

Effect of Ethanol Extract of *Coccinia grandis* Lin leaf on Glucose and Cholesterol lowering activity

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ABSTRACT

Aims: To investigate the effect of ethanol extract of *Coccinia grandis* Lin (Cucurbitaceae) leaf in glucose and cholesterol lowering activity in animal model.

Study design: Extraction, glucose and cholesterol lowering activity evaluation.

Place and Duration of Study: Department of Pharmacy, North South University, Dhaka between June 2012 and December 2012.

Methodology: Glucose and cholesterol lowering effect of the ethanol extract of *C. grandis* leaf was evaluated using the alloxan-induced diabetic rat and compared the activity with diabetic control and antidiabetic drug (Glibenclamide). Ethanol extract (25mg/kg) of *C. grandis* and Glibenclamide were administered to normal and experimental diabetic rats for the duration of 10 days.

Results: Phytochemical screening showed the presence of alkaloids, flavonoids, saponins, cardenolides and polyphenols in significant amounts. In the alloxan-induced diabetic rat model, *C. grandis* (25 mg/kg) significantly ($p < 0.05$) lowered fasting blood glucose levels. *C. grandis* extract (25 mg/kg) also produced significant ($p < 0.05$) total cholesterol lowering and HDL increasing ($p < 0.05$) effects. Surprisingly, body weight was increased significantly ($p < 0.05$) in the *C. grandis* treated diabetic group.

Conclusion: These results suggest that the ethanol extract of *C. grandis* leaf possesses significant glucose and cholesterol lowering activity in animal model, thus supporting the usage of the plant in traditional medicine as an anti-diabetic medication.

Keywords: Antidiabetic activity, Glibenclamide, *Coccinia grandis*.

1. INTRODUCTION

High glucose and cholesterol levels are two well-known independent major factors that accelerate the risk of cardiovascular disease [1, 2]. Intolerance of glucose metabolism is commonly observed in diabetic patient. This intolerance is closely associated with the cholesterol level in the body. Insulin action is also influenced by the cholesterol metabolism [3]. Extensive research work has been carried out over the years for exploring the involvement of biochemical markers and the management of cardiac disease and associated complications like diabetes.

31 A large number of medicinal plants have been studied over the years to discover new
32 chemical entities from plant source which possesses blood glucose [4,5] and cholesterol [6]
33 lowering actions. In this context, here *C. grandis* were investigated to discover its glucose
34 and cholesterol lowering activity in animal model. Phytochemicals produced by the plants
35 like *C. grandis* is used in the treatment of diabetes in Asian countries as Ayurvedic remedies.

36 *C. grandis* Linn is a medicinal plant under the family Cucurbitacea. *C. grandis* is also known
37 as its synonyms *Coccinia indica* and *Coccinia cordifolia* [5]. Phytochemical screening of *C.*
38 *grandis* reported the presence of triterpenoides [7], carotenoides [8], flavonoids [9], alkaloids
39 [10] and fatty acids [11].

40 It has been found that extract of *C. grandis* leaves showed glucose lowering activity in the
41 blood. In addition, decreased phagocyte activities of macro-phages in alloxan-induced
42 diabetic animal indicate its activity in reducing lipid peroxidation. Pectin of *C. indica* is
43 responsible for providing this hypoglycemic action in diabetic model [12]. Human trials [5,13]
44 confirm glucose lowering action [13]. In a double blind clinical trial, 61 healthy volunteers
45 were taken *C. grandis* (20 g leaves) and 61 healthy volunteers were taken placebo meals in
46 their dinner. All of the volunteers were maintained a 10-hour fasting period. The authors
47 reported that low postprandial blood glucose levels in the plant group than the control groups
48 in both 1 hour and 2 hour postprandial period [5]. The authors reported that *C. grandis*
49 reduces phosphorylase activity, increases liver glycogen and thus lower the blood glucose
50 level [12]. Moreover, glucose lowering effect can be achieved by inhibiting key
51 gluconeogenic enzyme, glucose-6-phosphatase in animal model [14].

52 Singh et al., in 2007 reported that polyphenols of *C. grandis* lowers the plasma lipid level
53 followed by a beneficial effect on HDL and its ratio with total cholesterol in dyslipidemic
54 hamster model [15]. The authors were also stated that the action of polyphenols was
55 mediated through peroxisome proliferators activated receptor- α (PPAR α) by catabolizing
56 triglycerides and improving HDLC/TC ratio for the maintenance of lipid-glucose homeostasis
57 in hamster model [15].

