

# Effect of low radiation dose on cisplatin induced hepato-testicular damage in male rats.

## ABSTRACT

**Aim:** The aim of this study was to evaluate the synergistic effect of low radiation dose with the chemotherapeutic drugs in order to find possible way to lessen the harmful effects during chemo-radiotherapy.

**Study Design:** Randomized controlled experiment.

**Place and Duration of Study:** Experimental Animal Unit, Drug Radiation Research Department, National Center for Radiation Research and Technology, Cairo Egypt.

**Methodology:** Estimation of antioxidant activity of low radiation dose on oxidative stress induced by cisplatin administration at a dose of (10 mg/kg bwt) in male albino rat.

**Results:** Results of experiment revealed that cisplatin administration caused a significant increase in serum alanine transaminase (GPT) activity ( $38.58 \pm 2.060$ ) and FSH level ( $8.162 \pm 1.424$ ) accompanied with a decrease in serum albumin ( $3.492 \pm 0.253$ ), and Butyryl Cholein Esterase (BChE) ( $65.35 \pm 12.61$ ). In Liver and testis, GSH content ( $68.00 \pm 2.391$  &  $24.93 \pm 4.778$ ) as well as cytochromes P450 levels ( $0.3875 \pm 0.0727$  &  $0.2167 \pm 0.0459$ ) showed a significant decrease as compared to the normal control level respectively. In addition the level of Fe, Cu and Zn showed no significant changes in liver and appeared to be significantly decrease as in case of corresponding trace elements in testis organs. On the other hand, exposing to low dose of radiation (0.5 Gy) post-cisplatin treatment effectively prevented these alterations and maintained the antioxidant status.

**Conclusion:** Data from present results revealed that low radiation dose have the existence as an antioxidant and antitumor agents which may be useful to use as a synergistic agents with the chemotherapeutic drug.

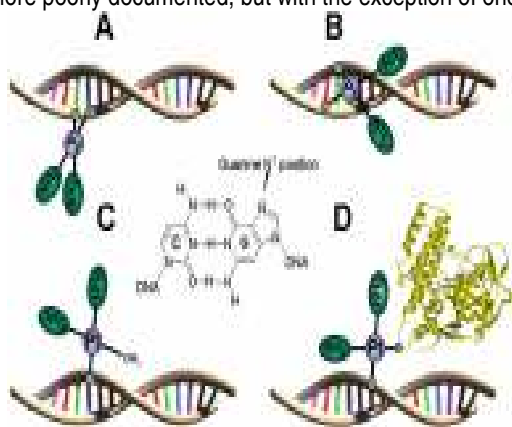
*Keywords: Low Radiation Dose, Cisplatin, Cytochromes P450, Butyryl Cholein Esterase.*

## 1. INTRODUCTION

Cisplatin is one of the most potent anticancer drugs used in chemotherapy. It is highly effective in the treatment of testicular and ovarian cancers and is also widely employed for treating bladder, cervical, head and neck, esophageal and small cell lung cancer [1]. Despite being an effective anti-proliferative agent, the clinical usage of cisplatin is limited by various side effects including nephro-toxicity, hepato-toxicity, neurotoxicity and oto-toxicity. The anticancer activity of this drug is attributed to its capacity to form covalent bond at N-7 position of Guanine residues of DNA leading to formation of 1, 2 or 1,3 inter-strand crosslink and a lesser extent of the interstrand crosslink (Fig. 1). These adduct cisplatin DNA disrupts the cellular replication and transcription machinery [2]. Many labs have demonstrated that cisplatin generates free radicals leading to oxidative and nitrosative stress which results into such deleterious effects *in vivo* [3, 4]. It is generally accepted that binding of cisplatin to genomic DNA (gDNA) in the cell nucleus is the main event responsible for its antitumor properties [5]. Thus, the damage induced upon binding of cisplatin to gDNA may inhibit transcription, and/or DNA replication mechanisms. Subsequently, these alterations in DNA processing would trigger cytotoxic processes that lead to cancer cell death. The cytotoxicity of cisplatin is considered to be due to a combination of factors, including peroxidation of the cell membrane, mitochondrial dysfunction, inhibition of protein synthesis, and DNA injury [6].

Till the last decade of the last century radiotherapy was the only therapeutic option for patients with locally advanced cancer [7]. There are some trails of combined cisplatin and radiation therapies have been reported and it has proved to be hopeful method to improve the treatment results of invasive bladder cancer [8]. The effectiveness of the combination of cisplatin and radiation in experimental malignant tumor is reported by Wodinsky and his colleagues [9] in leukaemic cell. Morris and his colleagues noted a similar effect in mouse leukaemia [10]. Cytotoxic chemotherapy has been shown to give good response rates in patients with good kidney function and no prior radiation therapy. Cisplatin is the most effective single agent [11], and has been shown in cell lines to be synergistic with radiotherapy. Mechanisms underlying the interaction between drugs and radiation may include inhibition of potentially lethal or sublethal damage repair, and increasing radio-sensitivity of hypoxic cells [12]. It has been widely used prior to surgery or radiotherapy with the aim of reducing tumour volume and facilitating local treatment. It may have the additional benefit of controlling micrometastatic

32 disease. Combined chemo-radiotherapy seems to offer substantial benefit for women with cervical cancer. However,  
33 acute toxicity, predominantly haematological and gastrointestinal, was increased with chemo-radiation [13]. Acute side  
34 effects are generally of short duration and resolve with medical management, while the late complications of radiotherapy  
35 lead to damage which can be difficult to reverse, and may permanently impair quality of life. Details of late morbidity are  
36 more poorly documented, but with the exception of one trial [14].



