbH, Germany) (Sera Pak, USA).

191

192 **2.8 Statistical analysis:**

193 Data from the experiments were analyzed using the Statistical Package for Social Sciences (SPSS)
194 software for Windows version 12 (SPSS Inc, Chicago Illinois, USA). Values were expressed as
195 mean±SD (Standard Deviation), Analysis of variance (ANOVA, Bonferroni Post Hock Test) and
196 independent sample 't'-test were done as the test of significance $p \leq 0.05$ was considered as the
197 minimal level of statistical significance.

198

199 **3. RESULTS**

200 **3.1. Acute effect of *Tamarindus indica* seed powder on blood glucose level of normal (non-**
201 **diabetic), type I and type II diabetic model rats**

202 Blood glucose level was analyzed at the fasting level and the results showed that the seed powder
203 had no significant effect on the fasting state of normal rats but lowered the serum glucose level 2.43%
204 at 60 min and 7.68% at 120 min. As expected glibenclamide, the positive control, lowered serum
205 glucose level significantly both at 60 minutes ($p=0.000$) and at 120 minutes ($p=0.000$) compared with
206 water control and powder treated groups (Table 1).

207

208 **Table 1: Effect of *T. indica* seed powder on fasting serum glucose level (M±SD) of normal, type**
209 **I and type II diabetic model rats:**

Group	Min 0 (mmol/l)	Min 60 (mmol/l)	Min 120 (mmol/l)
Normal rat			
Water control(n =6)	8.05±0.51	7.63±0.49	6.99±0.73
Glibenclamide(n =6)	8.39±0.76	5.67±0.88*	4.83±0.48*
<i>T. indica</i> seed powder (n =8)	7.82±0.68	7.63±0.41	7.22±0.61
Type I diabetic model rat			

Water control (n = 6)	22.04±1.28	20.53±3.09	19.40±3.92
Insulin (n =6)	21.49±1.58	4.30±1.87*	2.32±0.37*
T. indica seed powder (n =6)	21.38±1.51	20.84±1.90	19.51±2.66
Type-II diabetic model rat			
Water control(n =6)	8.35±1.57	8.87±2.50	10.53±3.88
Glibenclamide(n =6)	8.55±1.55	8.01±1.91	7.42±1.56*
T. indica seed powder (n =8)	9.10±1.12	9.93±2.37	9.57±1.94

210

211 Results are expressed as **mean ±standard** deviation (M±SD). One-way ANOVA (Bonferroni test) was done for
 212 comparing the different group and for the test of significance. *P= 0.000 when compared when compared with
 213 water control; n=number of rats.

214

215 The oral glucose tolerance test (OGTT) was performed and the results showed that powder had
 216 glucose lowering effect but non-significantly and the glibenclamide treated group showed a significant
 217 fall in serum glucose level at 75 minutes (p=0.000) (Table 2).

218

219 **Table 2: Effect of T .indica seed powder on serum glucose level (M±SD) of normal, type I and**
 220 **type II diabetic model rats when seed powder was fed simultaneously with glucose load:**

Group	Min 0 (mmol/l)	Min 30 (mmol/l)	Min 75 (mmol/l)	iobv
Normal rat				
Water control(n = 7)	6.38±0.51	8.36±0.68	7.79±0.75	3.39±1.71
Glibenclamide(n = 6)	5.84±0.66	7.27±0.38	5.55±0.78*	1.14±2.18
T. indica seed powder (n = 7)	6.15±1.21	8.64±0.90	7.92±0.63	4.26±3.28
Type-I diabetic model rat				
Water control(n = 7)	20.90±3.33	26.90±2.62	25.25±3.79	10.35±7.06

Insulin(n = 6)	21.86±2.16	18.38±6.48*	9.15±4.77**	-16.19±9.95
T. indica seed Powder (n = 6)	20.88±1.80	28.67±2.56	25.63±3.23	12.53±6.35
Type-II diabetic model rat				
Water control(n = 6)	7.64±1.42	17.10±4.24	17.36±3.42	19.17±8.80
Glibenclamide(n = 6)	9.01±1.40	16.25±0.98	12.72±0.90*	10.95±3.01
T. indica seed powder (n = 7)	8.21±2.10	16.97±1.86	15.49±2.40	16.03±6.24

221 Results are expressed as mean ±standard deviation (M±SD). One-way ANOVA (Bonferroni test) was done for
 222 comparing the different group and for the test of significance. *P= 0.000 when compared when compared with
 223 water control; iobv =Sum of the increments over the basal value. n=number of rats.

