

***In vitro* antioxidant potential of *Momordica charantia* fruit extracts**

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*Running Title: Antioxidative effects of *Momordica charantia**

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ABSTRACT

This research investigated the antioxidant potential of *Momordica charantia* fruit extracts in ethanol and ethyl acetate. The extracts have been assessed for DPPH free radical scavenging effect, FeCl₃ reducing power and superoxide scavenging effect. In DPPH method IC₅₀ value of ascorbic acid, ethanol and ethyl acetate extract were found 2.19 µg/ml, 111.87 µg/ml and 157.03 µg/ml respectively. In power reducing method, IC₅₀ value of ascorbic acid ethanol and ethyl acetate extract were found 50 µg/ml, 931.63 µg/ml and 754.86 µg/ml respectively. In super oxide scavenging method, IC₅₀ value of curcumin, ethyl acetate and ethanol extract were found 29.51 µg/ml, 331.26 µg/ml and 489.77 µg/ml respectively. The results of all three *in vitro* antioxidant assays exhibited that *M. charantia* possess relatively moderate antioxidant property than standards. The data obtained in the *in vitro* models clearly establish the antioxidant potency of the fruits extracts.

Keywords: *Antioxidant, Ethanol, Ethyl acetate, Petroleum ether, Momordica charantia, DPPH.*

INTRODUCTION

Oxidative stress is among the major causative factors in induction of any chronic and degenerative diseases including atherosclerosis, ischemic heart disease, ageing, diabetes mellitus, cancer, immunosuppression, neurodegenerative diseases and others [1]. Oxidative process is one of the most important routes for producing free radicals in foods, drugs and even in living systems [2]. The most effective path to eliminate and diminish the action of such free radicals, which cause the oxidative stress, is antioxidative defense mechanisms. Antioxidants are those substances which possess free radical chain reaction breaking properties. Recently there has been an upsurge of interest in the therapeutic potential medicinal plants as antioxidants in reducing oxidative stress-induced tissue injury. Among the numerous naturally occurring antioxidants; ascorbic acid, carotenoids and phenolic compounds are more effective. They are known to inhibit lipid peroxidation,

1 scavenge free radicals and active oxygen species by propagating a reaction cycle and to chelate
2 heavy metal ions.

3 Fruits have many health beneficiary functions. Recent research has confirmed that consumption of
4 fruits and vegetables can reduce the risk of stroke and cancer [3, 4, 15, and 21] as well as
5 inflammation and problems caused by aging [1]. This risk reduction is related to the presence of
6 antioxidative agents in fruits. They fight free radicals by supplying them the electron they lack, and
7 thus neutralize them. There are various types of antioxidants that our body needs to operate
8 optimally. Different antioxidants scavenge different free radicals, some work directly, while others
9 work indirectly as catalysts to boost our own body's production of antioxidants. Therefore, we need a
10 multitude of vitamins, minerals and enzymes to operate proficiently, so we need a wide range of
11 antioxidants [12, 13].

12 *Momordica charantia* L. (Cucurbitaceae), locally known as tit korla, has important role as a source of
13 carbohydrates, proteins, vitamins, minerals and other nutrients in human diet, which are necessary for
14 maintaining proper health [18]. *M. charantia* fruit is also very important economic source of proteins,
15 minerals, and calories of vitamins, essential for human nutrition [19]. Researchers reported the
16 antihyperglycemic [8], anti-migratory [14], anti-proliferatory [6] effects of the different extract and
17 compounds of *M. charantia*. As a part of our continuous work on medicinally important plants, we
18 report here the antioxidative effects of ethanol and ethyl acetate extracts of *M. charantia* in reducing
19 power model, DPPH free radical scavenging model and superoxide scavenging model.

20

21 **MATERIALS AND METHODS**

22 **Chemicals and reagents**

23 All chemicals used were of analytical grade. Ascorbic acid, nitro blue tetrazolium (NBT), trichloroacetic
24 acid, 2, 2-diphenyl-1-picryl hydrazyl free radical (DPPH) were purchased from Sigma-Aldrich,
25 (Germany). Ethyl acetate (98%) and absolute ethanol (99.5%) were also procured from Sigma (India).