**Fig. (1)** Main adducts formed after binding of cisplatin to DNA.  
(A) 1,2-intrastrand cross-link, (B) interstrand cross-link,  
(C) mono- functional adduct, and (D) protein-DNA cross-  
link. The main site of attack of cisplatin to DNA (N7 of  
guanine) is shown in the central panel [2].

49 Low dose of radiation induce various effects including radio-protective response [15], as well as activation of immune  
50 function [16]. It has been well documented that the immune function is linked to the release of radical oxygen species  
51 (ROS). Access amount of ROS is commonly eliminated by endogenous antioxidant system, thereby preventing injury to  
52 DNA [17]. It exerts protection against other challenges involving radicals and causing a beneficial effect by temporarily  
53 shielding the hit cell against radicals produced through endogenous processes. Low dose of radiation has been observed  
54 to stimulate the radical detoxification system, enhancement of DNA repair rates and induce immune competence that  
55 associate with an increase in number of cytotoxic lymphocytes, even causing a reduction of the incidence of metastatic  
56 cancer [18]. Our pervious study indicated that treatment with low dose of gamma rays (0.5 Gy) ameliorate harmful effects  
57 induced by TCE due to the effect of gamma radiation as a stimulant of radical detoxification [19].

58 According to many clinical studies, when chemotherapy and radiotherapy are concurrently administered improve  
59 effectively greater rather than at different times [20]. Combined chemo-radiotherapy presents some problems; however,  
60 with more sever adverse events, resulting in a reduced treatment compilation rate [21]. On the other hand, according to  
61 the study of Murphy and Morton [22] and Barcellos-Hoff, [23], low radiation dose administartion to entire body increased  
62 the action of the protective process in living organism including the overproduction lymphocytes that significantly  
63 prevented or impaired tumor growth. From this point of view, the current study's aim is to investigate the synergistic effect  
64 of low radiation dose in the treatment with cisplatin as a chemotherapeutic drug inducing oxidative stress.

## 65 2. MATERIAL AND METHODS

### 66 2.1. Animals

67 Male albino rats weighing approximately 120-150 g were used for this experiment. They were housed in polypropylene  
68 cages in an air conditioned room with temperature maintained at  $25^{\circ}\text{C} \pm 3^{\circ}\text{C}$ , relative humidity of  $50\% \pm 5\%$  and 12 h  
69 alternating light and dark cycles. The rats were provided with a nutritionally adequate chow diet and drinking water *ad*  
70 *libitum* throughout the study. Experiments were begun after a four-week acclimatization period.

### 72 2.2. Chemicals

73 Cisplatin was purchased from Central Drug House (Egypt). The chemicals used in this experiment were obtained from  
74 Sigma Chemical (USA). Kits used in this experiments were purchased from Bio-Diagnostics (UK).

### 75 2.3. Radiation process:

76 A single dose whole body irradiation (0.5 Gy) was performed with rats, using gamma rays by Cesium 137 irradiation unit,  
77 National Center for Radiation Research and Technology (NCRRT), with the dose rate 0.74589 rad/ sec. The gamma  
78 cesium cell was calibrated by alanine dosimetry relative to a primary standard. Correction were made daily for humidity,  
79 temperature, and barometric pressure.

### 80 2.4. Experimental design

81 Adult male albino rats were divided into 4 groups of 6 rats each and treated as follows:  
82 **Control group** (normal, untreated), received distilled water. **Cis. group**, received freshly dissolved cisplatin in 1 ml  
83 distilled water at a dose of 10 mg/ kg body weight (I.P); **IRR group** was exposed to a single low gamma radiation dose of  
84 0.5 Gy and **Cis. & IRR group** was administered with Cisplatin, 24 hr after cisplatin administration the animals were  
85 exposed to a single low dose of gamma rays (0.4 Gy). The dose of cisplatin was decided on the basis of Máthé and his  
86 colleagues [24]. Twenty four hours after radiation exposure the animals were decapitated and the blood was collected for  
87 biochemical analysis. Liver and testes were divided into two portions. One portion were excised, homogenized in ice-cold  
88 saline and utilized for various biochemical analyses and the second portion was used for trace elements analysis.

### 89 **2.5. Biochemical analysis**

90 The blood samples were collected directly from the animals by heart puncturing. They were centrifuged at 3000 rpm for  
91 15 min., clear sera were collected and stored in a refrigerator. The activities of GPT, butyryl cholinesterase (BChE) and  
92 FSH levels as well as concentrations of albumin in serum were analyzed. Liver and testis were minced and homogenized  
93 (10 % w/ v) in ice-cold normal saline solution. The homogenate was centrifuged at 3,000 rpm for 15-20 min at 4 °C. The  
94 resultant supernatant was used for estimation of GSH content and cytochrome P450 activity. Serum  
95 butyrylcholinesterase activity is quantitatively measured according to the methods published in Munshaw and his  
96 colleagues [25]. FSH level was detected in serum samples using elisa kit supplied by (Kamiya, biomedical company,  
97 USA) according to manufacturer instruction. The elisa kit was read using 2100 elisa reader. Serum albumin level was  
98 determined according to the method of Doumas and his colleagues [26]. In addition GSH content in both liver and testis  
99 was measured by method of Beutler and his colleagues [27]. Liver and testis cytochrome P450 activity assayed is  
100 detected using a spectrofluorometer [28]. Serum butyrylcholinesterase and FSH activities as well as liver and testis P450  
101 were carried out at the Central Laboratory, Radioisotope Dept., AEA, Giza, Egypt, while serum GPT activity and Albumin  
102 concentration, as well as tissue GSH content were performed with a Helios Thermo-Spectronic spectrophotometer  
103 (Thermo Spectronic, UK).

