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PART 1:

Journal Name:	British Microbiology Research Journal
Manuscript Number:	**2013_BMRJ_3708**
Title of the Manuscript:	**Detection of Mycobacterium tuberculosis Complex in Lung Specimen of Slaughtered Cattle and Goats By a DNA Based multiplex- Polymerase Chain Reaction and Ziehl-Neelsen Methods In Jos, Nigeria.**

General guideline for Peer Review process is available in this link:
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- This form has total 9 parts. Kindly note that you should use all the parts of this review form.

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PART 2: Review Comments

	Reviewer's comment	**Author's comment** *(if agreed with reviewer, correct the manuscript and highlight that part in the manuscript. It is mandatory that authors should write his/her feedback here)*
Compulsory REVISION comments	Abstract: Abbreviations should respect and these should be the same throughout the text. As an example see the corrections I suggest at the bottom of this page in Aims (1) Introduction: There are several spelling mistakes in the name of members of the MTBC. I copy the example as the introduction could begin (2) **However, the most serious problem of this study is the difference in the sequence of the primers JB21 and JB22 previously described (J. G. Rodriguez, G. A. Meja, P. Del Portillo, M. E. Patarroyo, and L. A. Murillo, Microbiology 141:2131–2138, 1995) and the sequence reported in the present manuscript.** **The same for the primers INS1 and INS2 described by Kolk et al in J of Clinical Microbiology, p. 2567-2575 , 1992.** **DNA amplification section in the present paper, page 10, says:** **PCR was performed in a 50µl reaction volume comprised of 26.5µl nuclease-free water (Fermetas®), 5µl dNTPs (10mM), 1µl each of JB21, + JB 22 primers for 500bp (JB21, 5´–CGTCCGCTGATGCAAGTGC-3´ and JB22, 5´-CGTCCGCTGACCTCAAGAAAG-3´) INS1+INS2 primers for 245bp (INS1, 5´–CCTGCGAGCGTAGGCGTCGG-3´ and INS2, 5´- CTCGTCCAGCGCCGCTTCG-3´).** **While primers reported in the literature are:** **The species-specific primers proposed by Rodríguez et al. to identify by PCR *M. bovis* strains were JB21 (5′-TCGTCCGCTGATGCAAGTGC) and JB22 (5′-CGTCCGCTGACCTCAAGAAG)** **The primers INS1 and INS2 reported to amplify 245pb by PCR of IS6110 were INS1 5'-CGTGAGGGCATCGAGGTGGC-3' and INS2 5'-GCGTAGGCGTCGGTGACAAA-3'.**	

Minor REVISION comments		
Optional/General comments	1 Aims : Determine percentage of acid fast bacilli (AFB) present in samples by Ziehl-Neelsen stain (ZN), detect members of *Mycobacterium tuberculosis* complex (MTBC) using a multiplex PCR (m-PCR), compare number of positive samples detected by ZN and m–PCR, determine prevalence rates of **MTBC** between in cattle and goat and the rates of detecting agents of tuberculosis using **ZN and m-PCR** in lung specimens of slaughtered cattle and goats in the study area. 2 Introduction: Members of the *Mycobacterium tuberculosis* complex (MTBC) are slow-growing, aerobic intracellular and acid-fast bacilli that belong to the family **Mycobacteriaceae**. MTBC refers to a genetically closely related group of Mycobacterium. It has a number of member include, *M. bovis*, *M. africanum*, *M. microti*, *M. bovis* Bacillus Calmette-Guerin (BCG), ***M. caprae***, *M. pinnipedii*, and ***M. canettii*** [3]. These	

	bacteria are the causative agent of tuberculosis in humans and animals, [1]. *Mycobacterium bovis* is closely related to the species *Mycobacterium tuberculosis,* etiologic agent of tuberculosis in human. *Mycobacterium bovis,* etiologic agent of tuberculosis in cattle, can also cross the species barrier and cause tuberculosis (Tb) in humans [2] and other animals such as goat, deer, and cat.	

Reviewer Details:

Name:	**Romano, María Isabel**
Department, University & Country	**Instituto de Biotecnología, INTA, Buenos Aires , Argentina**