

Effect of low radiation dose on cisplatin induced hepato-testicular damage in male rats.

ABSTRACT

Aim: The aim of this study was to evaluate the synergistic effect of low radiation dose with the chemotherapeutic drugs in order to find possible way to lessen the harmful effects during chemo-radiotherapy.

Study Design: Randomized controlled experiment.

Place and Duration of Study: Experimental Animal Unit, Drug Radiation Research Department, National Center for Radiation Research and Technology, Cairo Egypt.

Methodology: Estimation of antioxidant activity of low radiation dose on oxidative stress induced by cisplatin administration at a dose of (10 mg/kg bwt) in male albino rat.

Results: Results of experiment revealed that cisplatin administration caused a significant increase in serum alanine transaminase (GPT) activity (38.58 ± 2.060) and FSH level (8.162 ± 1.424) accompanied with a decrease in serum albumin (3.492 ± 0.253), and Butyryl Cholein Esterase (BChE) (65.35 ± 12.61). In Liver and testis, GSH content (68.00 ± 2.391 & 24.93 ± 4.778) as well as cytochromes P450 levels (0.3875 ± 0.0727 & 0.2167 ± 0.0459) showed a significant decrease as compared to the normal control level respectively. In addition the level of Fe, Cu and Zn showed no significant changes in liver and appeared to be significantly decrease as in case of corresponding trace elements in testis organs. On the other hand, exposing to low dose of radiation (0.5 Gy) post-cisplatin treatment effectively prevented these alterations and maintained the antioxidant status.

Conclusion: Data from present results revealed that low radiation dose have the existence as an antioxidant and antitumor agents which may be useful to use as a synergistic agents with the chemotherapeutic drug.

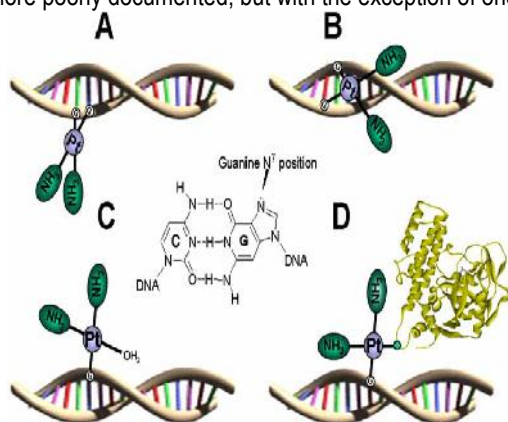
Keywords: Low Radiation Dose, Cisplatin, Cytochromes P450, Butyryl Cholein Esterase.

1. INTRODUCTION

Cisplatin is one of the most potent anticancer drugs used in chemotherapy. It is highly effective in the treatment of testicular and ovarian cancers and is also widely employed for treating bladder, cervical, head and neck, esophageal and small cell lung cancer [1]. Despite being an effective anti-proliferative agent, the clinical usage of cisplatin is limited by various side effects including nephro-toxicity, hepato-toxicity, neurotoxicity and oto-toxicity. The anticancer activity of this drug is attributed to its capacity to form covalent bond at N-7 position of Guanine residues of DNA leading to formation of 1, 2 or 1,3 inter-strand crosslink and a lesser extent of the interstrand crosslink (Fig. 1). These adduct cisplatin DNA disrupts the cellular replication and transcription machinery [2]. Many labs have demonstrated that cisplatin generates free radicals leading to oxidative and nitrosative stress which results into such deleterious effects *in vivo* [3, 4]. It is generally accepted that binding of cisplatin to genomic DNA (gDNA) in the cell nucleus is the main event responsible for its antitumor properties [5]. Thus, the damage induced upon binding of cisplatin to gDNA may inhibit transcription, and/or DNA replication mechanisms. Subsequently, these alterations in DNA processing would trigger cytotoxic processes that lead to cancer cell death. The cytotoxicity of cisplatin is considered to be due to a combination of factors, including peroxidation of the cell membrane, mitochondrial dysfunction, inhibition of protein synthesis, and DNA injury [6].

Till the last decade of the last century radiotherapy was the only therapeutic option for patients with locally advanced cancer [7]. There are some trails of combined cisplatin and radiation therapies have been reported and it has proved to be hopeful method to improve the treatment results of invasive bladder cancer [8]. The effectiveness of the combination of cisplatin and radiation in experimental malignant tumor is reported by Wodinsky and his colleagues [9] in leukaemic cell. Morris and his colleagues noted a similar effect in mouse leukaemia [10]. Cytotoxic chemotherapy has been shown to give good response rates in patients with good kidney function and no prior radiation therapy. Cisplatin is the most effective single agent [11], and has been shown in cell lines to be synergistic with radiotherapy. Mechanisms underlying the interaction between drugs and radiation may include inhibition of potentially lethal or sublethal damage repair, and increasing radio-sensitivity of hypoxic cells [12]. It has been widely used prior to surgery or radiotherapy with the aim of reducing tumour volume and facilitating local treatment. It may have the additional benefit of controlling micrometastatic

32 disease. Combined chemo-radiotherapy seems to offer substantial benefit for women with cervical cancer. However,
 33 acute toxicity, predominantly haematological and gastrointestinal, was increased with chemo-radiation [13]. Acute side
 34 effects are generally of short duration and resolve with medical management, while the late complications of radiotherapy
 35 lead to damage which can be difficult to reverse, and may permanently impair quality of life. Details of late morbidity are
 36 more poorly documented, but with the exception of one trial [14].



