

Antibacterial activity of phenolic compounds derived from *Ginkgo biloba* sarcotestas against food-borne pathogens

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ABSTRACT

Aims: We investigated the antibacterial activity of three groups of phenolic compounds obtained from the chloroform (CHCl₃) extract of the fleshy seed coat (sarcotestas) of *Ginkgo biloba* on several bacterial pathogens.

Study design: An experimental study.

Methodology: Inhibition of microbial growth was measured by an agar diffusion method and susceptibility tests were performed by the broth microdilution method. Bactericidal effect of *Ginkgo biloba* compound 5-7 against *Salmonella enteric* serovar Typhimurium was assessed by time-kill assay.

Results: *Ginkgo biloba* compounds 5-7 and 8-10 showed high antimicrobial activity against Gram-positive and Gram-negative bacteria, including several food-borne pathogens. In particular, compounds 5-7 and 8-10, containing phenolic acids and bilobols, respectively, were highly effective against *Salmonella enteric* serovar Typhimurium, *Listeria monocytogenes*, *Listeria innocua*, *Streptococcus pyogenes*, *Escherichia coli*, and *Shigella dysenteriae*. On the opposite, compounds 1-4, containing cardanols, showed little antibacterial activity. Compounds 5-7 exerted a bactericidal and bacteriolytic effect on *Salmonella enteric* serovar Typhimurium with a minimal inhibitory concentration (MIC) and a minimal bactericidal concentration (MBC) of 8.3 µg ml⁻¹.

Conclusion: The results of this study indicate that phenolic compounds derived from *Ginkgo biloba* sarcotestas, because of their strong inhibitory characteristics towards food pathogens, can be considered ideal candidates for possible application in food microbiology due to their natural origins.

Keywords: *Ginkgo biloba* compounds; antibacterial activity; *Salmonella enterica* serovar Typhimurium; natural antimicrobials; food microbiology; food-borne pathogens

1. INTRODUCTION

Ginkgo biloba is a tree native to China that reaches 30-40 meters height. It belongs to the Ginkgoaceae family and its beneficial effects on human health are largely known. In fact, Ginkgo has been used therapeutically for centuries in Traditional Chinese Medicine [1]. Currently, it represents one of the top-selling herbs in health food stores in the United States [2]. Among several compounds, *Ginkgo biloba* contains flavonoids (ginkgo-flavone glycosides) and terpenoids (ginkgolides and bilobalides). Its traditional preparations are used to ameliorate peripheral vascular disease, such as intermittent claudication and cerebral deficiency. Moreover, the ginkgolides inhibit platelet aggregation and the initial symptoms of arteriosclerosis [3-4].

In Western medicine, dry extracts of *Ginkgo biloba* leaves, known as Egb761 [3] are reported to improve mood and cognitive performance, to protect memory deficits and central nervous system disorders, to alleviate symptoms of mild/moderate Alzheimer-type dementia, to possess antidepressant and antioxidant properties [5-8]. Moreover, increasing evidence suggests a beneficial use of Egb in treating cardiovascular diseases [9-10].

Despite the numerous works on the healthy properties and the antimicrobial activity of *Ginkgo biloba* leaves constituents, little is known about the antimicrobial activity of seed extracts [11-18].

The aim of this study is to evaluate the *in vitro* antibacterial activity of three mixtures of compounds, named 1-4, 5-7, and 8-10, derived from the fleshy seed coat (sarcotestas) of *Ginkgo biloba*. These compounds, previously characterized as cardanols, phenolic acids, and bilobols, respectively [19] (Fig. 1), have been screened for antimicrobial activity against 11 bacterial strains, including Gram-positive, Gram-negative, pathogenic and safe strains. For mixtures 5-7 and 8-10, the minimal inhibitory and bactericidal concentrations were determined. Finally, we analyzed the kinetics of *Salmonella enterica* serovar Typhimurium growth inhibition by compounds 5-7.

