

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34

**Study of the hypoglycemic effect of *Tamarindus indica* Linn. seeds on non-diabetic and diabetic model rats.**

**Abbreviation:**

- STZ = Streptozotocin
- BW = Body weight
- GOD-POD = Glucose-Oxidase and Peroxidase
- GI = Gastro intestine
- SPSS = Statistical Package for Social Sciences
- SEM = Standard error of mean
- SD = Standard Deviation
- OGTT = Oral glucose tolerance test
- DM = Diabetes Mellitus
- i.p = intraperitoneal

35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52

53 **ABSTRACT**

54 Aim: The present study was undertaken to evaluate the antidiabetic effects of the *Tamarindus indica*  
55 Linn seed in normal (non-diabetic), type-I and type-II model rats and to investigate their effect on  
56 gastrointestinal motility and intestinal glucose absorption.

57 Methodology: *T. indica* seed powder was used at a dose of 1.25g/kg bw/10 ml water. Male Long-Evans  
58 rats (160-210g body weight) were used for the experiment. Experiments were done in non-diabetic and  
59 streptozotocin-induced diabetic model rats with a single feeding in different prandial states and blood  
60 was collected. An intestinal perfusion technique was used to study the effects of *T. indica* seed powder  
61 on intestinal glucose absorption in normal and type-II model rats. Gut motility was evaluated using  
62 barium sulfate milk. Glucose was measured by Glucose oxidase-peroxidase (GOD-POD) method.

63 Result: The screening results showed that *T. indica* seed powder had no effect on fasting or  
64 postprandial serum glucose level of normal and type-I diabetic rat. The seed powder also showed no  
65 hypoglycemic effect in the fasting state and no antihyperglycemic effect in type-II model rats when fed  
66 simultaneously with oral glucose load, but it exhibited significant antihyperglycemic effect when the seed

67 powder was fed 30 minutes prior to the glucose load at 105 minutes ( $p < 0.03$ ). Glibenclamide  
68 significantly lowered postprandial serum glucose levels of non-diabetic and type-II diabetic model rats  
69 ( $p < 0.02-0.001$ ). *T. indica* exerted inhibition on glucose absorption in type-II rats during the whole  
70 perfusion period when compared with control. On the other hand, *T. indica* seed powder significantly  
71 inhibited the gastrointestinal motility in type-II rats.

72 Conclusion: The present data suggest that *T. indica* possesses antihyperglycemic properties in type-II  
73 rats which are at least partly due to its inhibitory effect on intestinal glucose absorption. This effect  
74 cannot be attributed to the acceleration of intestinal transit.

75

76 **KEY WORDS:** Anti-hyperglycemic, *Tamarindus indica*, streptozotocin, type-I diabetes, type-II  
77 diabetes, gastro intestine.

78

79

80

81

82

### 83 1. INTRODUCTION

84 Diabetes mellitus is a disease due to abnormality of carbohydrate metabolism and it is mainly linked  
85 with low blood insulin level or insensitivity of target organs to insulin. It is the most prevalent chronic  
86 disease in the world affecting a large population and its prevalence is about 6.8% [1].

87

88 Hyperglycemia and hyperlipidemia are two important characteristic of diabetes mellitus, an endocrine  
89 disorder based disease. In modern medicine, no satisfactory effective therapy is still available to cure  
90 diabetes mellitus [2]. Though pharmaceutical drugs like sulfonylureas and biguanides are used for the  
91 treatment of diabetes but these are either too expensive or have undesirable side effects or  
92 contraindications [3, 4]. In recent years, there has been renewed interest in plant medicine [5, 6, 7] for  
93 the treatment against different diseases as herbal drugs are generally out of toxic effect [8, 9] reported  
94 from research work conducted on experimental model animal. Although in human, whether there is  
95 any toxic effect are not investigated but anti-diabetic effect of various plants having "folk medicine  
96 reputation" have been screened [10].

97

98 *Tamarindus indica* Linn. (*family*- Caesalpinaceae [Fabaceae]), locally known as Tetul tree, is found  
99 throughout the South Asian region and some portions of Africa. It is a large **ample**, evergreen tree, 12-  
100 18 m high with round bushy crown and comparatively smaller bole. Phytochemical investigation  
101 revealed the presence of many active constituents, such as phenolic compounds, cardiac glycosides,  
102 L-(-)mallic acid, tartaric acid, the mucilage and pectin, arabinose, xylose, galactose, glucose, and  
103 uronic acid [11, 12]. The fruit pulp contains large quantities (16-18%) of tartaric, citric, malic and  
104 acetic acids, potassium tartrate, invert sugar, gum and pectin. It also contains traces of oxalic acid.  
105 Seed testa contains a fixed oil. Seeds cotyledons contain albuminoids, fat and carbohydrates [13, 14].  
106 Traditional healers claim that the seed of the plant possess antidiabetic properties. Scientific reports  
107 also support the hypoglycemic activity of this plant [15, 16, 17, 18]. However, no published report  
108 supports the underlying mechanism of the hypoglycemic effect of *T. indica* seed powder **in** normal,  
109 STZ induced type-I and type-II diabetic model rats.