58 Aims of the present study

59 We aimed to explore the glucose, total cholesterol lowering and HDL improving property of
60 *C. grandis* and investigate the induced body weight changes. Alloxan is used in our current
61 study to induce diabetes in animals. In rodent model alloxan selectively destroys pancreatic
62 beta cell which in turn shut down the production of insulin. In the presence of thiols, alloxan
63 generates reactive oxygen species (ROS) which in turn initiate toxic action in the beta cell by
64 free radical formation. Alloxan was administered for 10 days to simulate diabetes in the
65 animals as this was supported by previous papers [16, 17].

66 Hypothesis

67 We hypothesized that *C. grandis* will be able to lower blood glucose and total cholesterol
68 level while improve HDL level in the plasma. The extract of *C. grandis* leave might have a
69 distinct mechanism to provide glucose and cholesterol lowering activity in animal model. We
70 also thought that *C. grandis* will not affect the total body weight.

71

72 **2. MATERIAL AND METHODS**

73

74 **2.1 Plant Material**

75 The leaves of *C. grandis* were collected in December, 2012 from Jessore city in Bangladesh
76 and authenticated by Bangladesh National Herbarium. The leaves were initially dried under
77 shade and grinded.

78 2.2 Preparation of extract

79 The powder *C. grandis* leaves (1 kg) were mixed with ethanol in a 250mL flask with mild
80 shaking. The flasks were closed with cotton plug and aluminum foil at 48 hours at room
81 temperature. The extract was filtered through Whatman filter paper (No.1), concentrated
82 using a rotary evaporator at low temperature (40-50°C). The extracts were preserved in
83 airtight containers and kept at 4°C until further use.

84 2.3 Phytochemical Screening

85 Qualitative phytochemical tests were conducted for the identification of alkaloids, flavonoids,
86 terpenoids, saponins, polyphenols and cardenolides in the *C. grandis* extract [18, 19, 20, 21].
87 *Test for alkaloids:* Dragendorff's test: Plant extract (2 ml) and dilute HCl (0.2 ml) were taken
88 together in a test tube. Then 1 ml of Dragendorff's reagent was added. Orange brown
89 precipitate in the test tube indicates the presence of alkaloids in the extract sample. *Test for*
90 *cardenolides:* At first, the extract was dissolved in pyridine. Then a few drops of 2% sodium
91 nitroprusside and a few drops of 20 % NaOH were added. A deep red color which faded to a
92 brownish yellow indicates the presence of cardenolides. *Test for flavonoids:* A few drops of
93 concentrated HCl were added to a small amount of extract solution. Immediate appearance
94 of a red color indicates the presence of flavonoids. *Test for saponins:* 1 ml solution of the
95 extract was diluted to 20 ml with distilled water and shaken in a graduated cylinder for 15
96 minutes. 1 cm layer of foam indicates the presence of saponins. *Test for polyphenols:*
97 Polyphenols was identified by using the method described in Singh et al. in 2007 [15]. *Test*
98 *for terpenoids:* Salkowski test: 5 ml of the extract solution was mixed in 2 ml of chloroform,
99 and concentrated sulfuric acid (3 ml) was carefully added to form a layer. A reddish brown
100 coloration of the interface was formed to show positive results for the presence of
101 terpenoids.

102 2.4 Test animals

103 Test animals were collected from International Cholera and Dysentery Disease Research, in
104 Bangladesh (icddr,b). Albino rats (wistar strain) of both sexes weighing 175 g (average) were
105 used for the study. Munasinghe et al. in 2011 were also recruited both gender in their study
106 [5]. They were individually housed in polypropylene cages in well-ventilated rooms, under
107 hygienic conditions. Feeding of animals was done ad libitum, along with drinking water and
108 maintained at natural day night cycle.