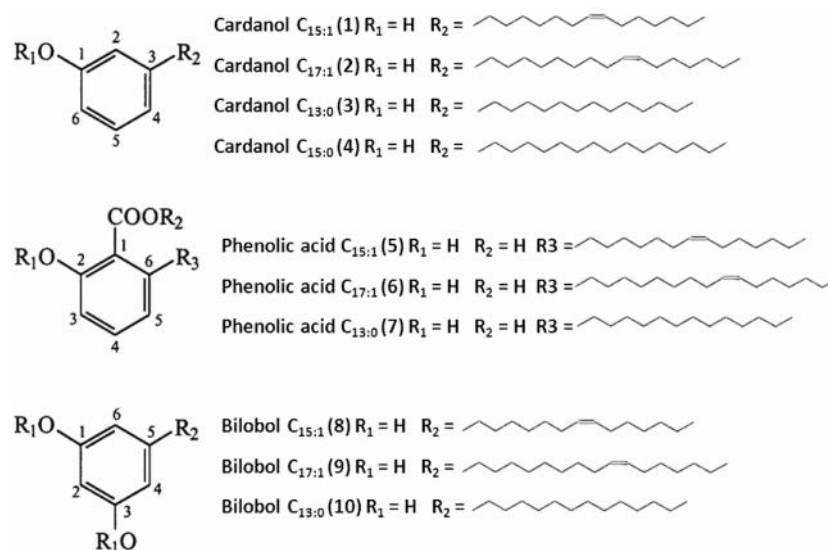


Fig. 1. Structures of *Ginkgo biloba* compounds. Adapted from Lee et al. (1998)

2. MATERIAL AND METHODS

2.1 Bacterial strains and media

Bacterial strains used in the present study were from the author's Institute collection or from the American Type Culture Collection (ATCC, Rockville, MD, U.S.A.). *Escherichia coli*, *Enterobacter aerogenes*, *Pseudomonas aeruginosa*, *Salmonella enterica* serovar Typhimurium, *Shigella dysenteriae* ATCC 11835, *Staphylococcus aureus*, *Streptococcus pyogenes*, *Vibrio mediterranei*, and *Vibrio vulnificus* were grown in Nutrient Broth (NB, Becton Dickinson, Cockeysville, USA). *Listeria innocua* and *Listeria monocytogenes* were propagated in Tryptone Soya Broth (TSB, Oxoid Limited, Basingstoke, UK). The plating medium for the activity assay was Tryptone Soya Agar (TSA) containing 3% TSB and 1.5% agar (Oxoid). For *Vibrio mediterranei* and *Vibrio vulnificus*, each medium was supplemented with 3% NaCl according to the halophilic features of the strains.

For the Minimal Inhibitory Concentration (MIC) and Minimal Bactericidal Concentration (MBC) determination, Mueller-Hinton II broth (BBL Microbiology Systems, Cockeysville, Md, USA) and TSA were used, respectively.

2.2 *Ginkgo biloba* compounds

The fresh sarcotestas of *Ginkgo biloba* L. (Ginkgoaceae) were collected from the ginkgo trees in Korea and were identified by Dr. Dae Suk Han, emeritus

professor, College of Pharmacy, Seoul National University, as previously reported [19]. The fresh sarcotestas were extracted in chloroform (CHCl_3) and purified by chromatographic methods exactly as described [19]. The structures of compounds 1-10 shown in Fig. 1 were identified by IR, ^1H NMR, ^{13}C NMR, and MS and confirmed by comparison with those of literature data [19 and references therein]. The 10 phenolic compounds were pooled in three groups: cardanols (compounds 1-4), phenolic acids (compounds 5-7), and bilobols (compounds 8-10) (Fig. 1). For the present study, all these compounds were dissolved in dimethyl sulfoxide (DMSO, Merck, Darmstadt, Germany), and kept in aliquots at -20°C , before use in antimicrobial tests.