37
 38
 39
 40
 41
 42
 43
 44
 45
 46
 47
 48
Fig. (1) Main adducts formed after binding of cisplatin to DNA. (A) 1,2-intrastrand cross-link, (B) interstrand cross-link, (C) mono- functional adduct, and (D) protein-DNA cross-link. The main site of attack of cisplatin to DNA (N7 of guanine) is shown in the central panel [2].

49 Low dose of radiation induce various effects including radio-protective response [15], as well as activation of immune
 50 function [16]. It has been well documented that the immune function is linked to the release of radical oxygen species
 51 (ROS). Access amount of ROS is commonly eliminated by endogenous antioxidant system, thereby preventing injury to
 52 DNA [17]. It exerts protection against other challenges involving radicals and causing a beneficial effect by temporarily
 53 shielding the hit cell against radicals produced through endogenous processes. Low dose of radiation has been observed
 54 to stimulate the radical detoxification system, enhancement of DNA repair rates and induce immune competence that
 55 associate with an increase in number of cytotoxic lymphocytes, even causing a reduction of the incidence of metastatic
 56 cancer [18]. Our pervious study indicated that treatment with low dose of gamma rays (0.5 Gy) ameliorate harmful effects
 57 induced by TCE due to the effect of gamma radiation as a stimulant of radical detoxification [19].

58 According to many clinical studies, when chemotherapy and radiotherapy are concurrently administered improve
 59 effectively greater rather than at different times [20]. Combined chemo-radiotherapy presents some problems; however,
 60 with more sever adverse events, resulting in a reduced treatment compilation rate [21]. On the other hand, according to
 61 the study of Murphy and Morton [22] and Barcellos-Hoff, [23], low radiation dose administartion to entire body increased
 62 the action of the protective process in living organism including the overproduction lymphocytes that significantly
 63 prevented or impaired tumor growth. From this point of view, the current study's aim is to investigate the synergistic effect
 64 of low radiation dose in the treatment with cisplatin as a chemotherapeutic drug inducing oxidative stress.

65 2. MATERIAL AND METHODS

66 2.1. Animals

67 Male albino rats weighing approximately 120-150 g were used for this experiment. They were housed in polypropylene
 68 cages in an air conditioned room with temperature maintained at $25\text{ }^{\circ}\text{C} \pm 3\text{ }^{\circ}\text{C}$, relative humidity of $50\% \pm 5\%$ and 12 h
 69 alternating light and dark cycles. The rats were provided with a nutritionally adequate chow diet and drinking water *ad*
 70 *libitum* throughout the study. Experiments were begun after a four-week acclimatization period.

72 2.2. Chemicals

73 Cisplatin was purchased from Central Drug House (Egypt). The chemicals used in this experiment were obtained from
 74 Sigma Chemical (USA). Kits used in this experiments were purchased from Bio-Diagnostics (UK).

75 2.3. Radiation process:

76 A single dose whole body irradiation (0.5 Gy) was performed with rats, using gamma rays by Cesium 137 irradiation unit,
 77 National Center for Radiation Research and Technology (NCRRT), with the dose rate 0.74589 rad/ sec. The gamma
 78 cesium cell was calibrated by alanine dosimetry relative to a primary standard. Correction were made daily for humidity,
 79 temperature, and barometric pressure.

80 2.4. Experimental design

81 Adult male albino rats were divided into 4 groups of 6 rats each and treated as follows:

82 **Control group** (normal, untreated), received distilled water. **Cis. group**, received freshly dissolved cisplatin in 1 ml
83 distilled water at a dose of 10 mg/ kg body weight (I.P); **IRR group** was exposed to a single low gamma radiation dose of
84 0.5 Gy and **Cis. & IRR group** was administered with Cisplatin, 24 hr after cisplatin administration the animals were
85 exposed to a single low dose of gamma rays (0.4 Gy). The dose of cisplatin was decided on the basis of Máthé and his
86 colleagues [24]. Twenty four hours after radiation exposure the animals were decapitated and the blood was collected for
87 biochemical analysis. Liver and testes were divided into two portions. One portion were excised, homogenized in ice-cold
88 saline and utilized for various biochemical analyses and the second portion was used for trace elements analysis.

89 2.5. Biochemical analysis

90 The blood samples were collected directly from the animals by heart puncturing. They were centrifuged at 3000 rpm for
91 15 min., clear sera were collected and stored in a refrigerator. The activities of GPT, butyryl cholinesterase (BChE) and
92 FSH levels as well as concentrations of albumin in serum were analyzed. Liver and testis were minced and homogenized
93 (10 % w/ v) in ice-cold normal saline solution. The homogenate was centrifuged at 3,000 rpm for 15-20 min at 4 °C. The
94 resultant supernatant was used for estimation of GSH content and cytochrome P450 activity. Serum
95 butyrylcholinesterase activity is quantitatively measured according to the methods published in Munshaw and his
96 colleagues [25]. FSH level was detected in serum samples using elisa kit supplied by (Kamiya, biomedical company,
97 USA) according to manufacturer instruction. The elisa kit was read using 2100 elisa reader. Serum albumin level was
98 determined according to the method of Doumas and his colleagues [26]. In addition GSH content in both liver and testis
99 was measured by method of Beutler and his colleagues [27]. Liver and testis cytochrome P450 activity assayed is
100 detected using a spectrofluorometer [28]. Serum butyrylcholinesterase and FSH activities as well as liver and testis P450
101 were carried out at the Central Laboratory, Radioisotope Dept., AEA, Giza, Egypt, while serum GPT activity and Albumin
102 concentration, as well as tissue GSH content were performed with a Helios Thermo-Spectronic spectrophotometer
103 (Thermo Spectronic, UK).