109 2.5 Induction of diabetes and treatment

110 The solution of Alloxan monohydrate (10 mg/ml) was prepared in ice-cold citrate buffer 0.1 M
111 pH 4.5 and kept in ice. Then the solution was administered intraperitoneally to the animals
112 within 5 minutes at a dose of 50 mg/kg body weight. Alloxan was chosen to induce diabetes
113 due to its availability and widely reported in previous research [17, 22]. After 48 hours of
114 administration, diabetic model rats having glycosuria and hyperglycemia were taken for the
115 experiment. Rats fasted over night were used for induction of diabetes. Rats were divided
116 into two sets; diabetic and non-diabetic. Group I (n=12) received normal diet and served as
117 normal control. Group II (n=12) consists of alloxan-induced rats receiving normal diet and
118 serving as diabetic control. Group III (n=12) consists of alloxan-induced rats receiving
119 Glibenclamide at 0.5 mg/kg body weight once a day orally for 10 days. Group IV (n=12)

120 consists of alloxan-induced rats receiving *C. grandis* Linn (25 mg/kg) once a day orally for 10
 121 days. Group V (n=12) consists of normal rats receiving *C. grandis* (25 mg/kg) once a day
 122 orally for 10 days. Blood samples were collected through the tail vein just prior to and on day
 123 10 after drug administration. The blood glucose level was measured using Glucometer. Total
 124 cholesterol and HDL were measured using kinetic enzymatic method [23] and Gordon D.J.
 125 (1989) method [24] for all the samples.

126 2.6 Statistical Analysis

127 The results of statistical analysis for animal experiment were expressed as mean \pm SEM and
 128 were evaluated by one-way ANOVA followed by Dunnet's multiple comparisons. The results
 129 obtained were compared with the vehicle control group. The $p < 0.05$ were considered to be
 130 statistically significant. All the statistical tests were carried out using SPSS statistical
 131 software.

132 3. RESULTS AND DISCUSSION

133
 134 Phytochemical screening of the extract of *C. grandis* revealed the presence of various
 135 bioactive components of which alkaloid, cardenolides, flavonoids and polyphenols were the
 136 most prominent (Table 1).

137 **Table 1:** Phytochemical investigation of ethanol extract of *C. grandis* leaf.

138

Test	Alkaloids	Cardenolides	Flavonoids	Terpenoids	Saponins	Polyphenols
Observation	+	++	+++	+	+++	+

139 '+++'= indicates presence in high concentration; '++'= indicates presence in moderate
 140 concentration; '+'= indicates presence in trace concentration;

141

142 Blood glucose levels were measured for all the animals. The blood glucose and total
 143 Cholesterol level at day 10 in control (group I), alloxan-induced diabetic animal (group II),
 144 glibenclamide treated diabetic group (group III), *C. grandis* extract treated diabetic (group IV)
 145 and *C. grandis* extract treated normal group (group V) is summarized in Table 2. The
 146 differences were significant between control and diabetic control, diabetic control and
 147 Glibenclamide treated diabetic group, diabetic control and *C. grandis* treated diabetic group.
 148 Total Cholesterol levels were also determined for all animals.

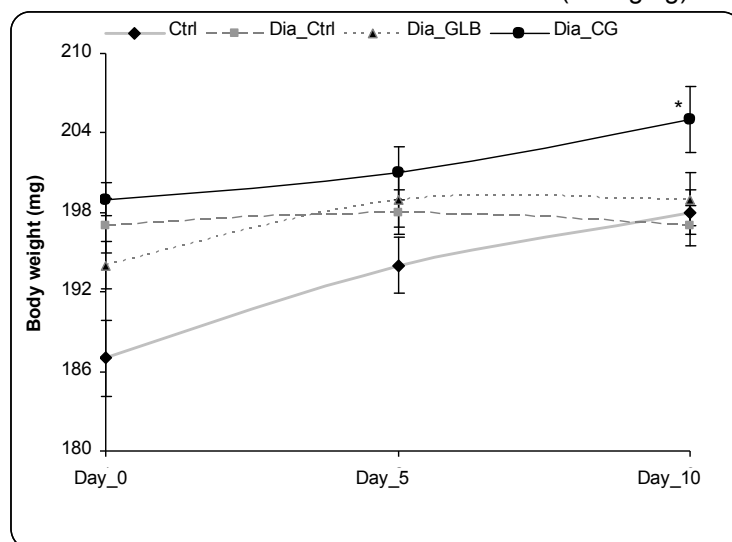
149 **Table 2.** Glucose and cholesterol level in serum between control and experimental animal
 150 groups.