2.3 Antibacterial assay

We used an agar diffusion method [20]. Briefly, suspensions of bacterial strains grown overnight in appropriate media were adjusted to 10^8CFU ml^{-1} , inoculated in 8 ml of semi-solid Tryptone Soya Agar (0.7% agar) and poured over TSA plates (agar 1.5%). After cooling and drying, 10 μl of each *Ginkgo biloba* mixture, e.g., 1-4, 5-7, 8-10, corresponding to 33.3 μg , were applied using a micropipette and allowed to diffuse. The plates were then inverted and incubated for 18-24 h at the optimal temperature of the test organism. The presence of a clear zone around the spot indicated growth inhibition and the diameter of the zone of inhibition was measured. As a positive control, a mixture of the following antibiotics was used: Pen/Strep (penicillin 5000 IU/ml; streptomycin 5000 $\mu\text{g ml}^{-1}$ (Life Technologies, Milan, Italy). DMSO and Nutrient broth were employed as negative controls. In a different experiment, serial dilutions of *Ginkgo biloba* mixtures 1-4, 5-7, 8-10 were spotted as above to verify the dose-dependence of the inhibition in solid media. The highest concentration applied was 3330 $\mu\text{g ml}^{-1}$.

2.4 Susceptibility testing

The in vitro activities of the *Ginkgo biloba* compounds against the most sensitive bacteria (*Salmonella enterica* serovar Typhimurium, *Listeria monocytogenes*, *Listeria innocua*, *Streptococcus pyogenes*, *Escherichia coli*, *Shigella dysenteriae* ATCC 11835) were determined by the EUCAST (European Committee on Antimicrobial Susceptibility Testing) broth microdilution method [21]. Briefly, microtiter plates containing serial 10-fold dilutions of each *Ginkgo biloba* compound, with concentration range varying from 3.33 to 3330 $\mu\text{g ml}^{-1}$, were inoculated in the presence of single strains to yield the appropriate density (10^5CFU/ml). The plates were incubated aerobically for 24 h at 37°C . For a more accurate measurement of the MIC, the lowest concentrations of the compounds with activity underwent further two-fold dilutions. The wells without any visible growth of microorganisms

were sub-cultured in order to determine the **MBC** that was defined as the lowest concentration of the test compound that killed all the bacteria.

2.5 Bactericidal effect of *Ginkgo biloba* compound 5-7 against *Salmonella enterica* serovar Typhimurium

Nutrient Broth (NB) containing either 40 or 80 μ l (corresponding to 133.2 and 266.4 μ g, respectively) of *Ginkgo biloba* compounds 5-7 was inoculated in an overnight culture of *Salmonella enterica* serovar Typhimurium, grown at 37 °C in NB adjusted to O.D₅₉₀ = 0.1. Growth of *Salmonella enterica* serovar Typhimurium was monitored by measuring the optical density of broth cultures at 590 nm at various times, up to 24 h. Viable bacterial cells were enumerated by standard plate count method. Medium plus bacterial culture, and medium plus bacterial culture containing 80 μ l DMSO were tested as controls.

3. RESULTS AND DISCUSSION

The spectrum of activity of *Ginkgo biloba* compounds was very broad. High levels of inhibitory activity were detected against *Salmonella enterica* serovar Typhimurium, *Listeria monocytogenes*, *Listeria innocua*, *Streptococcus pyogenes*, *Escherichia coli*, *Shigella dysenteriae*, while a minor activity was evidenced against *Enterobacter aerogenes*, *Vibrio vulnificus*, *Staphylococcus aureus*. Finally, *Pseudomonas aeruginosa* and *Vibrio mediterranei* were not affected (Table 1). Compounds 5-7 and 8-10 showed the highest efficacy, while compound 1-4 had little antimicrobial activity. In particular, *Salmonella enterica* serovar Typhimurium and *Listeria* spp. showed the highest sensitivity to *Ginkgo biloba* compounds.