### 104 **2.6. Atomic absorption analysis**

105 Liver and testes tissues were digested in a mixture of conc. HNO<sub>3</sub> and H<sub>2</sub>O<sub>2</sub> (5:1). The digestion was completely by using  
106 micro-wave oven (mls- 1200 mega) and the digested sample were diluted with de-ionized water to a fixed volume [29].  
107 The selected element was estimated quantitatively by atomic absorption spectrophotometer. *Unicam 939* Hallow cathode  
108 lamps were used to determine Fe, Cu and Zn (MT) in air acetylene flame [30]. Concentration of elements in tissues was  
109 calculated by using calibration curve prepared from their stock solution (1mg). The concentration of elements per grams  
110 tissues could be determined by equation:

$$111 \text{Elemental content in the tissue } (\mu\text{g/g}) = c \cdot v \cdot n / m,$$

112 Where: *c* (μg/ml) is the concentration of the metal measured in the AAS sample of volume *v* (ml), *n* is the dilution factor  
113 and *m* (g) is the mass of the tissue taken. Wet tissue weights were used for calculating the metal concentration in tissues  
114 [31].

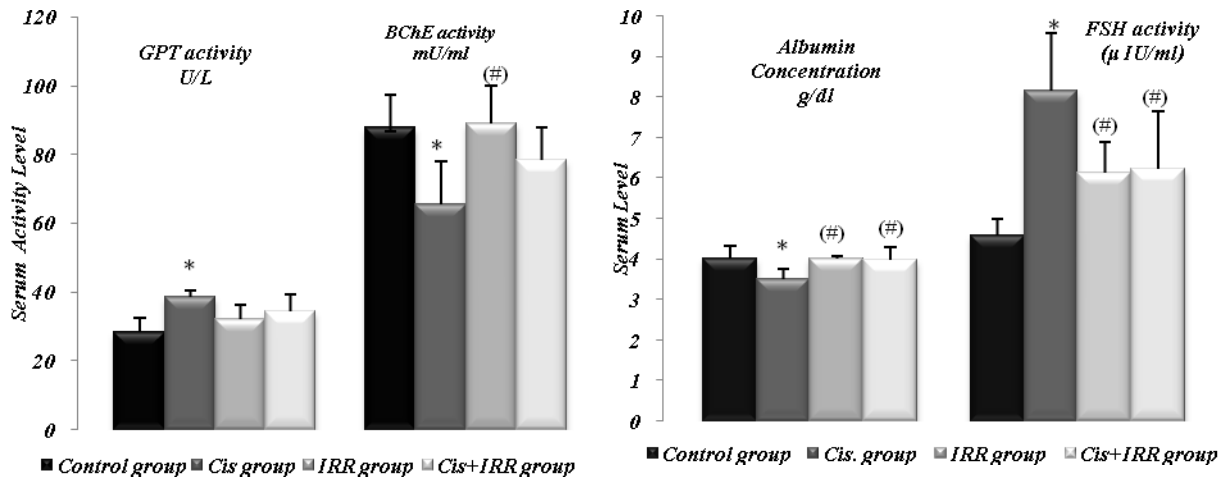
### 116 **2.7. Statistical Analysis:**

117 To assess the significant level of influence caused by low dose of radiation in cisplatin administrated rats, one way  
118 analysis of variances (ANOVA) followed by Tukey's multiple comparison test was used. Data are representative of 8  
119 independent experiments carried out in triplicate. Statistical analysis was performed by using Graph-Pad software, San  
120 Diago, CA, USA) Differences were considered statistically significant when the P value was less than 0.05.

## 122 **3. RESULTS**

123 Cisplatin is one of the most potent anticancer drugs used for the treatment of different types of cancer including testicular  
124 cancer. In spite of its significant anticancer activity, the clinical use of cisplatin is often limited by its undesirable side  
125 effect due to the influence of oxidative stress. Results indicated in fig. (2) in addition to table (1-4) evaluate the effect of  
126 cisplatin and the synergistic effect of low irradiation dose (0.5 Gy) on liver and testis tissues.

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**Fig. 2:** Effects of cisplatin administration and/ or radiation exposure on some serum biochemical parameters 24 hours post radiation exposure (n=6)

Data are presented as mean (± SD). \* Significantly different from the control group (P<0.05). (#)Significantly different from the Cis. group (P<0.05). IRR: Low radiation dose of 0.5 Gy gamma rays, Cis group: cisplatin administration.

### 133 3.1. Cisplatin demonstrated toxicity in both liver and testis:

#### 134 3.1.1. Cisplatin induced liver damage:

135 Cisplatin administration caused a significant decrease in serum albumin concentration as well as BChE activity which  
136 recorded -12.9% and -25.6% (P<0.05) as compared to the normal control level. This decrease in serum albumin and  
137 BChE levels accompanied with a significant increase in serum GPT activity which calculated 35% comparing to the  
138 corresponding control group Fig. (2).