Parameter (mg/dl)	Group-I: Control (n=12)	Group-II: Diabetic Control (n=12)	Group-III: Diabetic + GLB (0.5 mg/kg) (n=12)	Group-IV: Diabetic + CG treated (25 mg/kg) (n=12)	Group-V: CG treated (25 mg/kg) (n=12)
Glucose	119.6 \pm 1.4	248.1 \pm 1.6***	101.11 \pm 1.7**	108.42 \pm 1.2*	107.06 \pm 2.8*
Total Cholesterol	124.7 \pm 1.5	238.1 \pm 1.2***	106.08 \pm 1.8**	111.78 \pm 11.2*	112.63 \pm 4.5*
HDL	42.23 \pm 2.7	37.51 \pm 2.87*	38.81 \pm 5.91*	49.67 \pm 5.87**	48.81 \pm 3.32*

151 GLB= Glibenclamide; CG= *C. grandis* extract; Data were represented as the mean \pm SEM.
152 Data were analyzed by one way ANOVA followed by Dunnet's multiple comparison. The
153 criterion for statistical significance was *** $p < 0.001$, ** $p < 0.01$ and * $p < 0.05$.
154

155 The glucose level was significantly ($p < .05$) higher in alloxan-induced diabetic group (group
156 II) than *C. grandis* treated diabetic group (group IV). The total cholesterol level was
157 significantly ($p < .05$) lower in *C. grandis* treated diabetic group (group IV) than alloxan-
158 induced diabetic group (group II) while HDL level was higher ($p < .05$) in *C. grandis* treated
159 diabetic group (group IV) than alloxan-induced diabetic group (group II).
160

161 **Figure 1.** Effect of *C. Grandis* extracts on the body weight of diabetic rats. Ctrl: Control;
162 Dia_Ctrl: Diabetic Control; Dia_GLB: Diabetic animal treated with Glibenclamide; Dia_CG:
163 Diabetic animal treated with *C. Grandis* extract (25 mg/kg).



164 Data were represented as the mean \pm SEM. Data were analyzed by one way ANOVA
165 followed by Dunnet's multiple comparison. The criterion for statistical significance was $p < .05$
166 which is shown in Diabetic + *C. grandis* group on day 10.
167
168

169 Total body weights were also measured for all animals on day zero (before administration of
170 extract), day 5 and day 10. Surprisingly the body weight was raised in the alloxan-induced
171 diabetic animals that are treated with *C. grandis* leaf extract (group IV). Average body
172 weights of other animal groups were remaining unchanged (Figure 1).

173

174 The result of this study shows that ethanol extract of *C. grandis* leaf lowers serum glucose
175 and cholesterol level in the alloxan-induced diabetic rats while improving HDL level in the
176 serum. One unanticipated finding was that body weight was increased in *C. grandis* treated
177 diabetic control group.

178 As expected blood glucose lowering effect was found in this study. Even, the lowering of
179 blood glucose levels was consistent over long term treatment with ethanol extract of *C.*
180 *grandis*. This result indicates that the extract was able to improve blood glucose tolerance.
181 There are so many mechanisms are involved in blood glucose lowering activity. Present
182 study have not specifically designed to explore specific pathway involve in glucose lowering
183 activity. However, previous research studies have reported the following mechanisms of *C.*

184 *grandis* by which it may achieve blood glucose lowering activity. In 1979, Azad et al.
185 published a paper in which they studied *C. indica* in the treatment of diabetes. They
186 demonstrated that *C. indica* reduces phosphorylase activity, increases liver glycogen and
187 thus lower the blood glucose level [12]. In 1992, Hossain and coworkers published a paper
188 in which they studied hypoglycemic effects of *C. indica*. They demonstrated that *C. indica*
189 may inhibit key gluconeogenic enzyme, glucose-6-phosphatase in animal model [14]. In
190 2007, Rahul et al. *reported* a new mechanism in their review study. They demonstrated that
191 *C. grandis* may diminish the phosphorylase activity by suppressing glucose 6-phosphatase
192 enzyme [25]. Hence, through the various mechanisms as reported in the previous published
193 paper, glycogen in the liver will not be converted into glucose. Moreover, inhibition of
194 phosphorylase activity may produce an opportunity to increased glycogen synthesis in the
195 liver. In result, glucose may enter into the liver and deposit there as glycogen form. The total
196 mechanisms will ultimately help to remove excess glucose from the blood.