110

111 The purpose of this work was to explore the possible hypoglycemic / anti-hyperglycemic activity of *T.*  
112 *indica* seed extract **in** type-I and type-II diabetic rats as well as to investigate the possible mode of  
113 action beyond this activity.

114

## 115 **2. MATERIALS AND METHODS**

### 116 **2.1. Plant materials and preparation of test sample**

117 *Tamarindus indica* Linn. seeds were collected from Kushtia, a district of Bangladesh. The plant was  
118 identified by the Bangladesh National Herbarium (voucher specimen no. DACB-38203). The seeds  
119 were collected from the fresh fruit pulp and were washed carefully and dried for two days at 40°C in  
120 an oven. Then the seeds were crushed in an electric grinder to make fine powder. Afterwards the  
121 powder was stored immediately in the refrigerator at -20°C and **kept** in the same temperature up to  
122 end of the experiment.

123

### 124 **2.2. Experimental animals**

125 The study was conducted on adult male Long-Evans rats (weighing 160-210g) bred at the BIRDEM  
126 animal house maintained at a constant room temperature of 22±5°C, 40-70% humidity conditions and  
127 the natural day-night cycle with an *ad libitum* access to food except the day of experimental procedure

128 when animals were used after 12hrs fasting. The rats had no access to food during the whole period  
129 of blood sampling. The influence of circadian rhythms was avoided by starting all experiments at 8.30  
130 a.m.

131

### 132 **2.3. Induction of diabetes in rats**

133 Diabetes stimulating type-I was induced by a single intra-peritoneal injection of streptozotocin (STZ,  
134 Upjohn Company, Kalamazoo, MI USA) dissolved in 0.1 M citrate buffer, pH 4.5, at a dose of 65mg/kg  
135 body weight to adult rats [19]. On the 7<sup>th</sup> day rats (fasting blood glucose  $\geq 18$ mmol/l) were taken for  
136 carrying out the experiments.

137

138 Type-II diabetes was induced by a single intra-peritoneal injection of STZ at a dose of 90mg/ kg body  
139 weight, in citrate buffer to the 48 hours old pups as described by Bonner *et al.* [19]. Experiments were  
140 carried out 3 months after STZ injection an oral glucose (2.5g/kg bw) tolerance test was performed to  
141 check the blood glucose levels of the rats. Rats having blood glucose level 8-11 mmol/l at fasting  
142 condition were taken to carry out the experiments.

143

144 A total number of 250 rats were used to carry out the experiments, which include normal, type-I and  
145 type-II diabetic model rats. The animals were divided into 3 groups: Gr 1- water control group; Gr 2-  
146 glibenclamide treated positive control group in case of type-II diabetic model rats and inulin treated in  
147 case of type-I; Gr-3 *T. indica* seed powder treated group. Number of rats were 6-8 rats in each group.

### 148 **2.4. Acute effect on fasting and postprandial glucose level**

#### 149 **2.4.1. Fasting condition**

150 The powder (1.25g/kg bw/10ml) was fed to overnight fasting (12hrs) rats and blood samples were  
151 drawn at 0, 60 and 120 minutes [20]. The positive control group received glibenclamide (5mg/kgbw)  
152 for normal and type-II rats and insulin (10 $\mu$ l/rat) for type-I rats whereas the control group received only  
153 water (10ml/kg bw) [20]. Blood samples were collected by amputation of the tail tip under mild ethar  
154 anesthesia. The rats were kept unfed throughout the experimental period.

#### 155 **2.4.2. Postprandial condition**

156 *T.indica* seed powder (1.25 g/kg bw), with or without glucose (2.5g/10ml/kg bw), were fed to overnight  
157 fasted (12hrs) rats and blood samples were drawn at 0, 30, and 75 min when fed simultaneously with

158 glucose and at 0, 60 and 105 min when fed 30 min prior to glucose load. Both positive control and  
159 water control rats were fed with glucose solution (2.5g/10ml/kg bw) and water (10ml/kg bw) following  
160 glucose load [21].

### 161 **2.5. Effect of *T. indica* seed powder on intestinal glucose absorption**

162 An intestinal perfusion technique [22] was used to study the effects of *T.indica* seed powder on  
163 intestinal absorption of glucose in nondiabetic and type 2 diabetic rats fasted for 36 hours and  
164 anesthetized with sodium pentobarbital (50 mg/kg). The seed powder were added to a kreb's solution  
165 (g/L 1.02 CaCl<sub>2</sub>, 7.37 NaCl, 0.20 KCl, 0.065NaH<sub>2</sub>PO<sub>4</sub>.6H<sub>2</sub>O, 0.6 NaHCO<sub>3</sub>, pH 7.4), supplemented with  
166 glucose (54.0 g/L) and perfused at a perfusion rate of 0.5 mL/min for 30 min through the duodenum.  
167 The perfusate was collected from a catheter set at 40 cm. *T. indica* seed powder were added to kreb's  
168 solution to a final conc. of 25 mg/mL so that the amount of seed powder in the perfused intestine is  
169 equivalent to the dose of 1.25 g/kg. The control group was perfused only with kreb's buffer  
170 supplemented with glucose. The results were expressed as percentage of absorbed glucose,  
171 calculated from the amount of glucose in solution before and after the perfusion.