104 2.6. Atomic absorption analysis

105 Liver and testes tissues were digested in a mixture of conc. HNO₃ and H₂O₂ (5:1). The digestion was completely by using
106 micro-wave oven (mls- 1200 mega) and the digested sample were diluted with de-ionized water to a fixed volume [29].
107 The selected element was estimated quantitatively by atomic absorption spectrophotometer. *Unicam 939* Hallow cathode
108 lamps were used to determine Fe, Cu and Zn (MT) in air acetylene flame [30]. Concentration of elements in tissues was
109 calculated by using calibration curve prepared from their stock solution (1mg). The concentration of elements per grams
110 tissues could be determined by equation:

111 $\text{Conc. In } (\mu\text{g/g}) = [\text{Conc.}(\mu\text{g/ml}) / \text{sample weight}] \times \text{dil factor.}$

112 Wet tissue weights were used for calculating the metal concentration in tissues [31].

113

114 2.7. Statistical Analysis:

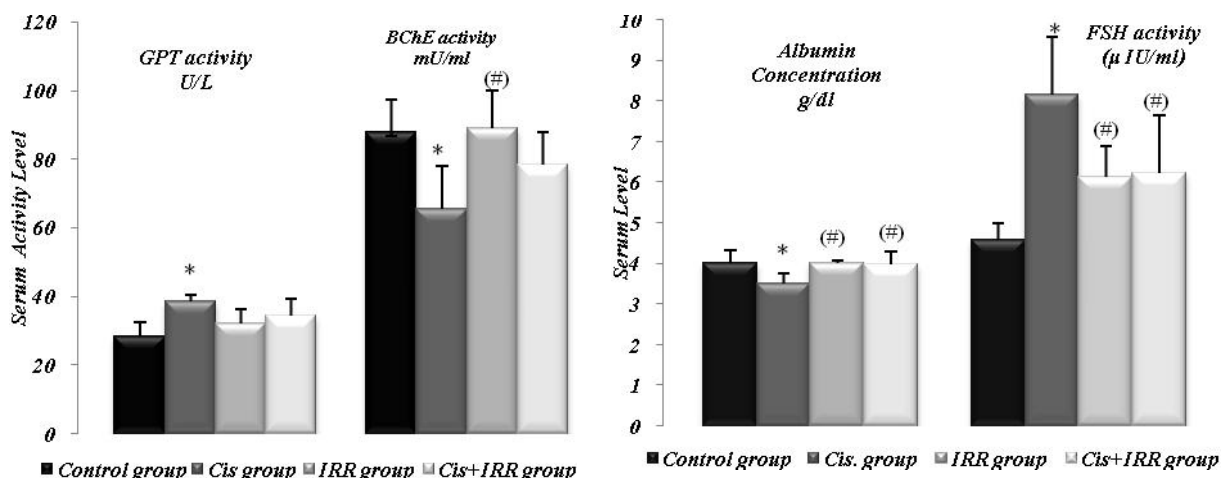
115 To assess the significant level of influence caused by low dose of radiation in cisplatin administrated rats, one way
116 analysis of variances (ANOVA) followed by Tukey's multiple comparison test was used. Data are representative of 8
117 independent experiments carried out in triplicate. Statistical analysis was performed by using Graph-Pad software, San
118 Diago, CA, USA) Differences were considered statistically significant when the P value was less than 0.05.

119

120 3. RESULTS

121 Cisplatin is one of the most potent anticancer drugs used for the treatment of different types of cancer including testicular
122 cancer. In spite of its significant anticancer activity, the clinical use of cisplatin is often limited by its undesirable side
123 effect due to the influence of oxidative stress. Results indicated in fig. (2) in addition to table (1-4) evaluate the effect of
124 cisplatin and the synergistic effect of low irradiation dose (0.5 Gy) on liver and testis tissues.

125



126
127
128
129
130

Fig. 2: Effects of cisplatin administration and/ or radiation exposure on some serum biochemical parameters 24 hours post radiation exposure (n=6)

Data are presented as mean (± SD). * Significantly different from the control group (P<0.05). (#)Significantly different from the Cis. group (P<0.05). IRR: Low radiation dose of 0.5 Gy gamma rays, Cis group: cisplatin administration.

131 **3.1. Cisplatin demonstrated toxicity in both liver and testis:**

132 **3.1.1. Cisplatin induced liver damage:**

133 Cisplatin administration caused a significant decrease in serum albumin concentration as well as BChE activity which
134 recorded -12.9% and -25.6% (P<0.05) as compared to the normal control level. This decrease in serum albumin and
135 BChE levels accompanied with a significant increase in serum GPT activity which calculated 35% comparing to the
136 corresponding control group Fig. (2).

137 On the other hand, a significant decrease in both liver GSH and P450 concentration that recorded -13.9% and -26.9%
138 (P<0.05) respectively as compared to the normal control level. Table (1) was indicated according to the cisplatin
139 administration.

140 In case of trace metals concentration, liver Fe, Zn as well as Cu concentration was estimated in the group of animals
141 treated with cisplatin only. Table (3) showed a non significant change on the level of Fe, Zn and Cu in cisplatin treatment
142 group when these data compared to the normal untreated control level.