224

225 When fed 30 min before to glucose load, it was found that glibenclamide treated group showed
 226 significant blood glucose lowering effect at 60 minutes (p = 0.001) and 105 minutes (p = 0.000)
 227 normal and type I diabetic model rats in comparison to water control and powder treated group (Table
 228 3). T. indica seed powder showed significant (p = 0.003) blood glucose lowering effect at 105 min. on
 229 type II diabetic model rats in Comparison to water control and glibenclamide treated group (Table 3;
 230 Fig. 1).

231

232 **Table 3: Effect of T. indica seed powder on serum glucose level (M±SD) of normal, type 1**
 233 **and type 2 diabetic model rats when seed powder was fed 30 minutes before to glucose load:**

Group	Min 0 (mmol/l)	Min 60 (mmol/l)	Min 105 (mmol/l)	iobv
Normal rat				
Water control (n =6)	6.64±1.09	9.05±0.61	8.51±0.70	3.27±2.86
Glibenclamide (n =6)	6.85±1.23	5.37±0.93*	3.83±0.48**	-4.49±1.90
T.indica seed Powder (n=8)	6.87±0.82	8.00±0.78	8.30±0.63	2.55±1.85
Type-I diabetic model rat				
Water control(n = 6)	24.26±2.03	31.71±3.62	30.78±1.20	9.97±6.66

Insulin(n = 7)	22.99±2.09	11.47±5.84 *	8.28±3.62*	-26.22±6.06
T. indica seed Powder (n = 6)	23.79±2.41	32.12±4.06	28.08±2.42	12.62±8.42
Type-II diabetic model rat				
Water control (n = 6)	7.96±1.79	19.65±4.36	19.30±4.27	23.01±9.99
Glibenclamide (n = 6)	9.05±1.78	18.99±5.37	16.09±4.68	19.98±8.91
T. indica seed Powder (n = 6)	9.59±1.72	17.10±2.31	14.65±2.82*	12.57±2.84*

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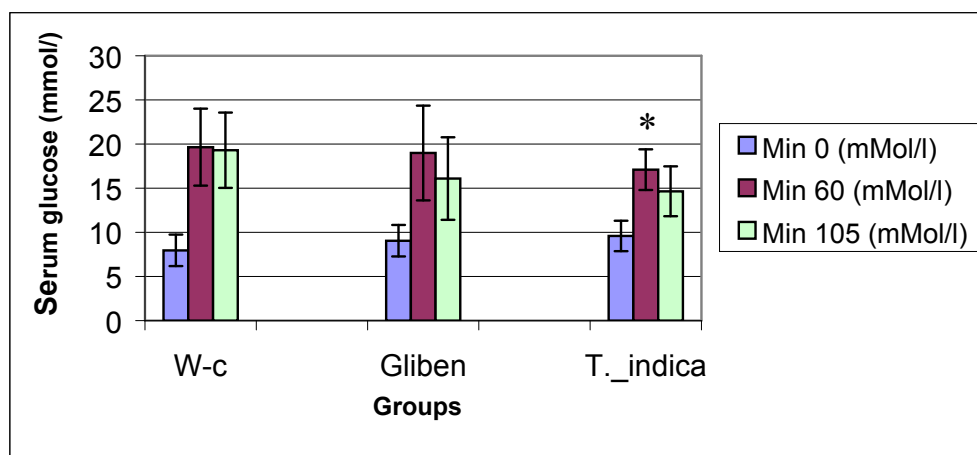
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Results are expressed as mean ±standard deviation (M±SD). One way ANOVA (Bonferroni test) was done for comparing the different group and for the test of significance. **P= 0.000.and. *p= 0.001.+ when compared when compared with water control; lobv =Sum of the increments over the basal value. n=number of rats.



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Figure 1: Acute effect of T. indica seed powder on serum glucose level on Type II diabetic model rats at 105 min. when feed at 30 min. before to glucose load. W-c=Water control.

3.2. **Effect of Tamarindus indica seed powder on gastrointestinal motility**

The length of GI traversed by BaSO₄ milk in powder treated group was lower than the water treated group. The inhibition in motility was not statistically significant in treated group of normal rats (Table 4). There was decreased percentage of length traversed by BaSO₄ with seed powder on type II

248 model rats in comparison to water control group. *T. indica* seed powder showed the significant motility
 249 effect (p=0.02).