### 172 **2.6. Gastrointestinal (GI) motility test:**

173 Gastrointestinal motility was evaluated by using barium sulfate (BaSO<sub>4</sub>) milk method [23].

174 The experiment was carried out by the method previously described by Chatterjee (1993).  
175 Distilled water at a dose of 10ml/kg bw was fed with smooth metallic tube to control rats and  
176 *T indica* powder (1.25g/kg bw/ 10ml) was fed to treated group. Baso4 milk was prepared by  
177 adding Baso<sub>4</sub> as 10% w/v in 0.5% CMC suspension. The milk was given to rats after 1 hour  
178 of administration of the test material. The rats were sacrificed 15 minutes after the  
179 administration of the milk. Before sacrificing the rats were anesthetized with di-ethyl ether.  
180 Then the abdominal part was opened and the intestinal part from pylorous to the ileoceccal  
181 junction as a total length of GI was taken into a petridish filled with distilled water for  
182 washing. It was then soaked by tissue paper to make it dry and taken into a white paper  
183 marked with 100 cm scale for measurement. The total gastrointestinal tract was measured  
184 first. Then the length traversed by Baso<sub>4</sub> was measured (white color). This length traversed  
185 by Baso<sub>4</sub> was expressed as a percentage of the total length of small intestine and the result of  
186 the test group was compared with that of control group (Chatterjee1993).

187

188

### 189 **2.7. Biochemical analysis:**

190 Serum glucose was measured on the same day by glucose-oxidase method (GOD-POD) using a  
191 commercial kit (Boehringer-Mannheim, GmbH, Germany) (Sera Pak, USA).

192

### 193 **2.8 Statistical analysis:**

194 Data from the experiments were analyzed using the Statistical Package for Social Sciences (SPSS)  
195 software for Windows version 12 (SPSS Inc, Chicago Illinois, USA). Values were expressed as  
196 mean±SD (Standard Deviation), Analysis of variance (ANOVA, Bonferroni Post Hock Test) and  
197 independent sample 't'-test were done as the test of significance  $p \leq 0.05$  was considered as the  
198 minimal level of statistical significance.

199

## 200 **3. RESULTS**

### 201 **3.1. Acute effect of *Tamarindus indica* seed powder on blood glucose level of normal (non- 202 diabetic), type- I and type- II diabetic model rats**

203 Blood glucose level was analyzed at the fasting level and the results showed that the seed powder  
204 had no significant effect on the fasting state of normal rats but lowered the serum glucose level 2.43%  
205 at 60 min and 7.68% at 120 min. As expected glibenclamide, the positive control, lowered serum  
206 glucose level significantly both at 60 minutes ( $p=0.000$ ) and at 120 minutes ( $p=0.000$ ) compared with  
207 water control and powder treated groups (Table 1).

208

209 **Table 1: Effect of *T. indica* seed powder on fasting serum glucose level (M±SD) of normal,  
210 type- I and type- II diabetic model rats:**

Group	Min 0 (mmol/l)	Min 60 (mmol/l)	Min 120 (mmol/l)
<b>Normal rat</b>			
<b>Water control(n =6)</b>	8.05±0.51	7.63±0.49	6.99±0.73
<b>Glibenclamide(n =6)</b>	8.39±0.76	5.67±0.88*	4.83±0.48*
<b><i>T. indica</i> seed powder (n =8)</b>	7.82±0.68	7.63±0.41	7.22±0.61
<b>Type I diabetic model rat</b>			

<b>Water control (n = 6)</b>	22.04±1.28	20.53±3.09	19.40±3.92
<b>Insulin (n =6)</b>	21.49±1.58	4.30±1.87*	2.32±0.37*
<b>T. indica seed powder (n =6)</b>	21.38±1.51	20.84±1.90	19.51±2.66
<b>Type-II diabetic model rat</b>			
<b>Water control(n =6)</b>	8.35±1.57	8.87±2.50	10.53±3.88
<b>Glibenclamide(n =6)</b>	8.55±1.55	8.01±1.91	7.42±1.56*
<b>T. indica seed powder (n =8)</b>	9.10±1.12	9.93±2.37	9.57±1.94

211

212 Results are expressed as **mean ±standard** deviation (M±SD). One-way ANOVA (Bonferroni test) was done for  
 213 comparing the different group and for the test of significance. \*P= 0.000 **when compared** with water control;  
 214 n=number of rats.

215

216 The oral glucose tolerance test (OGTT) was performed and the results showed that powder had  
 217 glucose lowering effect but non-significantly and the glibenclamide treated group showed a significant  
 218 fall in serum glucose level at 75 minutes (p=0.000) (Table 2).