143 **Table 1: Effects of cisplatin administration and/ or radiation exposure on liver GSH and 0-**
144 **cytochrome P450, 24 hours post radiation exposure (n=6)**

Treatment X ± SD	Control group	Cis group	IRR group	Cis & IRR group
GSH (mg/g tissue) X ± SD	78.94 ± 2.388	68.00 ± 2.391*	86.53 ± 2.705*(#)	76.00 ± 7.622(#)
Cytochrom P450 (pmole/ml/min) X ± SD	0.5300 ± 0.0337	0.3875 ± 0.0727*	0.6025 ± 0.0613	0.4825 ± 0.0591(#)

145 Data are presented as mean (± SD). *Significantly different from the control group (P<0.05). (#)Significantly different from the Cis.
146 group (P<0.05). IRR: Low radiation dose of 0.5 Gy gamma rays, Cis: cisplatin administration.

147 **3.1.2. Testicular toxicity due to cisplatin administration:**

148 Testicular toxicity was detected by measuring the activity of FSH in serum. Serum FSH showed a significant increase due
149 to the injection of cisplatin (10mg/kg b.wt I.P), the percentage increase in serum FSH was calculated 78%, (P<0.05) when
150 compared to the normal control level (Fig. 2).

151 Administration of cisplatin to the animal caused a significant decrease in both GSH content and P450 concentration in
 152 testis. The decrease in testis GSH recorded -23%, ($P<0.05$) and P450 (-48%, $P<0.05$) as compared to the control level
 153 (Table, 2).

154 Table (4) showed the effect of cisplatin administration on trace elements concentration, which indicated a significant
 155 decrease in Fe, Zn and Cu in testis tissue. The percentage decrease in all tested trace element concentration in testis
 156 was nearly the same (-29%, -29% and -22.4%, $P<0.05$) comparing to the normal untreated control respectively.

157 **Table 2:** Effects of cisplatin administration and/ or radiation exposure on testis GSH and
 158 cytochrome P450, 24 hours post radiation exposure ($n=6$)

Treatment $\bar{X} \pm SD$	Control group	Cis group	IRR group	Cis & IRR group
GSH (mg/g tissue) $\bar{X} \pm SD$	32.36 \pm 2.596	24.93 \pm 4.778*	39.30 \pm 4.200*(#)	35.38 \pm 3.061(#)
Cytochrom P450 (pmole/ml/min) $\bar{X} \pm SD$	0.4160 \pm 0.0427	0.2167 \pm 0.0459*	0.3780 \pm 0.0500(#)	0.4027 \pm 0.0460(#)

159 Data are presented as mean (\pm SD). *Significantly different from the control group ($P<0.05$). (#)Significantly different from the Cis.
 160 group ($P<0.05$). IRR: Low radiation dose of 0.5 Gy gamma rays, Cis: cisplatin administration.

161 **3.2. Low dose of radiation exposure showed no significant change in both liver and testis:**

162 **3.2.1. Low dose of radiation (0.5 Gy) induced no change in liver function:**

163 Exposure the animals to low dose of radiation (0.5) caused no significant changes in the concentration of serum albumin
 164 as well as the activities of GPT and BChE as compared to the normal control level (Fig. 2). Moreover, a significant
 165 increase was shown in the level of serum albumin and BChE recording 13% and 36% ($P<0.05$) comparing to the cisplatin
 166 treated group.

167 On the other hand, exposing the animals to 0.5Gy dose of whole body gamma radiation caused a significant increase in
 168 liver GSH content comparing to the normal control level. The increase in liver GSH content was calculated as 9.6% and
 169 27% as compared to the control and cisplatin treated group respectively (Table 1), while the concentration of P450
 170 showed no significant change comparing to normal control levels accompanied by a significant increase (74%, $P<0.05$) in
 171 regards to the cisplatin treated group.

172 Finally table (3) showed a non significant change in all tested trace element as compared to both control and cisplatin
 173 treated groups.

174 **3.2.2. Testicular state after low dose of gamma radiation (0.5 Gy):**

175 Exposure the animals to low dose of radiation (0.5 Gy) caused no significant changes in the concentration of serum FSH
 176 hormone as compared to the normal control level (Fig. 1). While, a significant increase was shown in the level of serum
 177 FSH that recorded 56.5% ($P<0.05$) comparing to the cisplatin treated group fig. (2).

178 In table (2) the effect of low dose of radiation showed a significant increase in testis GSH content regarding to both
 179 control untreated and cisplatin treated groups. The percentage of this increase was recorded 21.44% and 57.6%
 180 ($P<0.05$) respectively. In addition, non-significant increase was evaluated in P450 concentration due to exposing animals
 181 to low dose of radiation with a significant increase when the P450 level compared to cisplatin treated group (74%,
 182 $P<0.05$) table (2).

183 In case of trace metals concentration, testis Fe, Zn as well as Cu concentration was estimated in the group of animals
 184 exposed to the low dose of gamma radiation (0.5 Gy). Table (4) showed a non significant change on the level of Fe, Zn
 185 and Cu in low dose of radiation exposure group when these data compared to the normal untreated control level.