250

251 **Table 4: Effect of *Tamarindus indica* Linn seeds powder on gastrointestinal motility**
 252 **test by Baso₄ milk of normal rats and type-II diabetic model rats:**

253

Group	GI total length (cm)	Length traversed by BaSO ₄ (cm)	% of Length traversed by BaSO ₄
Normal rat			
Control (n=6)			
	118.33±9.83	56.33±6.18	47.93±7.04
<i>T indica</i> Seed Powder (n=6)	123.83±6.11	47.67±7.86	40.77±9.77
Type-II diabetic model rat			
Control (n=5)			
	121.00±5.47	58.00±6.78	47.89±4.88
<i>T indica</i> Seed Powder (n=5)	121.00±9.61	48.00±8.09	39.46±4.31*

254

GI= Gastro Intestine. Data are presented as mean ±standard deviation (M±SD) and Group was

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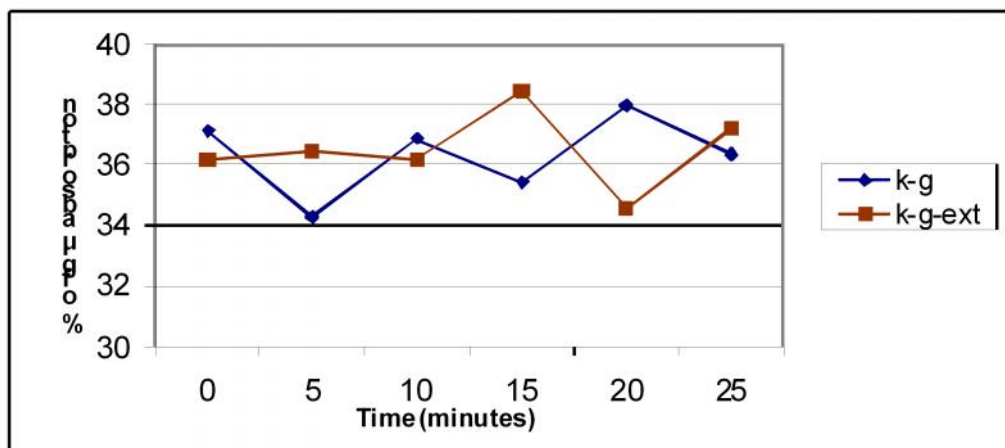
Compared by using independent samples't' test.*p =0.02. n=number of rats.

256

257 **3.3. Effect of *T indica* seed powder on upper intestinal glucose absorption**

258 It is evident that the upper intestinal glucose absorption was almost constant during 30 minutes of
 259 perfusion with glucose. When the seed powder solution was supplemented with the glucose solution, it
 260 showed no effect on intestinal glucose absorption in normal rats (Figure 2).

261



262

263

Figure 2: Effect of the *T. indica* seed powder on upper intestinal glucose absorption on

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normal rats.

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Results are presented as mean± SD (n=5). Rats were fasted for 36 hours and intestine was

266

perfused with glucose solution (54 g/l) with or without *T. Indica* seed powder (25mg/ml); glu=

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glucose, K-g= Kreb's solution, K-g-ext= Kreb's solution + glucose +seed powder juice.

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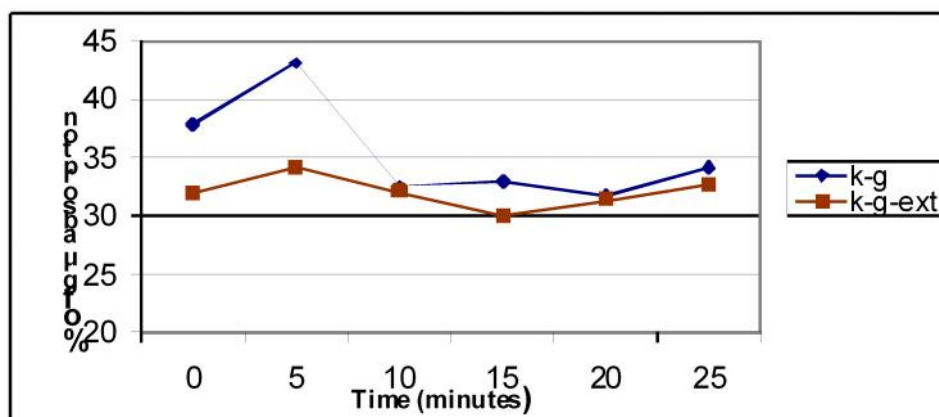
In case of type II model rats, intestinal glucose absorption was nearly constant during the 30 min of

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perfusion with glucose. There was a decrease in glucose absorption with glucose solution when

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supplemented with seed powder solution (Figure 3).



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Figure 3: Effect of *T. indica* seed powder on upper intestinal glucose absorption on type II

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diabetic rats.

275

Results are presented as mean± SD (n=6). Rats were fasted for 36 hours and intestine was perfused with

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glucose solution (54 g/l) with or without *T. Indica* seed powder (25mg/ml); glu= glucose, K-g= Kreb's

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solution, K-g-ext= Kreb's solution + glucose + Extract.