Table 1. Antibacterial activity of *Ginkgo biloba* **crude extracts** expressed as diameter of inhibition zone (mm)

Organisms	Diameter of inhibition zone (mm)*		
	Compounds 1-4**	Compounds 5-7**	Compounds 8-10**
<i>Escherichia coli</i>	4.1 \pm 0.3	21.4 \pm 0.7	17.0 \pm 0.6
<i>Enterobacter aerogenes</i>	4.3 \pm 0.2	10.8 \pm 0.4	8.8 \pm 0.6
<i>Listeria innocua</i>	5.5 \pm 0.4	22.2 \pm 0.9	20.2 \pm 0.8
<i>Listeria monocytogenes</i>	4.7 \pm 0.2	24.0 \pm 0.8	21.4 \pm 0.8
<i>Pseudomonas aeruginosa</i>	1.4 \pm 0.3	2.1 \pm 0.3	2.3 \pm 0.6
<i>Salmonella enterica</i> serovar Typhimurium	6.0 \pm 0.4	24.5 \pm 0.8	24.4 \pm 0.7
<i>Shigella dysenteriae</i>	3.7 \pm 0.4	16.0 \pm 0.8	15.5 \pm 0.6

<i>Staphylococcus aureus</i>	4.6 ± 0.2	8.7 ± 0.7	8.3 ± 0.7
<i>Streptococcus pyogenes</i>	4.0 ± 0.0	21.5 ± 0.6	17.5 ± 0.3
<i>Vibrio mediterranei</i>	1.2 ± 0.2	2.0 ± 0.5	1.9 ± 0.4
<i>Vibrio vulnificus</i>	3.5 ± 0.1	10.5 ± 0.4	8.8 ± 0.7

*The values are expressed as mean ± standard deviation (SD) of three independent experiments.

**Concentration applied: 3330 µg ml⁻¹.

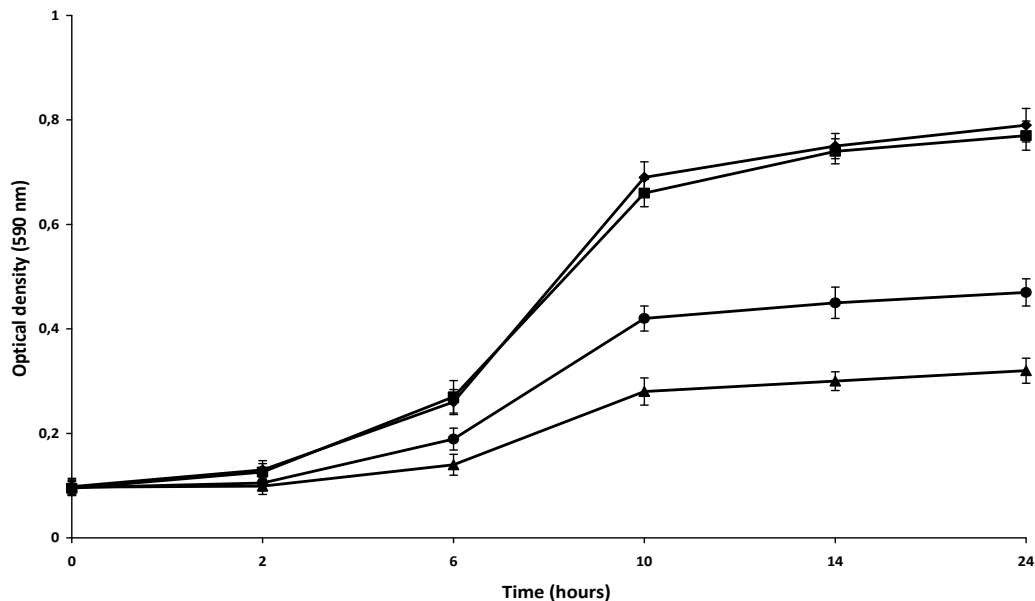
Analyzing the diameter of the inhibition zone (Table 1) and the results of susceptibility test (Table 2), mixture 5-7 was slightly more active than 8-10 against most of the strains tested. In fact, MIC and MBC values were in the range of 8.3-33.3 µg ml⁻¹ and 8.3-333.0 µg ml⁻¹, respectively, for mixture 5-7, while the same parameters ranged between 8.3-33.3 µg ml⁻¹ and 16.6-333.0 µg ml⁻¹ for compounds 8-10. More in details, compounds 5-7 was strongly inhibitory against *Salmonella enterica* serovar Typhimurium (MIC and MBC: 8.3 µg ml⁻¹), *Listeria monocytogenes* (MIC and MBC: 8.3 µg ml⁻¹ and 16.6 µg ml⁻¹, respectively), *Escherichia coli*, and *Streptococcus pyogenes* (Table 2).