197 In the present study, we found that *C. grandis* reduces total cholesterol level during the 10
198 days treatment period which was expected in the hypothesis. Moreover, phytochemical
199 screening showed the presence of polyphenol in *C. grandis* extract. Polyphenol has
200 cholesterol lowering activity. In this context, the present findings seem to be consistent with
201 earlier research [13, 14]. In 2007, Singh et al. isolate polyphenol from *C. grandis* leaf by
202 ethanol extract. The authors reported that polyphenol might be responsible for cholesterol-
203 lowering effects [15]. In 2011, Krishnakumari et al. studied the activity of methanol extract of
204 *C. grandis* on lipid profile in streptozotocin induced diabetic rats. The authors reported that
205 the treatment with methanol extract of *C. grandis* was able to reduce lipid profiles to the
206 normal level [26]. We also found that methanol extract of *C. grandis* was able to improve
207 HDL cholesterol level. This activity might be achieved due to the presence of polyphenol.
208 This compound may increase the HDL level in the blood.

209 It was surprising that the body weight of the diabetic animals those are treated with *C.*
210 *grandis* was increased. This weight enhancing effect was not found in glibenclamide group.
211 Excess deposition of fatty acids, conversion of glucose into fatty acid and other mechanisms
212 are might be responsible for this unwanted activity. Therefore, further research is needed to
213 explore the root cause of increased body weight.

214 **4. CONCLUSION**

215
216 This study indicates that the methanol extract of *C. grandis* possesses significant glucose
217 and total cholesterol lowering activity in the blood at 25mg/kg dose investigated on the
218 experimental laboratory animals. This could provide rationale for its traditional uses in the
219 management of diabetes and suggests for further investigation and isolation of biologically
220 active constituents responsible for the activity.
221
222

223 **COMPETING INTERESTS**

224

225 No competing interests exist.

226

227 **AUTHORS' CONTRIBUTIONS**

228

229 Author MMNU designed the study, wrote the protocol and wrote the first draft of the
230 manuscript. Author MSI performed the statistical analysis. Author MMAA and AR managed
231 the literature searches. Author MMAA finalize the manuscript. All authors read and approved
232 the final manuscript.

233

234 **CONSENT (WHERE EVER APPLICABLE)**

235

236 This section is not applicable in our paper.

237

238 **ETHICAL APPROVAL (WHERE EVER APPLICABLE)**

239

240 Experimental protocol was approved by Institutional Ethics Committee of the Department of
241 Pharmacy, North South University. Animals were handled in accordance with international
242 principles guiding the use and handling of experimental animals (United States National
243 Institutes for Health Publication, 1985).

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245 **REFERENCES**

246

- 247 1. Cohen HW, Hailpern SM, Alderman MH. Glucose-cholesterol interaction magnifies
248 coronary heart disease risk for hypertensive patients. *Hypertension*. 2004a;(43):
249 983-7.
- 250 2. Cohen HW, Sloop GD, Study P. Glucose interaction magnifies atherosclerotic risk
251 from cholesterol. *Atherosclerosis*. 2004b;172:115-20.
- 252 3. Hallikainen M, Toppinen L, Mykkanen H, Agren JJ, Laaksonen DE, Miettinen TA.,
253 Niskanen L, Poutanen KS, Gylling H. Interaction between cholesterol and glucose
254 metabolism during dietary carbohydrate modification in subjects with the metabolic
255 syndrome. *Am J Clin Nutr*. 2006;84:1385-92.
- 256 4. Bailey CJ, Day C. Traditional Plant Medicines as Treatments for Diabetes. *Diabetes*
257 *Care*. 1989;12(8):553-564
- 258 5. Munasinghe M, Abeysena C, Yaddehige IS, Vidanapathirana T, Piyumal KPB. Blood
259 Sugar Lowering Effect of *Coccinia grandis* (L.) J. Voigt: Path for a New Drug for
260 Diabetes Mellitus. *Exp Diabetes Res*. 2011; 978762.
- 261 6. Doornbos AM, Meynen EM, Duchateau GS, Van Der Knaap HC, Trautwein EA.
262 Intake occasion affects the serum cholesterol lowering of a plant sterol-enriched
263 single-dose yoghurt drink in mildly hypercholesterolaemic subjects. *Eur J Clin Nutr*.
264 2006;60:325-33.
- 265 7. Vaishnav MM, Jain P, Jogi SR, Gupta KR. Coccinioside-K, triterpenoid saponin from
266 *Coccinia indica*. *Orient. J. Chem*. 2001;17:465-468.
- 267 8. Barua AB, Goswami BC, Carotenoides of *Cephalandra indica* (*Coccinia indica*).
268 *Curr. Sci*. 1979;48:630-632.
- 269 9. Vaishnav MM, Gupta KR. Ombuin 3-O-arabinofuranoside from *Coccinia indica*.
270 *Fitoterapia* 1996;67:80.
- 271 10. Qudrat-i-Khuda M, Khaleque KA, Miah MA. Chemical investigations of *Cephalandra*
272 *indica*. I. Constituents of dry aerial parts. *Sci. Res. (Dacca)* 1965;2:27-31.
- 273 11. Siddiqui IA, Osman SM, Subbaram MR, Achaya KT. Fatty acid components of seed
274 fats from four plant families. *J. Oil Technol. Assoc. India*. 1973;5:8-9.