219

220 **Table 2: Effect of T .indica seed powder on serum glucose level (M±SD) of normal, type-I and**  
 221 **type-II diabetic model rats when seed powder was fed simultaneously with glucose load:**

<b>Group</b>	<b>Min 0 (mmol/l)</b>	<b>Min 30 (mmol/l)</b>	<b>Min 75 (mmol/l)</b>	<b>iobv</b>
<b>Normal rat</b>				
<b>Water control(n = 7)</b>	6.38±0.51	8.36±0.68	7.79±0.75	3.39±1.71
<b>Glibenclamide(n = 6)</b>	5.84±0.66	7.27±0.38	5.55±0.78*	1.14±2.18
<b>T. indica seed powder (n = 7)</b>	6.15±1.21	8.64±0.90	7.92±0.63	4.26±3.28
<b>Type-I diabetic model rat</b>				
<b>Water control(n = 7)</b>	20.90±3.33	26.90±2.62	25.25±3.79	10.35±7.06



<b>Insulin(n = 6)</b>	21.86±2.16	18.38±6.48*	9.15±4.77**	-16.19±9.95
<b>T. indica seed Powder (n = 6)</b>	20.88±1.80	28.67±2.56	25.63±3.23	12.53±6.35
<b>Type-II diabetic model rat</b>				
<b>Water control(n = 6)</b>	7.64±1.42	17.10±4.24	17.36±3.42	19.17±8.80
<b>Glibenclamide(n = 6)</b>	9.01±1.40	16.25±0.98	12.72±0.90*	10.95±3.01
<b>T. indica seed powder (n = 7)</b>	8.21±2.10	16.97±1.86	15.49±2.40	16.03±6.24

222 Results are expressed as mean ±standard deviation (M±SD). One-way ANOVA (Bonferroni test) was done for  
 223 comparing the different group and for the test of significance. \*P= 0.000 when compared when compared with  
 224 water control; iobv =Sum of the increments over the basal value. n=number of rats.

225

226 When fed 30 min before to glucose load, it was found that glibenclamide treated group showed  
 227 significant blood glucose lowering effect at 60 minutes (p = 0.001) and 105 minutes (p = 0.000)  
 228 normal and type-I diabetic model rats in comparison to water control and powder treated group (Table  
 229 3). T. indica seed powder showed significant (p = 0.003) blood glucose lowering effect at 105 min. on  
 230 type-II diabetic model rats in Comparison to water control and glibenclamide treated group (Table 3;  
 231 Fig. 1).

232

233 **Table 3: Effect of T .indica seed powder on serum glucose level (M±SD) of normal, type-I and**  
 234 **type-II diabetic model rats when seed powder was fed 30 minutes before to glucose load:**

<b>Group</b>	<b>Min 0 (mmol/l)</b>	<b>Min 60 (mmol/l)</b>	<b>Min 105 (mmol/l)</b>	<b>iobv</b>
<b>Normal rat</b>				
<b>Water control (n =6)</b>	6.64±1.09	9.05±0.61	8.51±0.70	3.27±2.86
<b>Glibenclamide (n =6)</b>	6.85±1.23	5.37±0.93*	3.83±0.48**	-4.49±1.90
<b>T.indica seed Powder (n=8)</b>	6.87±0.82	8.00±0.78	8.30±0.63	2.55±1.85
<b>Type-I diabetic model rat</b>				
<b>Water control(n = 6)</b>	24.26±2.03	31.71±3.62	30.78±1.20	9.97±6.66

<b>Insulin(n = 7)</b>	22.99±2.09	11.47±5.84 *	8.28±3.62*	-26.22±6.06
<b>T. indica seed Powder (n = 6)</b>	23.79±2.41	32.12±4.06	28.08±2.42	12.62±8.42
<b>Type-II diabetic model rat</b>				
<b>Water control (n = 6)</b>	7.96±1.79	19.65±4.36	19.30±4.27	23.01±9.99
<b>Glibenclamide (n = 6)</b>	9.05±1.78	18.99±5.37	16.09±4.68	19.98±8.91
<b>T. indica seed Powder (n = 6)</b>	9.59±1.72	17.10±2.31	14.65±2.82*	12.57±2.84*

235

236

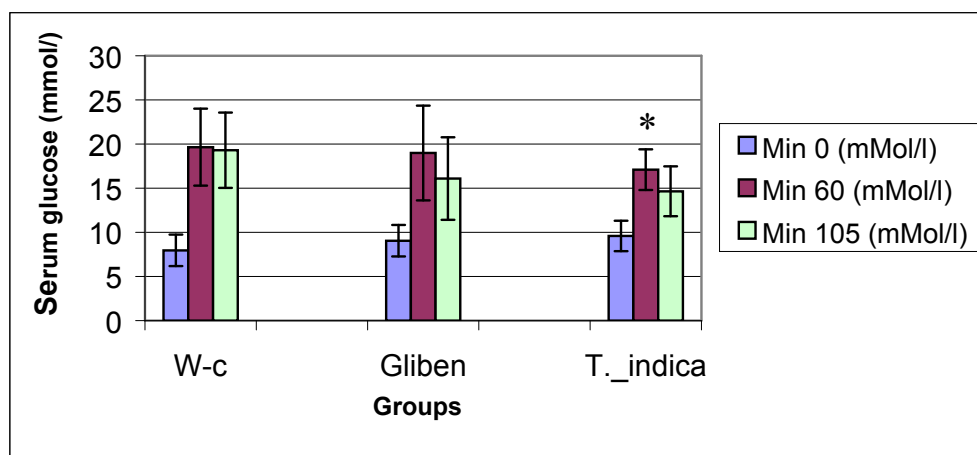
Results are expressed as mean ±standard deviation (M±SD). One way ANOVA (Bonferroni test) was done for comparing the different group and for the test of significance. \*\*P= 0.000.and. \*p= 0.001.+ when compared when compared with water control; lobv =Sum of the increments over the basal value. n=number of rats.

