

CURRENT TAXONOMY OF RHIZOBIA

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Abstract

Classification of rhizobia knows constant change. Recent studies have shown the existence of a great diversity among nitrogen-fixing bacteria isolated from different legumes. Currently, More than 98 species belonging to 13 genera of α - and β - proteobacteria have been described as rhizobia. The genera *Rhizobium*, *Mezorhizobium*, *Ensifer* (formerly *Sinorhizobium*), *Bradyrhizobium*, *Phyllobacterium*, *Microvirga*, *Azorhizobium*, *Ochrobacterium*, *Methylobacterium*, *Devosia*, *Shinella* (Class of α -proteobacteria), *Burkholderia*, *Cupriavidus* (formerly *Ralstonia*) (Class of β -proteobacteria) and some γ -proteobacteria, form the set of the bacteria known as legume's symbionts. There certainly is much to discover, since only 0.5% of known legumes were until now identified specifically for symbiotic relationship. The discovery of new symbionts associated with legumes is expected to create deep changes in the taxonomy of the rhizobia. A literature review of the currently recognized classification of rhizobia is presented in this paper.

Keywords: Rhizobia, Legume, Proteobacteria, Taxonomy, Classification.

1. Introduction

Rhizobia are soil bacteria able to form nodules and establish a symbiosis with the roots or the stems of leguminous plants. During the process of symbiosis, rhizobia reduce atmospheric nitrogen into a form directly assimilated by plants (ammonium).

The ability of rhizobia to fix nitrogen significantly reduced the use of chemical fertilizers in agriculture. In fact, nearly 90% of the nitrogen plant needs are well satisfied and the soil is enriched with nitrogen that will be used by subsequent crops, with no need to add chemical fertilizers. In general, this symbiosis has several advantages including improved agricultural productivity, maintenance and restoration of soil fertility, safety of expensive fertilizers currency and energy and limitation of groundwater's pollution by nitrates playing therefore a significant ecological and economical function.

The study of the rhizobial diversity is a valuable biological resource and attempts to find bacterial strains with interesting features to maximize the legumes productivity [1]. Such studies allow us not only to find new strains of rhizobia, but they also support research efforts to select efficient combinations of rhizobium-legume association.

Because of their ecological and economical importance, the diversity and taxonomy of these microorganisms have been extensively studied over the last twenty years. From one genus including

four species in 1981, the classification now includes at least 13 genus comprising more than 98 species and this number continue to increase [2].

Zakhia et al. (2004) proposed the term BNL (Bacteria Nodulating Legumes) to avoid confusion between the general term of rhizobium and genus name. Currently, all BNL described so far belong to the Proteobacteria class. The majority of them belong to genus of the α -Proteobacteria class, called *Rhizobium*, *Mesorhizobium*, *Bradyrhizobium*, *Azorhizobium*. In addition, new types were found in the α -Proteobacteria class, namely *Methylobacterium*, *Devosia*, *Microvirga*, *Ensifer* (formerly *Sinorhizobium*), *Ochrobactrum*, *Phyllobacterium* and *Shinella* [3].

The BNL were also found in the β -Proteobacteria class, such as *Burkholderia*, *Cupriavidus* (formerly *Ralstonia*) [4]. Finally, several studies reported the presence of γ -Proteobacteria class in black locust by the *Pseudomonas* sp. [5].

More than 19,700 legume species grow around the world, but only a few microsymbionts have been studied [6]. The increase in advanced study of legume's new species in different geographical regions opens new perspectives to isolate and to characterize more species of *Rhizobium*.

2. Phylogeny of rhizobia

The comparison of data obtained from bacterial strains or species with different molecular approaches allows the identification of groups of taxa. Different taxa obtained can be compared and used for tracing lines of descent commonly recognized under the name of phylogenetic tree (Figure 1). Thus, the phylogeny can be defined as the evolutionary history that reveals the existing relationships between groups of organisms with an ancestor located at a higher taxonomic level.