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314
12. Azad Khan AK, Akhtar S, Mahtab H. *Coccinia indica* in the treatment of patients with diabetes mellitus. *Bangladesh Med Res Counc Bull.* 1979;5:60-6.
 13. Kumar GP, Sudheesh S, Vijayalakshmi NR. Hypoglycaemic effect of *Coccinia indica*: mechanism of action. *Planta Med.* 1993;59:330-2.
 14. Hossain MZ, Shibib BA, Rahman R. Hypoglycemic effects of *Coccinia indica*: inhibition of key gluconeogenic enzyme, glucose-6-phosphatase. *Indian journal of experimental biology.* 1992;30(5):418-20.
 15. Singh GP, Gupta P, Rawat A, Puri BG, Maurya R. Antidyslipidemic activity of polyprenol from *Coccinia grandis* in high-fat diet-fed hamster model. *Phytomedicine: international journal of phytotherapy and phytopharmacology.* 2007;14(12):792-8.
 16. Moideen K, Sherief SH, Sengottuvelu S, Sivakumar T. Hepatoprotective and antioxidant activity of *Coccinia grandis* root extract against paracetamol induced hepatic oxidative stress in wistar albino rats. *International Journal of Research in Ayurveda & Pharmacy.* 2011;2(3):858-63.
 17. Doss A, Dhanabalan R. Anti-hyperglycaemic and Insulin Release Effects of *Coccinia grandis* (L.) Voigt Leaves in Normal and Alloxan Diabetic Rats. *Ethnobotanical Leaflets.* 2008;12:1172-75.
 18. Overtuff ML, Loose MDC. In vivo model system: the choice of the experimental model for the analysis of lipoproteins and atherosclerosis. *Curr. Opin. Lipidol.* 1992;3:179.
 19. Owens DR, Luzio SD, Coates PA. Insulin secretion and sensitivity in newly diagnosed NIDDM Caucasians in UK. *Diabetic Med.* 1996; 13;9Suppl 6:S19–S24.
 20. Harborne JB. *Phytochemical Methods: A guide to modern techniques of plant analysis.* 3rd ed. Chapman and Hall, London ISBN: 0341235727032, 1998;302.
 21. Sazada S, Verma A, Rather AA, Jabeen F, Meghvansi MK. Preliminary phytochemicals analysis of some important medicinal and aromatic plants. *Adv. in Biol. Res.* 2009;3:1883195.
 22. Soto C, Muriel P, Reyes JL. Pancreatic lipid peroxidation in alloxan-induced diabetes mellitus. *Archives of medical research.* 1994;25(4):377-80.
 23. Deeg R, Ziegenhorn J. Kinetic enzymic method for automated determination of total cholesterol in serum. *Clinical chemistry.* 1983;29(10):1798-802.
 24. Gordon DJ, Probstfield JL, Garrison RJ, Neaton JD, Castelli WP, Knoke JD. High-density lipoprotein cholesterol and cardiovascular disease. Four prospective American studies. *Circulation.* 1989;79(1):8-15.
 25. Gupta R, Bajpai KG, Johri S, Saxena AM. An Overview of Indian Novel Traditional Medicinal Plants with Anti-Diabetic Potentials. *African Journal of Traditional, Complementary and Alternative Medicines.* 2008; 5: 1–17.
 26. Krishnakumari SP, Bhuvaneswari, Rajeswari P. Ameliorative potential of *Coccinia grandis* extract on serum and liver marker enzymes and lipid profile in streptozotocin induced diabetic rats. *Ancient science of life.* 2011;31(1):26-30.