26 **Collection of plant material**

27 *M. charantia* fruits were collected from Chittagong region. Foreign materials of the fruits were
28 removed, dried in the sunlight for four consecutive days and crushed into fine powder. The powder
29 was dried at 40°C for 4 h by electric oven.

30 **Preparation of extract**

31 The powder of dried fruit was soaked in ethanol and ethyl acetate in separate conical flask for 12 days
32 with 3 days interval at room temperature ($28 \pm 2^\circ\text{C}$) with occasional shaking and stirring. The conical
33 flasks were sealed to avoid evaporation. After that the contents were filtered and the filtrate was
34 evaporated to dryness with rotary evaporator (RE 200, Bibby sterling, UK) under reduced pressure at
35 45°C. The blackish green crude extract was preserved at 4°C until further use.

36 **Antioxidative assay by *in vitro* methods**

37 **Free radical scavenging activity assay**

38 The antioxidative effect of the fruit of *M. charantia* ethanol and ethyl acetate extract was assessed by
39 the established method of Brand-William *et al.*, 1995 [5] with slight modifications. Briefly, the extracts
40 (20, 40, 60, 80, 100, 200, 400, 800 µg/ml) were prepared in ethanol and ethyl acetate. Positive control

1 ascorbic acid solution was made with the concentration between 1-100 µg/ml. DPPH solution
2 (0.004%) was prepared in ethanol and 5 ml of this solution was mixed with the same volume of extract
3 and standard solution separately. These solution mixtures were kept in dark for 30 min to read
4 absorbance at 517 nm using a spectrophotometer (UV-1601 Shimadzu, Kyoto, Japan). The degree of
5 DPPH purple decolorization to DPPH yellow indicated the scavenging efficiency of the extract. Lower
6 absorbance of the reaction mixture indicated higher free radical scavenging activity. The scavenging
7 activity against DPPH was calculated using the equation.

8 Percent of scavenging activity = $[(A-B)/A] \times 100$, where, A was the absorbance of control (DPPH
9 solution without the sample), B was the absorbance of DPPH solution in the presence of the sample
10 (extract/ascorbic acid). The control (ascorbic acid) was conducted in the same manner, except that
11 distilled water was added instead of sample.

12

13 **Reducing power assay**

14 The reducing power of the fruit extracts was determined according to Oyaizu ,1986 [17]. A 1.0 ml of
15 extract solution (100, 500, 1000, 2000, 5000 µg/ml) was mixed with 2.5 ml phosphate buffer (0.2 M,
16 pH 6.6) and 2.5 ml potassium ferricyanide (1%, w/w), and then mixture was incubated at 50°C for 20
17 min. After incubation at 50°C for 20 min, the solutions were mixed with 2.5 ml of 10% (w/w) trichloro-
18 acetic acid and then centrifugation at 3000 rpm for 10 min. The supernatant (2.5 ml) was mixed with
19 2.5 ml of distilled water and 0.5 ml of 0.1% ferric chloride. The resulting solution was read at 700 nm.
20 Increased absorbance of the reaction mixture indicated increasing reducing power.

21 **Super oxide scavenging activity assay**

22 In this method, superoxide radical is generated by the addition of sodium hydroxide to air saturated
23 dimethyl sulfoxide (DMSO). The generated superoxide remains stable in solution, which reduces nitro
24 blue tetrazolium (NBT) into formazan dye at room temperature and that can be measured at 560 nm.
25 Briefly, to the reaction mixture containing 1 ml of alkaline DMSO (1 ml DMSO containing 5 mM NaOH
26 in 0.1 ml water) and 0.3 ml of the extract in freshly distilled DMSO at various concentrations, added
27 0.1 ml of NBT (1 mg/ml) to give a final volume of 1.4 ml. The absorbance was measured at 560 nm.
28 The percentage of super oxide radical scavenging by the extracts and standard compounds were
29 calculated as follows:

30
$$\% \text{ superoxide scavenging activity} = \frac{\text{Test absorbance} - \text{control}}{\text{Test absorbance}} \times 100$$