139 On the other hand, a significant decrease in both liver GSH and P450 concentration that recorded -13.9% and -26.9%  
140 (P<0.05) respectively as compared to the normal control level. Table (1) was indicated according to the cisplatin  
141 administration.

142 In case of trace metals concentration, liver Fe, Zn as well as Cu concentration was estimated in the group of animals  
143 treated with cisplatin only. Table (3) showed a non significant change on the level of Fe, Zn and Cu in cisplatin treatment  
144 group when these data compared to the normal untreated control level.

145 **Table 1: Effects of cisplatin administration and/ or radiation exposure on liver GSH and 0-**  
146 **cytochrome P450, 24 hours post radiation exposure (n=6)**

Treatment X ± SD	Control group	Cis group	IRR group	Cis & IRR group
GSH (mg/g tissue) X ± SD	78.94 ± 2.4	68.00 ± 2.4*	86.53 ± 2.7*(#)	76.00 ± 7.6(#)
Cytochrom P450 (pmole/ml/min) X ± SD	0.5300 ± 0.034	0.3875 ± 0.073	0.6025 ± 0.061	0.4825 ± 0.059(#)

147 Data are presented as mean (± SD). \*Significantly different from the control group (P<0.05). (#)Significantly different from the Cis.  
148 group (P<0.05). IRR: Low radiation dose of 0.5 Gy gamma rays, Cis: cisplatin administration.

#### 149 3.1.2. Testicular toxicity due to cisplatin administration:

150 Testicular toxicity was detected by measuring the activity of FSH in serum. Serum FSH showed a significant increase due  
151 to the injection of cisplatin (10mg/kg b.wt I.P), the percentage increase in serum FSH was calculated 78%, (P<0.05) when  
152 compared to the normal control level (Fig. 2).

153 Administration of cisplatin to the animal caused a significant decrease in both GSH content and P450 concentration in  
 154 testis. The decrease in testis GSH recorded -23%, ( $P<0.05$ ) and P450 (-48%,  $P<0.05$ ) as compared to the control level  
 155 (Table, 2).

156 Table (4) showed the effect of cisplatin administration on trace elements concentration, which indicated a significant  
 157 decrease in Fe, Zn and Cu in testis tissue. The percentage decrease in all tested trace element concentration in testis  
 158 was nearly the same (-29%, -29% and -22.4%,  $P<0.05$ ) comparing to the normal untreated control respectively.

159 **Table 2:** Effects of cisplatin administration and/ or radiation exposure on testis GSH and  
 160 cytochrome P450, 24 hours post radiation exposure ( $n=6$ )

<i>Treatment</i> <i>X ± SD</i>	<i>Control group</i>	<i>Cis group</i>	<i>IRR group</i>	<i>Cis &amp; IRR group</i>
GSH (mg/g tissue) <i>X ± SD</i>	32.36 ± 2.6	24.93 ± 4.8*	39.30 ± 4.2*(#)	35.38 ± 3.1(#)
Cytochrom P450 (pmole/ml/min) <i>X ± SD</i>	0.4160 ± 0.043	0.2167 ± 0.046*	0.3780 ± 0.05(#)	0.4027 ± 0.046(#)

161 Data are presented as mean ( $\pm$  SD). \*Significantly different from the control group ( $P<0.05$ ). (#)Significantly different from the *Cis*.  
 162 group ( $P<0.05$ ). IRR: Low radiation dose of 0.5 Gy gamma rays, *Cis*: cisplatin administration.

163 **3.2. Low dose of radiation exposure showed no significant change in both liver and testis:**

164 **3.2.1. Low dose of radiation (0.5 Gy) induced no change in liver function:**

165 Exposure the animals to low dose of radiation (0.5) caused no significant changes in the concentration of serum albumin  
 166 as well as the activities of GPT and BChE as compared to the normal control level (Fig. 2). Moreover, a significant  
 167 increase was shown in the level of serum albumin and BChE recording 13% and 36% ( $P<0.05$ ) comparing to the cisplatin  
 168 treated group.

169 On the other hand, exposing the animals to 0.5Gy dose of whole body gamma radiation caused a significant increase in  
 170 liver GSH content comparing to the normal control level. The increase in liver GSH content was calculated as 9.6% and  
 171 27% as compared to the control and cisplatin treated group respectively (Table 1), while the concentration of P450  
 172 showed no significant change comparing to normal control levels accompanied by a significant increase (74%,  $P<0.05$ ) in  
 173 regards to the cisplatin treated group.

174 Finally table (3) showed a non significant change in all tested trace element as compared to both control and cisplatin  
 175 treated groups.

176 **3.2.2. Testicular state after low dose of gamma radiation (0.5 Gy):**

177 Exposure the animals to low dose of radiation (0.5 Gy) caused no significant changes in the concentration of serum FSH  
 178 hormone as compared to the normal control level (Fig. 1). While, a significant increase was shown in the level of serum  
 179 FSH that recorded 56.5% ( $P<0.05$ ) comparing to the cisplatin treated group fig. (2).

180 In table (2) the effect of low dose of radiation showed a significant increase in testis GSH content regarding to both  
 181 control untreated and cisplatin treated groups. The percentage of this increase was recorded 21.44% and 57.6%  
 182 ( $P<0.05$ ) respectively. In addition, non-significant increase was evaluated in P450 concentration due to exposing animals  
 183 to low dose of radiation with a significant increase when the P450 level compared to cisplatin treated group (74%,  
 184  $P<0.05$ ) table (2).