278

279 It denotes that *T indica* seed powder strongly affected the amount of absorbed glucose throughout the
280 notable period of experiment in type II rats. Figure 3 depicts the gradual fall in glucose absorption
281 during the whole perfusion period in type II rats compared to Krebs solution. Therefore, the obtained
282 results suggest that seed powder of the *T. indica* delays glucose absorption in the upper part of the
283 gastrointestinal tract.

284 .

285 **4. DISCUSSION**

286 Synthetic drugs such as Sulphonylureas and Biguanides are valuable in the treatment of DM, but their
287 uses are restricted by their limited action, pharmacokinetic properties, secondary failure rates and
288 accompanying side effects [24]. Moreover, these therapies only partially compensate for metabolic
289 derangements seen in diabetes and do not necessarily correct the fundamental biochemical lesion
290 [25]. As the incidence of diabetes is rising relatively around the world, there is an urgent need to
291 expand the range of effective palliatives available to patients.

292

293 The present study has been undertaken to screen the anti-hyperglycemic activity of *Tamarindus*
294 *indica* seed powder in nondiabetic, type I and type II diabetic model rats. The experimental approach
295 that has been followed, in addition to screening the hypo-/antihyperglycemic activity, gives an
296 approximate idea about the mechanism of action of the plant by analyzing the model, prandial states
297 and timing of hypoglycemic effect activity. Moreover, the study also extended to explore the possible
298 mechanism of action by elucidating the effect of the plant on gut motility and intestinal glucose
299 absorption.

300

301 Our results demonstrate that *T. indica* seed powder had no effect in the fasting state of nondiabetic,
302 type I or type II rats. At the post prandial state when the seed powder was administered
303 simultaneously with oral glucose load, no significant reduction was noticed with a single feeding in any
304 group of rats. On the contrary, when *T. indica* seed powder was administered half an hour before oral
305 glucose load in type I rats, the seed powder caused a significant attenuation in the rise of blood
306 glucose at 105 minutes compared to the control groups (glucose $M \pm SD$, mmol/l. 14.65 ± 2.82 in the
307 treated group Vs 19.30 ± 4.27 in the control group, $p < 0.03$). The antihyperglycemic effect of *T. indica*
308 seed in STZ induced diabetic rats have been found by other investigators [17].

309

310 *T. indica* seed powder was effective in type II diabetic model rats when fed 30 minutes before glucose
311 load. This effect might be due to a systemic action, i.e. as a result of the stimulation of Beta cells and
312 improving the insulin secretory capacity or enhancement of insulin action by the extract. This effect
313 could not be confirmed by our study since serum insulin level after a single feeding was not
314 determined. It has been claimed that the chronic aqueous extract of *T indica* improves glycemic status
315 [17, 26].

316

317 The inhibition of intestinal glucose absorption may contribute to the reduction of postprandial glucose
318 level. One of the objectives of the present study was to investigate whether the hypoglycemic effect is
319 related to the inhibition of glucose absorption in the gut. This was investigated in gut perfusion
320 experiment where *T. indica* seed powder showed strong inhibition of glucose absorption. This result
321 strongly suggests that the antihyperglycemic effect of *T. indica* as previously reported [26, 27] may be
322 due to, at least in part to the retardation of glucose absorption in the small intestine. *T. indica* is rich in
323 pectin's and dietary fibers such as cellulose, xylose and mucilage [16] and the presence of such
324 substances in the powder of *T. indica* may be responsible for the observed effect. Similar results have
325 been reported by some other scientists [16, 27]. Moreover, *T. indica* seed powder also inhibited the
326 BaSO₄ induced gastrointestinal motility in Type 2 rats. This result suggests that the decrease of
327 glucose absorption by *T. indica* seed powder is not achieved by an enhanced intestinal motility.

328

329 **5. CONCLUSION**

330 Based on the results of this study, it may be concluded that *T. indica* seed powder possesses
331 significant antihyperglycemic activity in type II diabetic model rats but not in type I and this is partly
332 due to inhibition of intestinal glucose absorption.

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342 (IPICS), Uppsala, Sweden and International Foundation for Sciences (IFS), Stockholm, Sweden and
343 Diabetic Association of Bangladesh in conducting this study.

344

345 **COMPETING INTERESTS**

346 Authors have declared that no competing interests exist.

347

348 **AUTHOR'S CONTRIBUTION**

349 Anzana Parvin, designed the proposal and protocol, performed the experiments and analysis; Md.
350 Morshedul Alam designed the protocol and wrote the first draft of manuscript; Md. Anwarul Haque,
351 Amrita Bhoumik and Liaquate Ali managed the study analysis; Begum Rokeya designed the protocol,
352 performed statistical analysis, managed the experiment. All authors approved the manuscript.

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