Nowadays, use of 16S rRNA sequence is the main tool in the study of microbial phylogeny. However, it has limitations to differentiate among close species and for this purpose several metabolic genes (housekeeping) have been proposed in several groups of bacteria. In rhizobia the two first genes analyzed were *recA* and *atpD* and currently they have been sequenced in many rhizobial species showing their usefulness in differentiation of species whose 16S rRNA genes are nearly identical. In the last years (mention duration) new schemes for identification and phylogenetic analysis of bacteria named MLSA (Multilocus sequence analysis) and MLST (Multilocus sequence typing) based in the analysis of several housekeeping genes have been applied for phylogenetic analyses of concrete groups of rhizobia [7].

Besides the housekeeping genes also named "core" genes, some "auxiliary" or "accessory" genes involved in the legume symbiosis are commonly included in species description of rhizobia and in some MLST analysis. The symbiotic genes from rhizobia are codified in plasmids in fast and some middle growing species and in symbiotic islands in the middle and slow growing ones. From these genes, the most studied in rhizobia are *nodD*, *nodA*, *nodC* and *nifH*. Nevertheless, the symbiotic genes are not useful in taxonomy because due to their ability to be transferred in nature from plasmids to islands, from bacteria to plants and among bacteria. Therefore the analysis of symbiotic genes is

overall useful to identify non-rhizobial species able to nodulate legumes and to carry out biogeography studies of legume endosymbionts [7].

3. Polyphasic taxonomy

Originally based on phenotypic criteria, the application of polyphasic terminology has been reviewed by the revolution of different molecular techniques of which mainly the PCR or Polymerase Chain Reaction [8]. Thus, the standard principles were proposed in 1991, for the description of new genus or species on the basis of numerical analysis which is the synthesis of the results of different phenotypic and molecular tests [9]. This new definition of the polyphasic approach allowed a proper systematic study of major bacterial groups, an assessment of taxonomic resolution of different techniques of diversity study [10] ~~—(Figure 2)~~ and an establishment of comparable databases between different applied microbiology laboratories [11].

The most commonly used molecular techniques are ~~elaborated in Figure 2~~: FT-IR technique (Fourier-transform infrared spectroscopy), which allows to study the diversity of bacteria at an intraspecific level [12], the technique of typing by PCR target (target PCR fingerprinting) [13] and especially the technique of DNA chips commonly known as DNA microarrays. In this last technique, the conception of the chips permits to detect simultaneously thousands of sequences or even to cover the entire genome of an organism [14]. By the same technique, Rüberg et al. (2003) have analyzed the entire genome of *Sinorhizobium meliloti* [15]. All these techniques can reveal more information about the diversity of rhizobia and can therefore provide more data available for their phylogenetic study.

~~Other~~ ~~News~~ methods are potential alternatives to DNA-based methods for rapid and reliable characterization of bacteria. ~~These most commonly used methods are:~~ Matrix-assisted laser desorption ionization–time-of-flight mass spectrometry (MALDI-TOF MS) ~~has been suggested as a fast and reliable method~~ for bacterial identification, based on the characteristic protein profiles for each microorganism. Using this technology it has been estimated that up to 99% of strains tested are correctly identified when ~~compared~~ with commercial phenotypic identification panels or *rrs* gene sequencing [16].

Thus, modern bacterial taxonomy is based on the integration of all phenotypic, genotypic and phylogenetic data for a more stable classification [17; 10]. The following paragraphs will discuss the current classification of rhizobia with this new approach.