185 In case of trace metals concentration, testis Fe, Zn as well as Cu concentration was estimated in the group of animals  
 186 exposed to the low dose of gamma radiation (0.5 Gy). Table (4) showed a non significant change on the level of Fe, Zn  
 187 and Cu in low dose of radiation exposure group when these data compared to the normal untreated control level.

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**Table 3:** Effects of cisplatin administration and/ or radiation exposure on levels of Fe, Zn and Cu in liver tissue 24 hours post radiation exposure (n=6)

Treatment X̄ ± SD	Control group	Cis. Group	IRR group	Cis. & IRR group
Fe (μg/g fresh tissue) X̄ ± SD	205.6±35	183.1±15	186.8±17.8	190.2±21.7
Zn (μg/g fresh tissue) X̄ ± SD	119.7±16.3	112.9±13.5	98.86±13.2	102.3±19.9
Cu (μg/g fresh tissue) X̄ ± SD	3.450 ± 0.49	3.308 ± 0.27	3.322 ± 0.3	3.087 ± 0.28

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Data are presented as mean (± SD). \*Significantly different from the control group (P<0.05). (#)Significantly different from the Cis. group (P<0.05). IRR: Low radiation dose of 0.5 Gy gamma rays, Cis: cisplatin administration.

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**3.3. The synergistic effect of low dose of radiation exposure with cisplatin administration in both liver and testis:**

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**3.3.1. Low dose of radiation (0.5 Gy) induced a significant amelioration in liver function:**

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Fig. (2) showed a full restoration in serum albumin concentration, GPT and BChE activity in the group of animals exposed to low dose 0.5 Gy gamma radiation after cisplatin treatment as regarded to the normal control value. In addition, a significant increase in both serum albumin and BChE levels recorded 13% and 20% (P<0.05) as compared to the cisplatin treated group respectively. In addition; animals exposed to low dose of radiation after cisplatin (10 mg/kg b.wt. I.P) treatment showed a non significant increase as compared to the serum GPT activity of cisplatin treated animals.

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According to data in table (2) the effect of low dose of gamma rays (0.5 Gy) showed a full amelioration as in case of both liver GSH and P450 concentration and their levels become more or less as the normal untreated control level. Comparing the liver GSH and P450 concentration in the group of animal treated with cisplatin before exposing to gamma rays together with cisplatin treated animals group showed a significant increase (11.8% and 24.5%, P<0.05) as in table (2).

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Table (4) showed the effect of cisplatin administration before exposing the animals with low dose of radiation on all tested trace elements concentration that indicated a non significant change in Fe, Zn and Cu in liver tissue as compared with either control untreated or cisplatin treated groups.

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**Table 4:** Effects of cisplatin administration and/ or radiation exposure on levels of Fe, Zn and Cu in testis tissue 24 hours post radiation exposure (n=6)

Treatment X̄ ± SD	Control group	Cis. Group	IRR group	Cis. & IRR group
Fe (μg/g fresh tissue) X̄ ± SD	97.91±16.9	69.61±4.6*	87.09±6.4	96.38±21.2(#)
Zn (μg/g fresh tissue) X̄ ± SD	128.8± 23.6	91.57±6.1*	106.6±13.1	116.7±21.3
Cu (μg/g fresh tissue) X̄ ± SD	1.885±0.39	1.462±0.12*	1.603±0.26	1.603±0.17

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Data are presented as mean (± SD). \*Significantly different from the control group (P<0.05). (#)Significantly different from the Cis. group (P<0.05). IRR: Low radiation dose of 0.5 Gy gamma rays, Cis.: cisplatin administration.

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**3.3.2. Low dose of radiation (0.5 Gy) and its effect on the testis tissue after cisplatin treatment:**

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Exposing cisplatin treated animals to low dose of gamma radiation (0.5 Gy) caused a non- significant decrease in serum FSH activity as compared to the normal control level that means a significant decrease as compared to the cisplatin treated animals only, which calculated 24% Fig. (2). In addition, the effect of whole body gamma radiation (0.5 Gy) on the animals treated with cisplatin only caused a significant increase in testis GSH content comparing to the cisplatin treated

218 group. This increase was calculated as 42% (Table 2) haunt with a non significant change when compared to the normal  
219 untreated group, while the concentration of P450 showed no significant change comparing to normal control levels  
220 accompanied by a significant increase (68%,  $P<0.05$ ) with regards to the cisplatin treated group.

221 In case of trace metals concentration, testis Fe, Zn as well as Cu concentration was estimated in the group of animals  
222 treated with cisplatin before exposing to the low dose of gamma radiation (0.5 Gy). Table (3) showed a significant change  
223 on the level of Fe, Zn and Cu in cisplatin treatment group when these data compared with either normal untreated control  
224 or cisplatin treated one.

#### 225 **4. Discussion**

226 Cisplatin is the most widely used antitumor drug especially in the treatment for testicular cancer, but its usage is limited  
227 by toxic effects on the reproductive system [32]. Testicular damage induced by cisplatin treatment was characterized by  
228 significant decreases in plasma testosterone level [33] that may be due to the increase in FSH level as in the current  
229 work, since according to Sasson and his colleagues the gonadal failure, which caused by cisplatin treatment, leads to  
230 high levels of FSH and LH [34].