186
 187

188 **Table 3:** Effects of cisplatin administration and/ or radiation exposure on levels of Fe, Zn and Cu in liver tissue 24 hours post radiation
189 exposure ($n=6$)

Treatment $\bar{X} \pm SD$	Control group	Cis. Group	IRR group	Cis. & IRR group
Fe ($\mu\text{g/g}$ fresh tissue) $\bar{X} \pm SD$	205.6 \pm 34.96	183.1 \pm 14.99	186.8 \pm 17.80	190.2 \pm 21.65
Zn ($\mu\text{g/g}$ fresh tissue) $\bar{X} \pm SD$	119.7 \pm 16.34	112.9 \pm 13.48	98.86 \pm 13.20	102.3 \pm 19.92
Cu ($\mu\text{g/g}$ fresh tissue) $\bar{X} \pm SD$	3.450 \pm 0.4913	3.308 \pm 0.2731	3.322 \pm 0.2994	3.087 \pm 0.2779

190 Data are presented as mean (\pm SD). *Significantly different from the control group ($P<0.05$). (#)Significantly different from the Cis.
191 group ($P<0.05$). IRR: Low radiation dose of 0.5 Gy gamma rays, Cis: cisplatin administration.

192 **3.3. The synergistic effect of low dose of radiation exposure with cisplatin administration in both liver and**
193 **testis:**

194 **3.3.1. Low dose of radiation (0.5 Gy) induced a significant amelioration in liver function:**

195 Fig. (2) showed a full restoration in serum albumin concentration, GPT and BChE activity in the group of animals exposed
196 to low dose 0.5 Gy gamma radiation after cisplatin treatment as regarded to the normal control value. In addition, a
197 significant increase in both serum albumin and BChE levels recorded 13% and 20% ($P<0.05$) as compared to the
198 cisplatin treated group respectively. In addition; animals exposed to low dose of radiation after cisplatin (10 mg/kg b.wt.
199 I.P) treatment showed a non significant increase as compared to the serum GPT activity of cisplatin treated animals.

200 According to data in table (2) the effect of low dose of gamma rays (0.5 Gy) showed a full amelioration as in case of both
201 liver GSH and P450 concentration and their levels become more or less as the normal untreated control level. Comparing
202 the liver GSH and P450 concentration in the group of animal treated with cisplatin before exposing to gamma rays
203 together with cisplatin treated animals group showed a significant increase (11.8% and 24.5%, $P<0.05$) as in table (2).

204 Table (4) showed the effect of cisplatin administration before exposing the animals with low dose of radiation on all tested
205 trace elements concentration that indicated a non significant change in Fe, Zn and Cu in liver tissue as compared with
206 either control untreated or cisplatin treated groups.

207 **Table 4:** Effects of cisplatin administration and/ or radiation exposure on levels of Fe, Zn and Cu in testis tissue 24 hours
208 post radiation exposure ($n=6$)

Treatment $\bar{X} \pm SD$	Control group	Cis. Group	IRR group	Cis. & IRR group
Fe ($\mu\text{g/g}$ fresh tissue) $\bar{X} \pm SD$	97.91 \pm 16.86	69.61 \pm 4.575*	87.09 \pm 6.388	96.38 \pm 21.22(#)
Zn ($\mu\text{g/g}$ fresh tissue) $\bar{X} \pm SD$	128.8 \pm 23.55	91.57 \pm 6.143*	106.6 \pm 13.13	116.7 \pm 21.26
Cu ($\mu\text{g/g}$ fresh tissue) $\bar{X} \pm SD$	1.885 \pm 0.3851	1.462 \pm 0.1201*	1.603 \pm 0.2568	1.603 \pm 0.1675

209 Data are presented as mean (\pm SD). *Significantly different from the control group ($P<0.05$). (#)Significantly different from the Cis.
210 group ($P<0.05$). IRR: Low radiation dose of 0.5 Gy gamma rays, Cis.: cisplatin administration.

211 **3.3.2. Low dose of radiation (0.5 Gy) and its effect on the testis tissue after cisplatin treatment:**

212 Exposing cisplatin treated animals to low dose of gamma radiation (0.5 Gy) caused a non- significant decrease in serum
213 FSH activity as compared to the normal control level that means a significant decrease as compared to the cisplatin
214 treated animals only, which calculated 24% Fig. (2). In addition, the effect of whole body gamma radiation (0.5 Gy) on the
215 animals treated with cisplatin only caused a significant increase in testis GSH content comparing to the cisplatin treated

216 group. This increase was calculated as 42% (Table 2) haunt with a non significant change when compared to the normal
217 untreated group, while the concentration of P450 showed no significant change comparing to normal control levels
218 accompanied by a significant increase (68%, $P<0.05$) with regards to the cisplatin treated group.

219 In case of trace metals concentration, testis Fe, Zn as well as Cu concentration was estimated in the group of animals
220 treated with cisplatin before exposing to the low dose of gamma radiation (0.5 Gy). Table (3) showed a significant change
221 on the level of Fe, Zn and Cu in cisplatin treatment group when these data compared with either normal untreated control
222 or cisplatin treated one.

223 4. Discussion

224 Cisplatin is the most widely used antitumor drug especially in the treatment for testicular cancer, but its usage is limited
225 by toxic effects on the reproductive system [32]. Testicular damage induced by cisplatin treatment was characterized by
226 significant decreases in plasma testosterone level [33] that may be due to the increase in FSH level as in the current
227 work, since according to Sasson and his colleagues the gonadal failure, which caused by cisplatin treatment, leads to
228 high levels of FSH and LH [34].