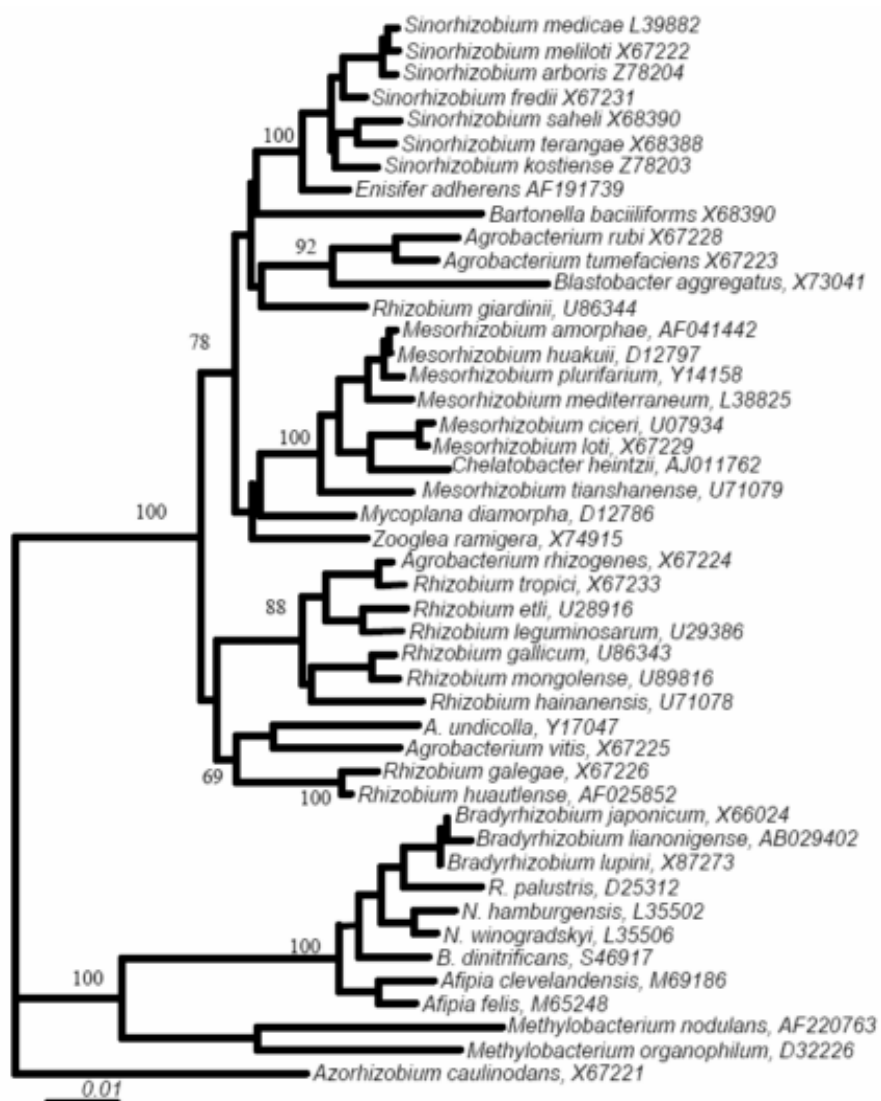


Figure 1: phylogenetic tree of rhizobia and related species built by the neighbor-joining method from the complete sequence of 16S rRNA [18].

	Family	Genus	Species	Strain
Labeling of DNA probes				
DNA sequencing				
Phenotypic analysis (APL...)				
RNA sequencing				
ADN:ARNr hybridization				
Chemotaxonomic analysis				
FAME				
ADN:ADN hybridization				
PCR/ARNr				
G+C%				
RFLP-ARDRA				
RAPD-DAF-APPCR				
AFLP				
REP-PCR				
Proteomic analysis of total cellular proteins				
Isoenzyme polymorphism				
Phage typing				
MLEE				
Serology				

Figure 2: Taxonomic resolution of the various techniques used to study the diversity of rhizobia [10;

4. Current classification of rhizobia

The first classifications were based on cross-inoculation tests between rhizobia and host plants for their identification. The host plant was not the only criteria taken into account for the classification of rhizobium which species were classified into two genus: the genus *Rhizobium* corresponding to strains with fast growth and genus *Bradyrhizobium* for slow-growing strains, based on their generation time and rate of growth on culture medium [20].