237

238

239

240



241

242

**Figure 1:** Acute effect of T. indica seed powder on serum glucose level on Type II diabetic model

243

rats at 105 min. when feed at 30 min. before to glucose load. W-c=Water control.

244

### 245 **3.2. Effect of Tamarindus indica seed powder on gastrointestinal motility**

246

The length of GI traversed by BaSO<sub>4</sub> milk in powder treated group was lower than the water treated

247

group. The inhibition in motility was not statistically significant in treated group of normal rats (Table

248

4).There was decreased percentage of length traversed by BaSO<sub>4</sub> with seed powder on type-II

249 model rats in comparison to water control group. *T. indica* seed powder showed significant **inhibitory**  
 250 **effect on gastrointestinal** motility (p=0.02).

251

252 **Table 4: Effect of *Tamarindus indica* Linn seeds powder on gastrointestinal motility**  
 253 **test by Baso<sub>4</sub> milk of normal and type-II diabetic model rats:**

254

Group	GI total length (cm)	Length traversed by BaSO <sub>4</sub> (cm)	% of Length traversed by BaSO <sub>4</sub>
<b>Normal rat</b>			
<b>Control (n=6)</b>			
	118.33±9.83	56.33±6.18	47.93±7.04
<b><i>T indica</i> Seed Powder (n=6)</b>	123.83±6.11	47.67±7.86	40.77±9.77
<b>Type-II diabetic model rat</b>			
<b>Control (n=5)</b>			
	121.00±5.47	58.00±6.78	47.89±4.88
<b><i>T indica</i> Seed Powder (n=5)</b>	121.00±9.61	48.00±8.09	39.46±4.31*

255 GI= Gastro Intestine. Data are presented as **mean ±standard** deviation (M±SD) and Group was

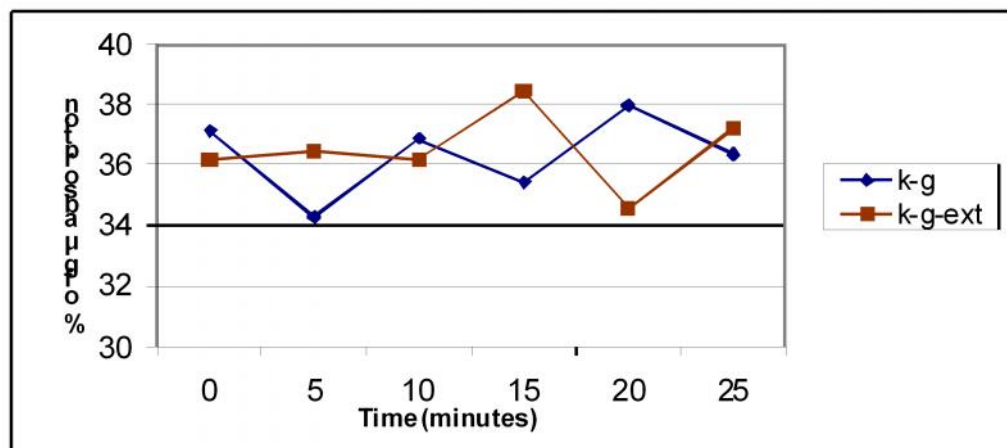
256 Compared by using independent samples 't' test.\*p =0.02. n=number of rats.

257

258 **3.3. Effect of *T indica* seed powder on upper intestinal glucose absorption**

259 It is evident that the upper intestinal glucose absorption was almost constant during 30 minutes of  
 260 perfusion with glucose. When the seed powder solution was supplemented with the glucose solution, it  
 261 showed no effect on intestinal glucose absorption in normal rats (Figure 2).