Table 2. MIC and MBC values for *Ginkgo biloba* compounds 5-7 and 8-10 expressed as µg ml⁻¹

Organisms	Compound 5-7		Compound 8-10	
	MIC	MBC	MIC	MBC
<i>Escherichia coli</i>	16.6	33.3	33.3	333.0
<i>Listeria innocua</i>	16.6	16.6	16.6	33.3
<i>Listeria monocytogenes</i>	8.3	16.6	16.6	33.3
<i>Salmonella enterica</i> serovar Typhimurium	8.3	8.3	8.3	16.6
<i>Shigella dysenteriae</i>	33.3	333.0	33.3	333.0
<i>Streptococcus pyogenes</i>	16.6	33.3	33.3	333.0

The analysis of growth inhibition kinetic triggered by compounds 5-7 against *Salmonella enterica* serovar Typhimurium indicated a bactericidal effect, as resulted by cell lysis (data not shown) and loss of turbidity in the growth medium (Fig. 2). Growth inhibition was detected starting from 2 h with a gradual increase over time. Both growth rate and cell density in the treated cultures decreased when the amount (volume) of compounds 5-7 added to the culture were doubled (Fig. 2). DMSO, used as negative control, did not inhibit *Salmonella enterica* serovar Typhimurium growth.

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Fig. 2. Inhibitory effect of *Ginkgo biloba* mixture 5-7 on *Salmonella enterica* serovar Typhimurium growth. *Ginkgo biloba* mixture 5-7, 40 µl (●) or 80 µl (▲) corresponding to 133.2 and 266.4 µg, respectively, was added to an early log-phase culture of *Salmonella enterica* serovar Typhimurium. Optical density (OD) at 590 nm was measured. A decrease in the optical density of the bacterial culture indicates an increase of inhibition. Controls were medium plus bacterial culture without compound 5-7 (♦), and medium plus bacterial culture containing 80 µl DMSO (■). Values represent mean of three separate experiments ± standard deviation (SD).

Fig. 3 shows the dose-effect of the three compounds against *Salmonella enterica* serovar Typhimurium in solid media. The linear correlation between volume of inoculum and size of inhibition zone is evident. The presence of a clear zone around the spotted area confirmed the bactericidal mode of action of compounds 5-7.

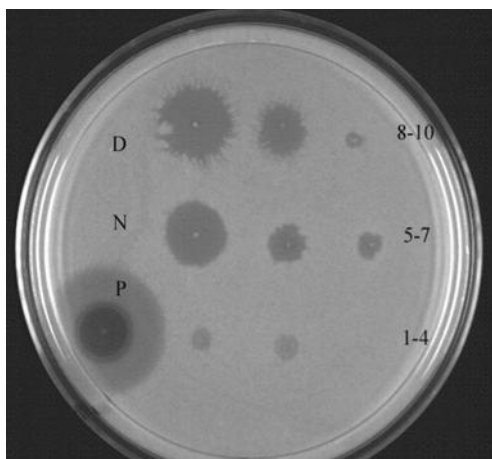


Fig.3. Antibacterial activity of *Ginkgo biloba* compounds. *Salmonella enterica* serovar Typhimurium (10^8 CFU ml^{-1}) was inoculated into 8 ml of molten TSA, and poured over TSA plates. Serial dilutions of compounds 8-10, 5-7, and 1-4 were spotted on the plate. Controls were DMSO (D), Nutrient Broth (N), and penicillin/streptomycin (P).