31

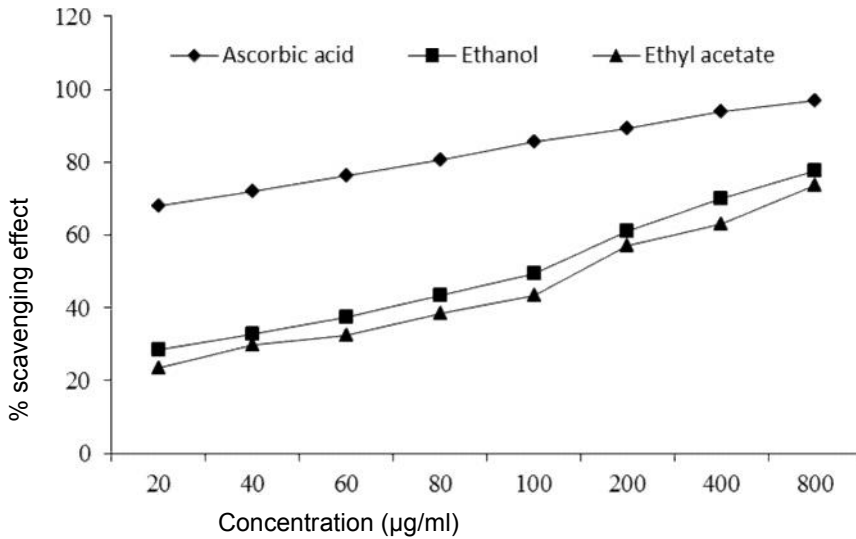
32 **RESULT**

33 **DPPH radical scavenging assay**

34 The radical scavenging effect of ethanol and ethyl acetate extract was summarized in Figure 1.
35 Results showed that both the extracts showed a dose dependent radical scavenging effect.

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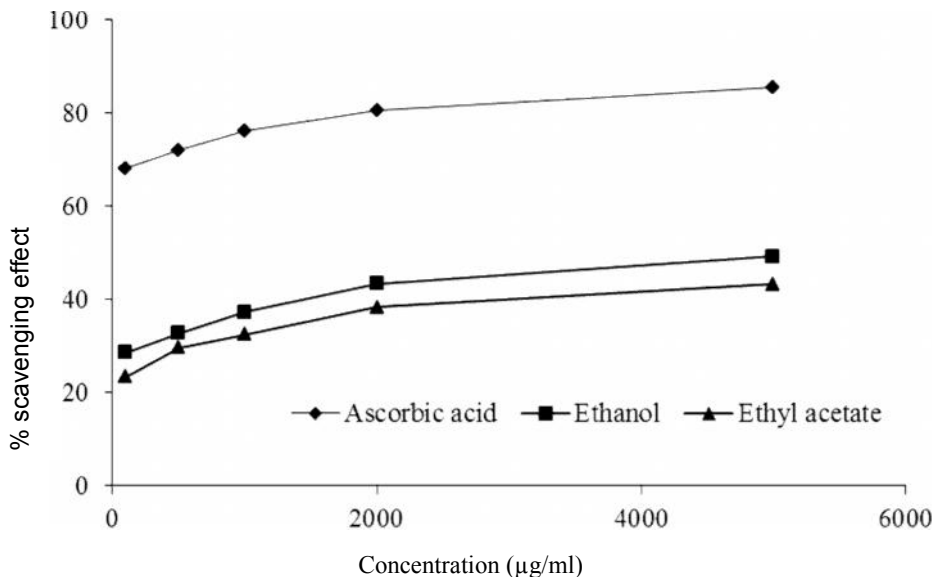
2 Figure 1: Relative percentage of scavenging activity for standard and *M. charantia* extracts by DPPH
3 method

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5 The most prominent scavenging effect of ethanol and ethyl acetate extracts were 77.65% and 73.75%
6 at a concentration of 800 µg/ml which were comparable to the highest activity (96.86) of ascorbic acid.
7 The inhibition concentration (IC₅₀) of the extract was determined by plotting a graph of scavenging
8 activity against the log concentration. The IC₅₀ value of ascorbic acid, ethanol and ethyl acetate
9 extracts was found 2.19 µg/ml, 111.87 µg/ml and 157.03 µg/ml respectively (Table 1).

10 **Reducing power by FeCl₃**

11 Results showed that the reducing power of the extracts increased with the concentrations. The
12 extracts showed a dose dependent effect in reducing power measurement. Ethanol and ethyl acetate
13 extracts showed the highest reducing power 70.51% and 70.83%, respectively, which were higher
14 than that of ascorbic acid (34.91%) (Figure 2).