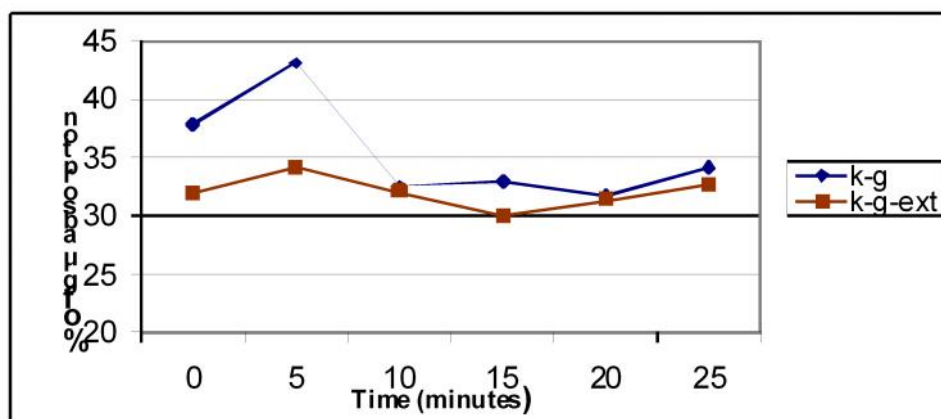
262



263  
264 **Figure 2: Effect of the *T. indica* seed powder on upper intestinal glucose absorption on**  
265 **normal rats.**

266 Results are presented as mean± SD (n=5). Rats were fasted for 36 hours and intestine was  
267 perfused with glucose solution (54 g/l) with or without *T. Indica* seed powder (25mg/ml); glu=  
268 glucose, K-g= Kreb's solution, K-g-ext= Kreb's solution + glucose +seed powder juice.

269  
270 In case of type-II model rats, intestinal glucose absorption was nearly constant during the 30 min of  
271 perfusion with glucose. There was a decrease in glucose absorption with glucose solution when  
272 supplemented with seed powder solution (Figure 3).



273  
274 **Figure 3: Effect of *T. indica* seed powder on upper intestinal glucose absorption on type-II**  
275 **diabetic rats.**

276 Results are presented as mean± SD (n=6). Rats were fasted for 36 hours and intestine was perfused with  
277 glucose solution (54 g/l) with or without *T. Indica* seed powder (25mg/ml); glu= glucose, K-g= Kreb's solution, K-  
278 g-ext= Kreb's solution + glucose + Extract.

280 It denotes that *T indica* seed powder strongly affected the amount of absorbed glucose throughout the  
281 notable period of experiment in type-II rats. Figure 3 depicts the gradual fall in glucose absorption  
282 during the whole perfusion period in type-II rats compared to Krebs solution. Therefore, the obtained  
283 results suggest that seed powder of the *T. indica* delays glucose absorption in the upper part of the  
284 gastrointestinal tract.

285 .

#### 286 **4. DISCUSSION**

287 Synthetic drugs such as Sulphonylureas and Biguanides are valuable in the treatment of DM, but their  
288 uses are restricted by their limited action, pharmacokinetic properties, secondary failure rates and  
289 accompanying side effects [24]. Moreover, these therapies only partially compensate for metabolic  
290 derangements seen in diabetes and do not necessarily correct the fundamental biochemical lesion  
291 [25]. As the incidence of diabetes is rising relatively around the world, there is an urgent need to  
292 expand the range of effective palliatives available to patients.

293

294 The present study has been undertaken to screen the hypoglycemic and anti-hyperglycemic activity of  
295 *Tamarindus indica* seed powder in nondiabetic, type-I and type-II diabetic model rats. The  
296 experimental approach that has been followed, in addition to screening the hypo-/antihyperglycemic  
297 activity, gives an approximate idea about the mechanism of action of the plant by analyzing the  
298 model, prandial states and timing of hypoglycemic activity. Moreover, the study was also extended to  
299 explore the possible mechanism of action by elucidating the effect of the plant on gut motility and  
300 intestinal glucose absorption.

301

302 Our results demonstrate that *T. indica* seed powder had no effect in the fasting state of nondiabetic,  
303 type-I or type-II rats. At the post prandial state when the seed powder was administered  
304 simultaneously with oral glucose load, no significant reduction was noticed with a single feeding in any  
305 group of rats. On the contrary, when *T. indica* seed powder was administered half an hour before oral  
306 glucose load in type-II rats, the seed powder caused a significant attenuation in the rise of blood  
307 glucose at 105 minutes compared to the control groups (glucose  $M\pm SD$ , mmol/l.  $14.65\pm 2.82$  in the  
308 treated group Vs  $19.30\pm 4.27$  in the control group,  $p<0.03$ ). The antihyperglycemic effect of *T. indica*  
309 seed in STZ induced diabetic rats have been found by other investigators [17].

310

311 *T. indica* seed powder was effective in type-II diabetic model rats when fed 30 minutes before glucose  
312 load. This effect might be due to a systemic action, i.e. as a result of the stimulation of Beta cells and  
313 improving the insulin secretory capacity or enhancement of insulin action by the extract. This effect  
314 could not be confirmed by our study since serum insulin level after a single feeding was not  
315 determined. It has been claimed that the chronic aqueous extract of *T indica* improves glycemic status  
316 [17, 26].