229 Once cisplatin has been administered in the body, it rapidly diffuses into tissues and is highly bound to plasma albumin
230 and other protein [35]. This may explain why cisplatin caused a decrease in serum albumin in the present work. In
231 addition, the decrease in both serum albumin concentration as well as butyryle cholinesterase with the increase in serum
232 GPT activities are further evidence that anti-tumoral treatment causes liver damage [36]. According to the observation of
233 Cepeda and his colleagues, there is a direct connection between the cellular concentrations of copper and platinum
234 recommended an active transport for cisplatin through copper transporter, which reduce the uptake of each other that
235 indicated as a significant decrease in testicular copper levels together with Zn and iron that may be due to cisplatin
236 treatment induced oxidative stress in testis [2]. Consequently, increase in lipid peroxidation (LPO) and depletion of
237 enzymes such as superoxide dismutase following cisplatin treatment in testis and liver clearly demonstrated failure of
238 their antioxidant defence system [33]. It may be noted that previous investigators reported the reduction of GSH levels
239 leads to elevation of LPO [37 & 38]. Therefore, depletion of GSH as in the present results might be inducing oxidative
240 stress by increasing the free radical generation leading to cell death in both tested organs.

241 Cytochrome p450 is the catalyst enzymes that are essential for the metabolism of many medications. Suppression of
242 P450 may be due to the consequence of inflammatory processes and can result in increased clinical toxicity of drugs with
243 a low therapeutic index [39]. Down regulation of testicular cytochrome P450 reported by Waxman and Change and
244 Masubuchi and his colleagues that was indicated in the present results [40 & 41]. However, exposing to low radiation
245 dose attenuated the level of P450 after cisplatin treatments. In addition, evidence reported by Oetari and his colleagues
246 indicates inactivation of glutathione- S- transferase accompanied by glutathione depletion and inhibition of cytochrome
247 p450 in liver [42]. Interestingly, the cytochrome p450 and GSH content in both liver and testis showed a significant
248 inhibition than the control group in our current study. Moreover, the active site of cytochrome p450 contain iron heme
249 center [43], so as one could explain the decrease in both cytochrome p450 and iron in testis as in present work.

250 Cisplatin augments the antitumor effect of a cytotoxic T-lymphocyte-mediated immunotherapy strategy, resulting in a
251 higher cure rate. This effect is associated with the enhanced ability of cytotoxic T lymphocytes to lyse tumor cells [44]. It
252 seems that functional activity of lymphocytes decreased at the stage of well-developed tumor, which promoted inhibition
253 of the lymphocyte defense properties. Cisplatin did not modify the structure and functions of lymphocytes and presumably
254 improved their energy status [45]. So to overcome the above problems and as the evidence of Murphy and Morton [22],
255 exposing to the low radiation dose can be used as a synergistic agent with cisplatin for cancer treatment since low
256 radiation dose administration to entire body increased the action of the protective process in living organism including the
257 overproduction lymphocytes that significantly prevented or impaired tumor growth. In addition, an interesting observation
258 remarked that sign of glutathione (GSH) protects against cisplatin cytotoxicity [46]. Of note is that the GSH administration
259 achieved a superior response rate despite having a larger average tumor burden during chemotherapy [47]. According to
260 current study animals exposed to low radiation dose alone (0.5 Gy) showed a significant increase in GSH content in both
261 liver and testis tissue, which may explain the antioxidant effects of low radiation dose against cisplatin induced oxidative
262 stress. These antioxidants did not interfere with chemotherapeutic effects - and mitigated cancer treatment toxicity [48].

263

264

265 **5. Conclusion:**

266 In conclusion, the obtainable results proved that low radiation dose have the existence as an antioxidant and antitumor
 267 agents which may be useful to use as a synergistic agents with the chemotherapeutic drug or even in case of tumors
 268 chemo-radiotherapy. Further studies should be needed to address this issue in different biochemical views.