231 Once cisplatin has been administered in the body, it rapidly diffuses into tissues and is highly bound to plasma albumin  
232 and other protein [35]. This may explain why cisplatin caused a decrease in serum albumin in the present work. In  
233 addition, the decrease in both serum albumin concentration as well as butyryle cholinesterase with the increase in serum  
234 GPT activities are further evidence that anti-tumoral treatment causes liver damage [36]. According to the observation of  
235 Cepeda and his colleagues, there is a direct connection between the cellular concentrations of copper and platinum  
236 recommended an active transport for cisplatin through copper transporter, which reduce the uptake of each other that  
237 indicated as a significant decrease in testicular copper levels together with Zn and iron that may be due to cisplatin  
238 treatment induced oxidative stress in testis [2]. Consequently, increase in lipid peroxidation (LPO) and depletion of  
239 enzymes such as superoxide dismutase following cisplatin treatment in testis and liver clearly demonstrated failure of  
240 their antioxidant defence system [33]. It may be noted that previous investigators reported the reduction of GSH levels  
241 leads to elevation of LPO [37 & 38]. Therefore, depletion of GSH as in the present results might be inducing oxidative  
242 stress by increasing the free radical generation leading to cell death in both tested organs.

243 Cytochrome p450 is the catalyst enzymes that are essential for the metabolism of many medications. Suppression of  
244 P450 may be due to the consequence of inflammatory processes and can result in increased clinical toxicity of drugs with  
245 a low therapeutic index [39]. Down regulation of testicular cytochrome P450 reported by Waxman and Change and  
246 Masubuchi and his colleagues that was indicated in the present results [40 & 41]. However, exposing to low radiation  
247 dose attenuated the level of P450 after cisplatin treatments. In addition, evidence reported by Oetari and his colleagues  
248 indicates inactivation of glutathione- S- transferase accompanied by glutathione depletion and inhibition of cytochrome  
249 p450 in liver [42]. Interestingly, the cytochrome p450 and GSH content in both liver and testis showed a significant  
250 inhibition than the control group in our current study. Moreover, the active site of cytochrome p450 contain iron heme  
251 center [43], so as one could explain the decrease in both cytochrome p450 and iron in testis as in present work.

252 Cisplatin augments the antitumor effect of a cytotoxic T-lymphocyte-mediated immunotherapy strategy, resulting in a  
253 higher cure rate. This effect is associated with the enhanced ability of cytotoxic T lymphocytes to lyse tumor cells [44]. It  
254 seems that functional activity of lymphocytes decreased at the stage of well-developed tumor, which promoted inhibition  
255 of the lymphocyte defense properties. Cisplatin did not modify the structure and functions of lymphocytes and presumably  
256 improved their energy status [45]. So to overcome the above problems and as the evidence of Murphy and Morton [22],  
257 exposing to the low radiation dose can be used as a synergistic agent with cisplatin for cancer treatment since low  
258 radiation dose administration to entire body increased the action of the protective process in living organism including the  
259 overproduction lymphocytes that significantly prevented or impaired tumor growth. In addition, an interesting observation  
260 remarked that sign of glutathione (GSH) protects against cisplatin cytotoxicity [46]. Of note is that the GSH administration  
261 achieved a superior response rate despite having a larger average tumor burden during chemotherapy [47]. According to  
262 current study animals exposed to low radiation dose alone (0.5 Gy) showed a significant increase in GSH content in both  
263 liver and testis tissue, which may explain the antioxidant effects of low radiation dose against cisplatin induced oxidative  
264 stress. These antioxidants did not interfere with chemotherapeutic effects - and mitigated cancer treatment toxicity [48].

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## 5. Conclusion:

In conclusion, the obtainable results proved that low radiation dose have the existence as an antioxidant and antitumor agents which may be useful to use as a synergistic agents with the chemotherapeutic drug or even in case of tumors chemo-radiotherapy. Further studies should be needed to address this issue in different biochemical views.