317

318 The inhibition of intestinal glucose absorption may contribute to the reduction of postprandial glucose  
319 level. One of the objectives of the present study was to investigate whether the hypoglycemic effect is  
320 related to the inhibition of glucose absorption in the gut. Since the antihyperglycemic effect of *T. indica*  
321 was found in type II rats, therefore, this gut perfusion experiment was investigated in normal and  
322 type-II rats, where *T. indica* seed powder showed strong inhibition of glucose absorption. This result  
323 strongly suggests that the antihyperglycemic effect of *T. indica* as previously reported [26, 27] may be  
324 due to, at least in part to the retardation of glucose absorption in the small intestine. *T. indica* is rich in  
325 pectin's and dietary fibers such as cellulose, xylose and mucilage [16] and the presence of such  
326 substances in the powder of *T. indica* may be responsible for the observed effect. Similar results have  
327 been reported by some other scientists [16, 27]. Moreover, *T. indica* seed powder also inhibited the  
328 BaSO<sub>4</sub> induced gastrointestinal motility in Type-II rats. This result suggests that the decrease of  
329 glucose absorption by *T. indica* seed powder is not achieved by an enhanced intestinal motility. It is  
330 now well established that diabetes mellitus is not a single disease entity, but a heterogenous group of disorders with  
331 a striking diversity of etiopathogenetic mechanisms as well as clinical manifestations [28]. It is also  
332 established that the basic pathophysiology of Type-I and Type-II diabetes is quite different. In type-II diabetes, there are  
333 multiple abnormalities in diverse tissues. So, a plant material can show glucose lowering effect in diverse way. It may not be  
334 active in type-I diabetes, but may be active in type-II diabetes, which we found in our study.

335

336

337 **5. CONCLUSION**

338 Based on the results of this study, it may be concluded that *T. indica* seed powder possesses  
339 significant antihyperglycemic activity in type II diabetic model rats but not in type I and this is partly  
340 due to inhibition of intestinal glucose absorption.

341

342

343

344

345

346

347

#### 348 **ACKNOWLEDGEMENT**

349 We gratefully acknowledge the financial support of International Program for the Chemical Sciences  
350 (IPICS), Uppsala, Sweden and International Foundation for Sciences (IFS), Stockholm, Sweden and  
351 Diabetic Association of Bangladesh in conducting this study.

352

#### 353 **COMPETING INTERESTS**

354 Authors have declared that no competing interests exist.

355

#### 356 **AUTHOR'S CONTRIBUTION**

357 Anzana Parvin, designed the proposal and protocol, performed the experiments and analysis; Md.  
358 Morshedul Alam wrote the first draft of manuscript; Md. Anwarul Haque, Amrita Bhoumik and Liaquate  
359 Ali managed the study analysis; Begum Rokeya designed the protocol, performed statistical analysis,  
360 managed the experiment. All authors approved the manuscript.

361

362

363

364

365

366

367

368

369

370  
371  
372  
373  
374  
375  
376  
377  
378

379 **REFERENCE**

- 380 1. Emmanuela G, Leslie M, Jesse AK, Ramiro G, Salvador V, Ruy LR, Wichai A, Mohsen N,  
381 Stephen L, Rafael L, Christopher JLM. Management of diabetes and associated  
382 cardiovascular risk factors in seven countries: a comparison of data from national health  
383 examination surveys. *Bulletin of the World Health Organization*. 2011; 89: 172-183
- 384 2. Ghosh S, Suryawanshi SA. Effect of *Vinca rosea* extracts in treatment of alloxan diabetes  
385 in male albino rats. *Indian Journal of Experimental Biology*. 2001; 39: 748-759.
- 386 3. Berger W. Incidence of severe side effects during therapy with sulphonylureas and  
387 biguanides. *Hormones Metabolic Res*. 1985; 17: 111-115.
- 388 4. Rang HP, Dale MM, Ritter JM. The endocrine system Pharmacology. In: *Pharmacology*.  
389 Longman Group Ltd., UK. 1991; 504-508.
- 390 5. Dubey GP, Dixit SP, Alok S. Alloxan-induced diabetes in rabbits and effect of herbal  
391 formulation D-400. *Indian Journal of Pharmacology*. 1994; 26: 225-226.
- 392 6. Prince PS, Menon VP, Pari L. Hypoglycemic activity of *Syzygium cuminii* seeds: effect on  
393 lipid peroxidation in alloxan diabetes rats. *Journal of Ethnopharmacology*. 1998; 61: 1-7.
- 394 7. Ladeji O, Omekarah I, Solomon M. Hypoglycemic properties of aqueous bark extract of  
395 *Ceiba pentandra* in streptozotocin-induced diabetic rats. *Journal of Ethnopharmacology*.  
396 2003; 84: 139-142.