However, discordant observations between the notion of bacterial growth speed and the host range showed a lot of doubt on the validity of this classification. This makes place otherwise to comparative methods such as the serology, the coefficient of Chargaff, RNA / DNA or DNA / DNA, hybridization analysis of plasmids, etc...

This period (specify period) marked the beginning of a new taxonomy based on the comparison of results from different phenotypic and biochemical analysis for the identification of symbiotic bacteria [21]. Since then, the isolation of rhizobia from an increasing number of plant species in the world and their characterization by modern polyphasic taxonomy has led to the description of other new genera and species.

The current literature describes more than 98 species grouped in 11 genus belonging to the subclass α -Proteobacteria and 2 genus belonging to the order of Burkholderiales in subclass β -Proteobacteria [17; 15; 22; 23], and finally, one genus belonging to the order Pseudomonadales in the subclass γ -Proteobacteria [24].

The complete list of valid species of rhizobia is recorded in the List of Prokaryotic Names with Standing in Nomenclature that is prepared, performed and revised by Dr. Euzéby (<http://www.bacterio.cict.fr>). According to this list currently the genus *Rhizobium* contains 33 species and includes ~~to the~~ former genera *Agrobacterium* and *Allorhizobium*. The genus *Sinorhizobium*, currently named *Ensifer*, contains 13 species, the genus *Shinella*, contains 1 specie, the genus *Mesorhizobium*, 29 species, the genus *Bradyrhizobium*, 15 species, the genus *Phyllobacterium*, 1 specie and the genus *Azorhizobium*, 3 species, the genus *Merhylobacterium*, 1 specie, the genus *Microvirga*, 3 species.

This high number of species able to nodulate legumes (Table 1) was unexpected only two decades ago and the previsions are that it will be increased in next times.

In summary, on the basis of the DNA ribosomal 16S sequence, the legume's symbionts belong to three main distinct phylogenetic subclasses (Figure 3).

Table 1: Current classification of different species of rhizobia [28]

Genus	Species	Host plant	References
Class :Alphaproteobacteria			
Order :Rhizobiales			
Family :Rhizobiaceae			
<u>Genus: Rhizobium</u>			
<i>R. leguminosarum</i>			[25]
<i>biovar viciae</i>	<i>Pisum, viciae, Lens, Lathyrus</i>		[25; 26]
<i>biovar trifolii</i>	<i>Trifolium pratense</i>		[29; 26]
<i>biovar phaseoli</i>	<i>Phaseolus vulgaris</i>		[25; 26]
<i>R. galegae</i>	<i>Galega, Leucaena</i>		[27]
<i>biovar officinalis</i>	<i>Galega orientalis</i>		[27]
<i>biovar orientalis</i>	<i>Galega officinalis</i>		[27]
<i>R. tropici</i>	<i>Phaseolus, Medicago, Macroptilium</i>		[28]
type IIA (<i>R. leucaenae</i>)			[29]
type IIB			[28]
<i>R. endophyticum</i>	<i>Phaseolus vulgaris</i>		[30]
<i>R. phaseoli</i>	<i>Phaseolus</i>		
<i>R. fabae</i>	<i>Vicia faba</i>		[31]
<i>R. etli</i>	<i>Phaseolus,</i>		[32]
<i>biovar mimosae</i>	<i>Mimosa affinis</i>		[33]
<i>biovar phaseoli</i>	<i>Phaseolus,</i>		[24]
<i>R. undicola</i>	<i>Neptunia natans</i>		[35]
<i>R. gallicum</i>	<i>Phaseolus vulgaris</i>		[36]
<i>biovar phaseoli</i>	<i>Phaseolus vulgaris</i>		[36]
<i>biovar gallicum</i>	<i>Phaseolus vulgaris</i>		[36]
<i>R. giardinii</i>	<i>Phaseolus vulgaris</i>		[36]
<i>biovar phaseoli</i>	<i>Phaseolus,</i>		[36]
<i>biovar giardinii</i>	<i>Phaseolus vulgaris</i>		[36]
<i>R. hainanensis</i>	<i>Desmodium sinuatum, Centrosema, etc.</i>		[37]
<i>R. huautlense</i>	<i>Sesbania herbacea</i>		[38]
<i>R. mongolense</i>	<i>Medicago ruthenica, Phaseolus</i>		[39]
<i>R. yanglingense</i>	<i>Amphicarpaea</i>		[40]
<i>R. larrymoorei</i>	<i>Ficus benjamina</i>		[41]
<i>R. indigoferae</i>	<i>Indigofera spp.</i>		[42]
<i>R. sullae</i>	<i>Hedysarum</i>		[43]
<i>R. loessense</i>	<i>Astragalus, Lespedeza</i>		[44]
<i>R. cellulosilyticum</i>	<i>Populus alba</i>		[45]
<i>R. miluonense</i>	<i>Lespedeza</i>		[46]
<i>R. multihospitium</i>	Multiple legume species		[47]
<i>R. oryzae</i>	<i>Oryza alta</i>		[48]