## 6. REFERENCES

1. Giaccone, G. Clinical perspectives on platinum resistance. *Drugs*. 2000; 59 (Suppl 4): 9–17.
2. Cepeda, V, Miguel, AF, Castilla, J, Alonso, C, Quevedo, C, and Pérez, JM. Biochemical Mechanisms of Cisplatin Cytotoxicity, Anti-Cancer Agents in Medicinal Chemistry. 2007; 7: 3-18
3. Chirino, YI, Pedraza- Chaverri, J. Role of oxidative and nitrosative stress in cisplatin-induced nephrotoxicity. *Exp Toxicol Pathol*. 2009; 61(3):223-42. doi: 10.1016/j.etp.2008.09.003. Epub 2008 Nov 4.
4. Hassan, I, Chibber, S., Khan, AA, Naseem, I. Riboflavin Ameliorates Cisplatin Induced Toxicities under Photoillumination. *PLoS ONE*. 2012; 7(5): e36273.
5. González, VM, Fuertes, MA, Alonso, C, Pérez, JM. *Mol. Pharmacol*. 2001; 59: 657.
6. Sadowitz, PD, Hubbard, BA, Dabrowiak, JC, Goodisman, J, Tacka, KA, Aktas, MK, Cunningham, MJ, Dubowy, RL, Souid, AK. Kinetics of cisplatin binding to cellular DNA and modulations by thiolblocking agents and thiol drugs. *Drug Metab. Dispos*. 2002; 30: 183–190.
7. Coia, L, Won, M, Lanciano, R, Marcial, VA, Martz, M, Hanks, G. The patterns of care outcome study for cancer of the uterine cancer: Results of the second national practice survey. *Cancer*. 1990; 66: 2451-2456.
8. Morri, y, Shara, H, Shima, H, Shimada, K, Arima, M, Ikoma, F. Combined cisplatin and radiation therapy in patients with invasive bladder cancer. *International Urology and Nephrology*. 1990; 22(4): 337- 343.
9. Wodinsky, I, Swiniarki, J, Kensler, CJ, Venditti, JM. Combination radiotherapy and chemotherapy for P388 lymphocytic leukemia in vivo *Cancer Chemother. Rep*. 1974; 4: 73.
10. Morris, CR, Blackwell, LH, Loveless, VS. Antileukemic of combination of radiation and malanats (1,2 diamininocyclohexane) platinum II (NSC. 224964) *J. Med*. 1977; 8: 254 (1977)
11. Omura, GA. (1996) Chemotherapy for stage IVB or recurrent cancer of the uterine cervix. *Journal of the National Cancer Institute*. 1996; 21:123–6.
12. Wallner, KE, Li, GC. Effect of cisplatin resistance on cellular radiation response. *Int J Radiat Onco Biol Phys*. 1987; 13:587–91.
13. Perez, CA, Grigsby, PW, Castro, Vita, H, Lockett, MA. (1995) Carcinoma of the uterine cervix. I. Impact of prolongation of treatment time and timing of brachytherapy on outcome of radiation therapy. *Int J Radiat Onco Biol Phys*. 32:1275–88.
14. Green, JA, Kirwan, JM, Tierney, JF, *et al*. Survival and recurrence after concomitant chemotherapy and radiotherapy for cancer of the uterine cervix: a systematic review and meta-analysis. *Lancet*. 2001; 358: 781-6.
15. Ishizuka, M., Fujimoto, Y, Itoh, Y, Kitagawa, K, Sano, M, Miyagawa, Y, Ando, A, Hiramatsu, M, Hirasawa, N, Ishihara, S, Nakashima, T, Yamada, K. Relationship between hematotoxicity and serum albumin level in the treatment of head and neck cancers with concurrent chemo-radiotherapy using cisplatin. *Jpn. J. Oncol*. 2011; 41(8): 973- 979.
16. Nogami, M, Huang, JT, James, SJ. Mice chronically exposed to low dose ionizing radiation possess splenocytes with increased levels of HSP70 mRNA, HSC72 and HSP70 and with an increased capacity to proliferate. *Int J Radiat Biol*. 1993; 63:775–783.



- 308 17. Kojima, S, Nakayama, K, Ishida, H. Low Dose  $\gamma$ -Rays Activate Immune Functions via Induction of Glutathione  
309 and Delay Tumor Growth. *J Radiat Res.* 2004; 45 (1): 33-39. doi: 10.1269/jrr.45.33
- 310 18. Feinendegen, LF. Evidence for beneficial low level radiation effects and radiation hormesis. *Brit J Radiol.* 2005;  
311 78: 3-7. doi:10.1259/bjr/63353075
- 312 19. Gharib, OA, Abd Ellatif, UA, Abdellah, NM, Mohammad, MA. Radio-protective response on the environmental  
313 pollutant induced oxidative stress. *Adv Bioscie Biotech.* 2012; 3: 989-996.
- 314 20. Kopp, AH, Eifel, PJ. Chemo-radiotherapy for cervical in 2010. *Curr Oncol. Rep.* 2011; 13:77-85.
- 315 21. Turner, J.E. (1995). *Atoms radiation and radiation protection (2<sup>nd</sup> Edition)*, John Willy & Sons, inc., New York.
- 316 22. Murphy, B, Morton, JJ. *J. Exper.* 1915; *Med.* 22: 204.
- 317 23. Barcellos-Hoff, MH, Park, C, Wright, EG. Radiation and the microenvironment - tumorigenesis and therapy. *Nat*  
318 *Rev Cancer.* 2005; 5: 867–875.
- 319 24. Máthé, A, Komka, K, Forczig, M, Szabó, D, Anderlik, P, Rozgonyi, F. The effect of different doses of cisplatin  
320 on the pharmacokinetic parameters of cefepime in mice. *Lab Anim.* 2006; 40:296–300
- 321 25. Munshaw, S, Hwang, HS, Torbenson, M, Quinn, J, Hansen, KD, Astemborski, J, Mehta, SH, Ray, SC, Thomas,  
322 DL, Balagopal, A. Laser captured hepatocytes show association of butyrylcholinesterase gene loss and fibrosis  
323 progression in hepatitis C-infected drug users. *Hepatology.* 2012; 56(2):544-54. doi: 10.1002/hep.25655.
- 324 26. Doumas, BT, Watson, WA, Biggs, HG. *Clin. Chim.Acta.* 1971; 31: 87-95
- 325 27. Beutler, E. *Red Cell Metabolism: A Manual of Biochemical Methods*, Grune and Stratton, New York. 1971.
- 326 28. Nerurkar, PV, Park, SS, Thomas, PE, Nims, RW, Lubet, RA. Methoxyresorufin and benzyloxyresorufin:  
327 substrates preferentially metabolized by cytochromes P4501A2 and 2B, respectively, in the rat and mouse.  
328 *Biochem Pharmacol.* 1993; 46: 933-943.
- 329 29. IAEA. *Elemental analysis of biological materials. Current problem and techniques with special reference to*  
330 *trace elements.* International Atomic Energy Agency, IAEA, Vienna. Technical Reports Series. 1980; 379.
- 331 30. Kingstone, HM, Jassie, L. *Introduction to microwave sample preparation. Theory and practice.* American  
332 *Chemical Society Professional Reference Book*, Washington DC. 1998; 263.
- 333 31. Gregus, Z, Klaassen, CD. Disposition of metals in rats: A comparative study of fecal urinary and biliary  
334 excretion and tissue distribution of eighteen metals. *Toxicology and Applied Pharmacology.* 1986; 85: 24-38.  
335 doi:10.1016/0041-008X(86)90384-4
- 336 32. Ciftci, O, Beytur, A, Cakir, O, Gurbuz, N, Vardi, N. Comparison of Reproductive Toxicity Caused by Cisplatin  
337 and Novel Platinum-N-Heterocyclic Carbene Complex in Male Rats. *Basic & Clinical Pharmacology &*  
338 *Toxicology.* 2011; 109: 328–333.
- 339 33. Fahmy, HA, Abd El-Azime, ASh, Gharib, OA. Possible Ameliorative Role of Low Dose of Radiation Against  
340 Cisplatin Induced Oxidative Stress and Tissue Damage in Male Rats. *Euro J Biol Medi Sci Resh.* 2013; 1(4):  
341 10-18.  
342
- 343 34. Sasson, R, Dantes, A, Tajima, K, Amsterdam, A. Novel genes modulated by FSH in normal and immortalized  
344 FSH-responsive cells: new insights into the mechanism of FSH action. *FASEB J.* 2003; 17(10):1256-66.
- 345 35. Judson, I, Kelland, LR. *New developments and approaches in the platinum arena.* 2000; 59: 29.
- 346