- 397 8. Geetha BS, Biju CM, Augusti KT. Hypoglycemic effect of leucodelphinidin derivative  
398 isolated from *Ficus bengalensis* (Linn.). *Indian Journal of Pharmacology*. 1994; 38: 220-222.
- 399 9. Rao BK, Sudarshan PR, Rajasekhar MD, Nagaraju N, Rao CA. Antidiabetic activity of  
400 *Terminalia pallida* fruit in alloxan-induced diabetic rats. *Journal of Ethnopharmacology*.  
401 2003; 85: 169-172.
- 402 10. Vats V, Grover JK, Rathi SS. Evaluation of antihyperglycemic effect of *Trigonella*  
403 *foenum-graecum* Linn, *Occium sanctum* Linn. and *Pterocarpus marsupium* Linn., in normal  
404 and alloxanised diabetic rats. *Journal of Ethnopharmacology*. 2002; 79: 95-100.
- 405 11. Rasu N, Saleem B, Nawaz R. Preliminary screening of four common Plants of family  
406 *Caesalpinia*caee. *Pak J Pharm Sci*. 1989; 2: 55-7.
- 407 12. Ibrahim E, Abbas SA. Chemical and biological evaluation of *Tamarindus indica* L.  
408 growing in Sudan. *Acta Hortic*. 1995; 390: 51-7.
- 409 13. Bhadoriya SS, Ganeshpurkar A, Narwaria J, Rai G, Jain AP. *Tamarindus indica*: Extent  
410 of explored potential. *Pharmacogn Rev*. 2011; 5(9): 73–81.
- 411 14. Ghani A. *Medicinal Plants of Bangladesh*, 2<sup>nd</sup> Ed. The Asiatic Society of Bangladesh,  
412 Dhaka. 2003; 331-332.
- 413 15. Iyer SR. *Tamarindus indica* Linn. In: Warriier, P.K., Nambiar , V.P.K., Kutty, C.R.  
414 (Eds.), *Indian Medicinal Plants* , vol .V. Orient Longman Limited , Madras. 1995; 235-236.
- 415 16. Ibrahim NA, EI-Gengaihi S, EI-Hamidi A, Bashandy SAE. Chemical and Biological  
416 Evaluation of *Tamarindus indica* Linn Growing in Sudan. *Acta- hortic*: Wageningen:  
417 International society for Horticultural science. 1995; 390, 51-57.
- 418 17. Maiti R, Jana D, Das UK, Ghosh D. Antidiabetic effect of aqueous extract of seed of  
419 *Tamarindus indica* in streptozotocin-induced diabetic rats. *Journal of Ethnopharmacology*.  
420 2004; 92(1): 85-91.

- 421 18. Ramchander T, Rajkumar D, Sravanprasad M, Venkateshwarlu Goli, Dhanalakshmi CH,  
422 Arjun. Antidiabetic activity of aqueous methanolic extracts of leaf of *Tamarindus indica*. Int.  
423 J Pharm & Phy Res. 2012; 4(1): 5-7.
- 424 19. Bonner S, Trent DF, Honey RN, Weir GC. Responses of neonatal rat islets on  
425 streptozotocin limited beta cell regeneration and hyperglycemia. Diabetes. 1981; 30: 64-69.
- 426 20. Ali L, Azad Khan AK, Mamun MIR, Mosihuzzaman M, Nahar N, Nur-E-Alam M,  
427 Rokeya B. Studies on Hypoglycemic Effects Fruit Pulp, seed and Whole plant of *Momordica*  
428 *charantia* on Normal and Diabetic Model Rats. Planta Medica. 1993; 59: 408-412.
- 429 21. Morshed MA, Haque A, Rokeya B, Ali L. Anti-Hyperglycemic effect of *Terminalia*  
430 *arjuna* bark extract on streptozotocin induced type 2 diabetic model rats. International  
431 Journal of Pharmacy and Pharmaceutical Sciences. 2011; 3(4): 449-453.
- 432 22. Swintosky Joseph V, Elzbieta Pogonowska- Wala. The in situ rat gut technique. A simple  
433 rapid, inexpensive way to study factors influencing drug absorption rate from the intestine.  
434 Pharmacy. 1982; 3(5): 163-167.
- 435 23. Chatterjee TK. Handbook on Laboratory mice and rats. Dept. of Pharmaceutical  
436 Technology, Jadavpur University. 1993.
- 437 24. Bailey CJ, Day C. Traditional Plant medicines as treatments for Diabetes. Diabetes Care.  
438 1989; 12(8): 553-564.
- 439 25. Tailor R, Agius L. The Biochemistry of Diabetes. Biochemistry Journal. 1988; 250: 650-  
440 740.
- 441 26. Maiti R, Das UK, Ghosh D. Attenuation of Hyperglycemia and Hyperlipidemia in  
442 Streptozotocin-induced Diabetic rats by aqueous extract of seed of *Tamarindus indica*. Biol.  
443 Pharm. Bull. 2005; 28(7): 1172-1176.

444 27. Shehla Imam, Iqbal Azhar, Hasan M. Mohtasheemul, Ali MS, Waseemuddin Ahmed S.  
445 Two triterpenes lupanone and lupeol isolated and identified from *Tamarindus indica* Linn.  
446 Pak. J. Pharmaceutical Science. 2007; 20(2): 125-127.

447 28. Ganda OP, Soeldner SS. Genetic, acquired, and related factors in the etiology of diabetes  
448 mellitus. Arch Intern Med, 1977, apr 137(4): 461- 469.

449  
450  
451  
452