269

270 **6. REFERENCES**

- 271 1. Giaccone, G. Clinical perspectives on platinum resistance. *Drugs*. 2000; 59 (Suppl 4): 9–17.
- 272 2. Cepeda, V, Miguel, AF, Castilla, J, Alonso, C, Quevedo, C, and Pérez, JM. Biochemical Mechanisms of
 273 Cisplatin Cytotoxicity, Anti-Cancer Agents in Medicinal Chemistry. 2007; 7: 3-18
- 274 3. Chirino, YI, Pedraza- Chaverri, J. Role of oxidative and nitrosative stress in cisplatin-induced nephrotoxicity.
 275 *Exp Toxicol Pathol*. 2009; 61(3):223-42. doi: 10.1016/j.etp.2008.09.003. Epub 2008 Nov 4.
- 276 4. Hassan, I, Chibber, S., Khan, AA, Naseem, I. Riboflavin Ameliorates Cisplatin Induced Toxicities under
 277 Photoillumination. *PLoS ONE*. 2012; 7(5): e36273.
- 278 5. González, VM, Fuertes, MA, Alonso, C, Pérez, JM. *Mol. Pharmacol*. 2001; 59: 657.
- 279 6. Sadowitz, PD, Hubbard, BA, Dabrowiak, JC, Goodisman, J, Tacka, KA, Aktas, MK, Cunningham, MJ, Dubowy,
 280 RL, Souid, AK. Kinetics of cisplatin binding to cellular DNA and modulations by thiolblocking agents and thiol
 281 drugs. *Drug Metab. Dispos*. 2002; 30: 183–190.
- 282 7. Coia, L, Won, M, Lanciano, R, Marcial, VA, Martz, M, Hanks, G. The patterns of care outcome study for cancer
 283 of the uterine cancer: Results of the second national practice survey. *Cancer*. 1990; 66: 2451-2456.
- 284 8. Morri, y, Shara, H, Shima, H, Shimada, K, Arima, M, Ikoma, F. Combined cisplatin and radiation therapy in
 285 patients with invasive bladder cancer. *International Urology and Nephrology*. 1990; 22(4): 337- 343.
- 286 9. Wodinsky, I, Swiniarki, J, Kensler, CJ, Venditti, JM. Combination radiotherapy and chemotherapy for P388
 287 lymphocytic leukemia in vivo *Cancer Chemother. Rep*. 1974; 4: 73.
- 288 10. Morris, CR, Blackwell, LH, Loveless, VS. Antileukemic of combination of radiation and malanats (1,2
 289 diamininocyclohexane) platinum II (NSC. 224964) *J. Med*. 1977; 8: 254 (1977)
- 290 11. Omura, GA. (1996) Chemotherapy for stage IVB or recurrent cancer of the uterine cervix. *Journal of the*
 291 *National Cancer Institute*. 1996; 21:123–6.
- 292 12. Wallner, KE, Li, GC. Effect of cisplatin resistance on cellular radiation response. *Int J Radiat Onco Biol Phys*.
 293 1987; 13:587–91.
- 294 13. Perez, CA, Grigsby, PW, Castro, Vita, H, Lockett, MA. (1995) Carcinoma of the uterine cervix. I. Impact of
 295 prolongation of treatment time and timing of brachytherapy on outcome of radiation therapy. *Int J Radiat Onco*
 296 *Biol Phys*. 32:1275–88.
- 297 14. Green, JA, Kirwan, JM, Tierney, JF, *et al*. Survival and recurrence after concomitant chemotherapy and
 298 radiotherapy for cancer of the uterine cervix: a systematic review and meta-analysis. *Lancet*. 2001; 358: 781-6.
- 299 15. Ishizuka, M., Fujimoto, Y, Itoh, Y, Kitagawa, K, Sano, M, Miyagawa, Y, Ando, A, Hiramatsu, M, Hirasawa, N,
 300 Ishihara, S, Nakashima, T, Yamada, K. Relationship between hematotoxicity and serum albumin level in the
 301 treatment of head and neck cancers with concurrent chemo-radiotherapy using cisplatin. *Jpn. J. Oncol*. 2011;
 302 41(8): 973- 979.
- 303 16. Nogami, M, Huang, JT, James, SJ. Mice chronically exposed to low dose ionizing radiation possess
 304 splenocytes with increased levels of HSP70 mRNA, HSC72 and HSP70 and with an increased capacity to
 305 proliferate. *Int J Radiat Biol*. 1993; 63:775–783.

- 306 17. Kojima, S, Nakayama, K, Ishida, H. Low Dose γ -Rays Activate Immune Functions via Induction of Glutathione
307 and Delay Tumor Growth. *J Radiat Res.* 2004; 45 (1): 33-39. doi: 10.1269/jrr.45.33
- 308 18. Feinendegen, LF. Evidence for beneficial low level radiation effects and radiation hormesis. *Brit J Radiol.* 2005;
309 78: 3-7. doi:10.1259/bjr/63353075
- 310 19. Gharib, OA, Abd Ellatif, UA, Abdellah, NM, Mohammad, MA. Radio-protective response on the environmental
311 pollutant induced oxidative stress. *Adv Bioscie Biotech.* 2012; 3: 989-996.
- 312 20. Kopp, AH, Eifel, PJ. Chemo-radiotherapy for cervical in 2010. *Curr Oncol. Rep.* 2011; 13:77-85.
- 313 21. Turner, J.E. (1995). *Atoms radiation and radiation protection (2nd Edition)*, John Willy & Sons, inc., New York.
- 314 22. Murphy, B, Morton, JJ. *J. Exper.* 1915; *Med.* 22: 204.
- 315 23. Barcellos-Hoff, MH, Park, C, Wright, EG. Radiation and the microenvironment - tumorigenesis and therapy. *Nat*
316 *Rev Cancer.* 2005; 5: 867-875.
- 317 24. Máthé, A, Komka, K, Forczig, M, Szabó, D, Anderlik, P, Rozgonyi, F. The effect of different doses of cisplatin
318 on the pharmacokinetic parameters of cefepime in mice. *Lab Anim.* 2006; 40:296-300
- 319 25. Munshaw, S, Hwang, HS, Torbenson, M, Quinn, J, Hansen, KD, Astemborski, J, Mehta, SH, Ray, SC, Thomas,
320 DL, Balagopal, A. Laser captured hepatocytes show association of butyrylcholinesterase gene loss and fibrosis
321 progression in hepatitis C-infected drug users. *Hepatology.* 2012; 56(2):544-54. doi: 10.1002/hep.25655.
- 322 26. Doumas, BT, Watson, WA, Biggs, HG. *Clin. Chim.Acta.* 1971; 31: 87-95
- 323 27. Beutler, E. *Red Cell Metabolism: A Manual of Biochemical Methods*, Grune and Stratton, New York. 1971.
- 324 28. Nerurkar, PV, Park, SS, Thomas, PE, Nims, RW, Lubet, RA. Methoxyresorufin and benzyloxyresorufin:
325 substrates preferentially metabolized by cytochromes P4501A2 and 2B, respectively, in the rat and mouse.
326 *Biochem Pharmacol.* 1993; 46: 933-943.
- 327 29. IAEA. *Elemental analysis of biological materials. Current problem and techniques with special reference to*
328 *trace elements.* International Atomic Energy Agency, IAEA, Vienna. Technical Reports Series. 1980; 379.
- 329 30. Kingstone, HM, Jassie, L. *Introduction to microwave sample preparation. Theory and practice.* American
330 *Chemical Society Professional Reference Book*, Washington DC. 1998; 263.
- 331 31. Gregus, Z, Klaassen, CD. Disposition of metals in rats: A comparative study of fecal urinary and biliary
332 excretion and tissue distribution of eighteen metals. *Toxicology and Applied Pharmacology.* 1986; 85: 24-38.
333 doi:10.1016/0041-008X(86)90384-4
- 334 32. Ciftci, O, Beytur, A, Cakir, O, Gurbuz, N, Vardi, N. Comparison of Reproductive Toxicity Caused by Cisplatin
335 and Novel Platinum-N-Heterocyclic Carbene Complex in Male Rats. *Basic & Clinical Pharmacology &*
336 *Toxicology.* 2011; 109: 328-333.
- 337 33. Fahmy, HA, Abd El-Azime, ASh, Gharib, OA. Possible Ameliorative Role of Low Dose of Radiation Against
338 Cisplatin Induced Oxidative Stress and Tissue Damage in Male Rats. *Euro J Biol Medi Sci Resh.* 2013; 1(4):
339 10-18.
340
- 341 34. Sasson, R, Dantes, A, Tajima, K, Amsterdam, A. Novel genes modulated by FSH in normal and immortalized
342 FSH-responsive cells: new insights into the mechanism of FSH action. *FASEB J.* 2003; 17(10):1256-66.
- 343 35. Judson, I, Kelland, LR. *New developments and approaches in the platinum arena.* 2000; 59: 29.
- 344