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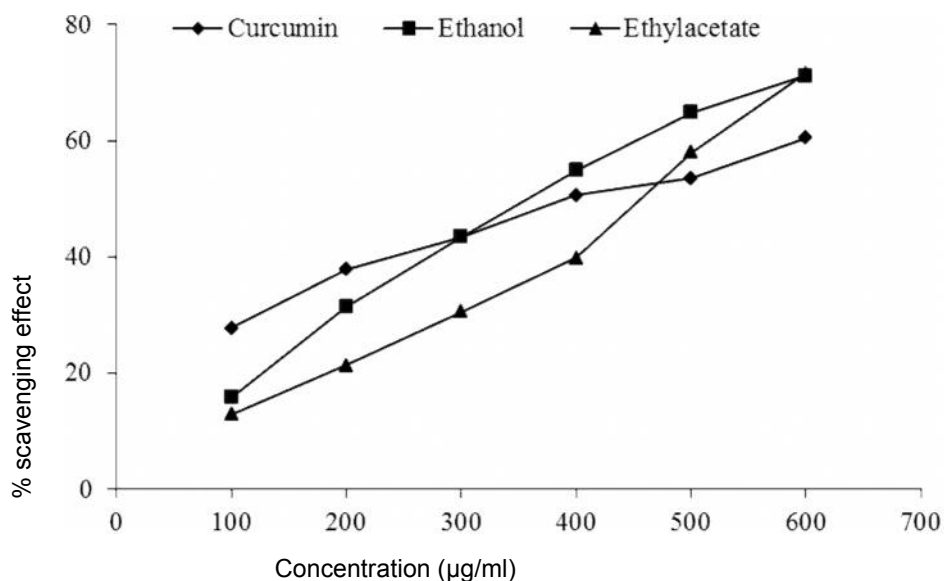
1 Figure 2. Relative percentage of scavenging activity for two solvents by reducing power method
2
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4 The percentage (%) of reducing power or % of inhibition was plotted against log concentration and
5 from the graph IC₅₀ value was calculated by linear regression analysis. IC₅₀ value of ascorbic acid,
6 ethanol and ethyl acetate extract were found 50 µg/ml, 931.63 µg/ml and 754.86 µg/ml, respectively
7 (Table 1).

8 Super oxide scavenging activity by alkaline DMSO method

9 Super oxide free radical was formed by alkaline DMSO which reacted with NBT to produce
10 colored diformazan. The ethanol and ethyl acetate displayed a dose dependent activity in inhibiting the
11 superoxide radicals. The best scavenging effect was shown 71.18% for ethanol and 71.58% for ethyl
12 acetate extract. These promising scavenging effects of ethanol and ethyl acetate extracts were
13 stronger than the reference agent curcumin (Figure 3).

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18 Figure 3 . Superoxide scavenging activity of *M. charantia*.

19

20 Scavenging activity (%) was plotted against log concentration and from the graph IC₅₀ was calculated
21 by linear regression analysis. IC₅₀ value of curcumin, *M. charantia* ethanol and ethyl acetate extract
22 was found 29.51, 489.77 µg/ml and 331.26 µg/ml, respectively (Table 1).

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1 Table 1. IC₅₀ Values of the extracts in different experimental models

Antioxidative models	Standard/ Samples	IC ₅₀ Values (µg/ml)
DPPH Free radical scavenging effect	Ascorbic acid	2.19
	Ethanol	111.87
	Ethyl acetate	157.03
Reducing effect	Ascorbic acid	50
	Ethanol	931.63
	Ethyl acetate	754.86
Superoxide scavenging effect	Curcumin	29.51
	Ethanol	489.77
	Ethyl acetate	331.26

2

3 DISCUSSION

4

5 Fruits contain large variety of antioxidants. Many methods are available to measure the antioxidative
 6 capacity of plant materials. Owing to the complexity of the oxidation-antioxidation process, no single
 7 testing method is capable of providing a comprehensive view of the antioxidative profile of a
 8 sample[15]. Therefore, a multi-method approach is necessary to assess antioxidative activity. In this
 9 study we used three different methods: DPPH free radical scavenging assay , ferric reducing power
 10 assay and superoxide scavenging assay.