Salmonellosis continues to be a major public health problem worldwide and *Salmonella enteric* serovar Typhimurium is one of the most prevalent serovars among *Salmonella* spp. causing gastroenteritis [22]. Rates of multidrug-resistance in *Salmonella* spp. increased considerably in recent years, primarily in response to antimicrobial usage in humans and food animals. An example is the global spread of multidrug-resistant *Salmonella enterica* serovar Typhimurium phage type DT104, resistant to ampicillin, chloramphenicol, streptomycin, sulphonamides and tetracyclines [23]. *Listeria monocytogenesis* an important pathogen which has been isolated from various foodstuffs such as meat, poultry, eggs, seafood, and represents a major concern to food manufacturers [24]. In recent years, there has been an increasing interest in the development of effective natural antimicrobials as food preservatives. *Ginkgo biloba* extracts used in this study showed antibacterial activity against a wide panel of bacteria, including most of the tested pathogens.

Previous studies regarding the antimicrobial activity of *Ginkgo biloba* extracts showed contrasting results. In fact, Mazzanti *et al.* [12] reported that the antimicrobial activity of three fractions of methanolic extracts of *Ginkgo biloba* leaves was effective towards Gram-positive bacteria only. Lee and Kim [13] found that *Ginkgo biloba* leaf-derived materials inhibited *Clostridium perfringens* and *Escherichia coli*, but did not inhibit the anaerobic intestinal bacteria, such as *Bifidobacterium bifidum*, a common and beneficial lactic acid bacteria, or *Lactobacillus acidophilus*. Sati and Joshi [14] showed that the methanolic extract of *Ginkgo biloba* leaves possessed inhibitory activity against *Escherichia coli*, *Bacillus subtilis* and some plant pathogenic bacterial

strains. Tao *et al.* [15] reported the antibacterial/antifungal activities and synergistic interactions between *Ginkgo biloba* polyphenols and eight compounds separated from *Ginkgo biloba* L. leaves lipids against *Salmonella enterica*, *Staphylococcus aureus* and *Aspergillus niger*.

Moreover, Boonkaew and Camper [16] observed that methanolic extracts from leaf and root tissue of *Ginkgo biloba* showed no inhibitory activity, but extracts from leaf and root derived callus inhibited the growth of *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Staphylococcus* spp., and *Streptococcus pyogenes*. No activity against *Escherichia coli* was detected.

Regarding studies reporting the antimicrobial activity of *Ginkgo biloba* seed extract constituents, Sawano *et al.* [17] isolated a protein from the seeds of *Ginkgo biloba* that inhibited the growth of some fungi, but did not exhibit antibacterial activity against *Escherichia coli*, while Choi *et al.* [18] showed that a chloroform fraction prepared from sarcotestas of *Ginkgo biloba* possessed inhibitory activity against vancomycin-resistant *Enterococcus* spp.

The results of the present study indicate that *Ginkgo biloba* sarcotestas-derived compounds possess a remarkably high inhibitory activity against a wide spectrum of Gram-positive and Gram-negative bacteria. It is worthwhile to note that the tested compounds were highly active against *Salmonella enterica* serovar Typhimurium and *Listeria monocytogenes*. Moreover, *Ginkgo biloba* extracts 5-7 and 8-10 were also inhibitory towards other important food-borne pathogens, such as *Shigella dysenteriae*, *Escherichia coli* and the human pathogens *Staphylococcus aureus* and *Streptococcus pyogenes*.

4. CONCLUSION

Increase in antibiotic resistance, including multiple antibiotic resistances among several groups of bacteria, is a global phenomenon [25-26]. Reports on resistance in *Listeria* spp. have been previously published [27-29]. Therefore, new antimicrobial substances to counteract antibiotic resistance is urgently need. *Ginkgo biloba* mixtures 5-7 and 8-10 may represent ideal, natural candidates in potential applications aimed to preserve microbiological contamination of foods.

Further studies are needed to evaluate the antimicrobial activity of the individual components of the *Ginkgo biloba* compounds used in the present study and to test them for cytotoxicity.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

AUTHORS' CONTRIBUTIONS

This work was carried out in collaboration between all authors. Authors AC and GLR designed the study, wrote the protocol and the draft of the manuscript. Authors AC, KR, and IT managed the analyses of the study and literature searches. Author JK prepared all the extracts of *Ginkgo biloba*. All authors read and approved the final manuscript.

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