- 345 36. Fasihi, M, Ghodrati-zadeh, S, Ghodrati-zadeh, S. Protective Effect of Captopril on Cisplatin Induced
346 Hepatotoxicity in Rat. *American-Eurasian. J Toxicol Sci.* 2012; 4(3): 131-134.
- 347 37. El-Maraghy, SA, Gad, MZ, Fahim, AT, Hamdy, M A. Effect of cadmium and aluminum intake on the antioxidant
348 status and lipid peroxidation in rat tissues. 2001; *J Biochem Mol Toxicol.* 2001; 15(4):207-14.
- 349 38. Gharib, OA, Ibrahim, NK. Oxidative damage in testes induced by 950 MHz simulating cellular phone. *Isotope
350 and Radiat Res.* 2010; 42(4) , 9 4 1 - 9 5 3.
- 351 39. Morgan, ET. Regulation of cytochromes P450 during inflammation and infection. *Drug Metab Rev.* 1997;
352 29:1129-1188.
- 353 40. Waxman, DJ, Chang, TKH. Hormonal regulation of liver cytochrome P450 enzymes. In *Cytochrome P450:
354 Structure, Mechanism, and Biochemistry* (2nd edition). Ortiz de Montellano P. R., editor. Plenum Press, New
355 York. 1995; 391-418.
- 356 41. Masubuchi, Y, Enoki, K, Horie, T. Down-regulation of hepatic cytochrome P450 enzymes in rats with
357 trinitrobenzene sulfonic acid-induced colitis *Drug Metab Dispos.* 2008; 36(3):597-603.
- 358 42. Oetari, S, Sudibyo, M, Commandeur, JN, Samhoedi, R, Vermeulen, NP. Effects of curcumin on cytochrome
359 P450 and glutathione S-transferase activities in rat liver. *Biochem Pharmacol.* 1996; 12,51(1):39-45.
- 360 43. Guengerich, FP. "Cytochrome p450 and chemical toxicology". *Chem. Res. Toxicol.* 2008; 21 (1): 70-83.
- 361 44. Merritt, RE, Mahtabifard, A, Yamada, RE, Crystal, RG, Korst, RJ. (2003) Cisplatin augments cytotoxic T-
362 lymphocyte-mediated antitumor immunity in poorly immunogenic murine lung cancer. *J Thorac Cardiovasc
363 Surg.* 2003; 126(5):1609-17.
- 364 45. Zamay, TN, Kolovskaya, OS, Zamay, GS, Borodina, NA. Effects of Cisplatin on Lymphocyte Structure and
365 Functions in Mice with Ehrlich Ascitic Carcinoma. *Bull Experi Biol & Medi.* 2011; 151 (1): 62-65.
- 366 46. Lu, y, Kawashima, A, Horri, I, Zhong, L. Effect of BSO and L- cysteine on drug induced cytotoxicity in primary
367 cell cultures: drug-, cell type- and species- specific difference. *Drug Chem. Toxicol.* 2004; 27: 269- 280.
- 368 47. Schmidinger, M, Budinsky, AC, Wenzel, C, Piribauer, M, Brix, R, Kautzky, M, Oder, W, Locker, GJ, Zielinski,
369 CC, Steger, GG. Glutathione in the prevention of cisplatin induced toxicities. A prospectively randomized pilot
370 trial in patients with head and neck cancer and non small lung cancer. *Wien Klin Wochenschr.* 2000; 112(14):
371 617-23.
- 372 48. Block, KI, Koch, AC, Mead, MN, Tothy, PK, Newman, RA, Gyllenhaal, C. Impact of antioxidant supplementation
373 on chemotherapeutic efficacy: A systematic review of the evidence from randomized controlled trials, *Cancer
374 Treatment Reviews.* 2007. doi:10.1016/j.ctrv.2007.01.005.