11 However, plants of high antioxidative effects can be pivotal sources of such uses [10]. In the present
 12 study, the antioxidative activity, in terms of the scavenging of the radical DPPH of the ethanolic and
 13 ethyl acetate extracts of *M. charantia* was determined and compared with ascorbic acid, the reference
 14 antioxidative agent. 1,1-Diphenyl-2-picrylhydrazyl (DPPH) is a stable organic nitrogen radical. DPPH
 15 radical is commercially available. It is regarded as a model for a lipophilic radical. The assay of the
 16 scavenging of DPPH radical is widely used to evaluate the antioxidant capacity of extracts from
 17 different plant materials. The proton-radical scavenging action has been known as an important
 18 mechanism of antioxidation. DPPH was used to determine the proton-radical scavenging action of the
 19 extracts, since it possesses a proton free radical and shows a characteristic absorption at 517 nm.
 20 The purple color of the DPPH solution rapidly turned into yellow once it encounters proton-radical
 21 scavengers. The intensity of the radical scavenging effect is measured by the calculated half-inhibition

1 concentration (IC_{50}), the efficient concentration required for decreasing initial DPPH concentration by
2 50%. IC_{50} was obtained by interpolation from linear regression analysis of data shown the IC_{50} values
3 were 111.87 $\mu\text{g/ml}$ for ethanol extract was and 157.03 $\mu\text{g/ml}$ for ethyl acetate extract suggesting that
4 ethanol extract had the stronger antioxidative potential of the extracts. However, both the scavenging
5 effects were biologically important because the cutoff value for antioxidative power is 1000 $\mu\text{g/ml}$.
6 Extracts or chemical agents with the values higher than this are not effective as antioxidants.
7 Ascorbic acid is used as reference standard because ascorbic acid impairs the formation of free
8 radicals in the process of intracellular substance formation throughout the body.

9

10 Reducing power assay method is based on the principle that substances, which have reduction
11 potential, react with potassium ferricyanide (Fe^{3+}) to form potassium ferrocyanide (Fe^{2+}), which then
12 reacts with ferric chloride to form ferric ferrous complex that has an absorption maximum at 700 nm.

13 The reducing capacity of a compound may serve as a significant indicator of its potential antioxidant
14 activity. The reducing power of *M. charantia* ethanol and ethyl acetate extract along with that of
15 ascorbic acid at concentrations between 5-50 $\mu\text{g/ml}$ showed that high absorbance indicates high
16 reducing power. The reducing power of the plant extract was increased as the amount of extract
17 concentration increases. In our study, the reducing power of extract was lower than that of ascorbic
18 acid.

19

20 The scavenging activity of the extract against superoxide radical generated in NaOH-alkaline DMSO-
21 NBT system, resulting in the formation of the blue formazan was studied in this research. The
22 generated superoxide remains stable in solution, which reduces nitro blue tetrazolium into formazan
23 dye at room temperature. Superoxide scavenger capable of reacting inhibits the formation of a red
24 dye formazan. The inhibition of formazan formation by the extract was reflected through the IC_{50} value
25 for ethanol and ethyl acetate extract, 489.77 $\mu\text{g/ml}$ and 331.26 $\mu\text{g/ml}$ respectively , which was
26 significantly ($p < 0.05$) different compared to that of curcumin, 29.51 $\mu\text{g/ml}$. This finding demonstrates
27 that *M. charantia* fruit extract is capable of non-enzymatically inhibiting the superoxide radical,
28 produced in biological system, which is a precursor of many ROS and is shown to be harmful for
29 various cellular components. Although the enzyme superoxide dismutase possessed in aerobic and
30 anaerobic organisms can catalyze the breakdown of superoxide radical.

31 CONCLUSION

32 The results stated above showed that the ethanolic extract of *M. charantia* possessed noteworthy
33 antioxidative effects in all the models. Whatever the solvent for extraction, the antioxidative effect of
34 *M. charantia* evidenced that it could be a very good source of natural medicines on standard
35 formulation.

36

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5 **CONFLICT OF INTEREST**

6 The authors have declared that there is no conflict of competing interest.

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