- 347 36. Fasihi, M, Ghodrati-zadeh, S, Ghodrati-zadeh, S. Protective Effect of Captopril on Cisplatin Induced  
348 Hepatotoxicity in Rat. *American-Eurasian. J Toxicol Sci.* 2012; 4(3): 131-134.
- 349 37. El-Maraghy, SA, Gad, MZ, Fahim, AT, Hamdy, M A. Effect of cadmium and aluminum intake on the antioxidant  
350 status and lipid peroxidation in rat tissues. 2001; *J Biochem Mol Toxicol.* 2001; 15(4):207-14.
- 351 38. Gharib, OA, Ibrahim, NK. Oxidative damage in testes induced by 950 MHz simulating cellular phone. *Isotope  
352 and Radiat Res.* 2010; 42(4) , 9 4 1 - 9 5 3.
- 353 39. Morgan, ET. Regulation of cytochromes P450 during inflammation and infection. *Drug Metab Rev.* 1997;  
354 29:1129–1188.
- 355 40. Waxman, DJ, Chang, TKH. Hormonal regulation of liver cytochrome P450 enzymes. In *Cytochrome P450:  
356 Structure, Mechanism, and Biochemistry* (2nd edition). Ortiz de Montellano P. R., editor. Plenum Press, New  
357 York. 1995; 391–418.
- 358 41. Masubuchi, Y, Enoki, K, Horie, T. Down-regulation of hepatic cytochrome P450 enzymes in rats with  
359 trinitrobenzene sulfonic acid-induced colitis *Drug Metab Dispos.* 2008; 36(3):597-603.
- 360 42. Oetari, S, Sudibyo, M, Commandeur, JN, Samhoedi, R, Vermeulen, NP. Effects of curcumin on cytochrome  
361 P450 and glutathione S-transferase activities in rat liver. *Biochem Pharmacol.* 1996; 12,51(1):39-45.
- 362 43. Guengerich, FP. "Cytochrome p450 and chemical toxicology". *Chem. Res. Toxicol.* 2008; 21 (1): 70–83.
- 363 44. Merritt, RE, Mahtabifard, A, Yamada, RE, Crystal, RG, Korst, RJ. (2003) Cisplatin augments cytotoxic T-  
364 lymphocyte-mediated antitumor immunity in poorly immunogenic murine lung cancer. *J Thorac Cardiovasc  
365 Surg.* 2003; 126(5):1609-17.
- 366 45. Zamay, TN, Kolovskaya, OS, Zamay, GS, Borodina, NA. Effects of Cisplatin on Lymphocyte Structure and  
367 Functions in Mice with Ehrlich Ascitic Carcinoma. *Bull Experi Biol & Medi.* 2011; 151 (1): 62-65.
- 368 46. Lu, y, Kawashima, A, Horri, I, Zhong, L. Effect of BSO and L- cysteine on drug induced cytotoxicity in primary  
369 cell cultures: drug-, cell type- and species- specific difference. *Drug Chem. Toxicol.* 2004; 27: 269- 280.
- 370 47. Schmidinger, M, Budinsky, AC, Wenzel, C, Piribauer, M, Brix, R, Kautzky, M, Oder, W, Locker, GJ, Zielinski,  
371 CC, Steger, GG. Glutathione in the prevention of cisplatin induced toxicities. A prospectively randomized pilot  
372 trial in patients with head and neck cancer and non small lung cancer. *Wien Klin Wochenschr.* 2000; 112(14):  
373 617-23.
- 374 48. Block, KI, Koch, AC, Mead, MN, Tothy, PK, Newman, RA, Gyllenhaal, C. Impact of antioxidant supplementation  
375 on chemotherapeutic efficacy: A systematic review of the evidence from randomized controlled trials, *Cancer  
376 Treatment Reviews.* 2007. doi:10.1016/j.ctrv.2007.01.005.