<i>R. pisi</i>	<i>Pisum sativum</i>	[49]
<i>R. mesosinicum</i>	<i>Albizia, Kummerowia Dalbergia</i>	[50]
<i>R. alamii</i>	<i>Arabidopsis thaliana</i>	[51]
<i>R. alkalisoli</i>	<i>Caragana intermedia</i>	[52]
<i>R. tibeticum</i>	<i>Trigonella archiducis-nicolai</i>	[53]
<i>R. tubonense</i>	<i>Oxytropis glabra</i>	[54]
<i>R. halophytocola</i>	Coastal dune plant	[53]
<i>R. radiobacter</i>	*	[55]
<i>R. rhizogenes</i>	*	[55]
<i>R. rubi</i>	*	[55]
<i>R. vitis</i>	*	[55]
<i>R. nepotum</i>	*	[56]
Genus: <i>Ensifer</i>		
<i>E. meliloti</i>	<i>Medicago, Melilotus, Trigonella</i>	[57]
<i>E. fredii</i>	<i>Glycine, Vigna, Cajanus</i>	
biovar <i>fredii</i>	<i>Glycine, Vigna, Cajanus</i>	[58]
biovar <i>siensis</i>	<i>Glycine</i>	[37]
<i>E. saheli</i>	<i>Acacia, Prosopis, Neptunia, Leucaena</i>	[59]
<i>E. teranga</i>	Different host plants	[59]
biovar <i>acaciae</i>	<i>Acacia</i>	[55]
biovar <i>sesbania</i>	<i>Sesbania</i>	[60]
<i>E. medicae</i>	<i>Medicago truncatula, Melilotus</i>	[61]
<i>E. arboris</i>	<i>Acacia, Prosopis</i>	[62]
<i>E. kostiense</i>	<i>Acacia, Prosopis</i>	[62]
<i>E. xingianense</i>	<i>Glycine max</i>	[63]
<i>E. adhaerens</i>	<i>Leucaena leucocephala</i>	[64]
<i>E. kummerowiae</i>	<i>Kummerowiae stipulaceae</i>	[42]
<i>E. americanum</i>	<i>Acacia</i>	[65]
<i>E. mexicanus</i>	<i>Acacia angustissima</i>	[66]
<i>E. numidicus</i>	<i>Medicago sativa</i>	[67]
Genus : <i>Shinella</i>		
<i>S. kummerowiae</i>	<i>Kummerowia stipulacea</i>	[50]
Family : <i>Phyllobacteriaceae</i>		
Genus : <i>Mesorhizobium</i>		
<i>M. loti</i>	<i>Lotus, Cicer, Anthyllis, Astragalus, etc.</i>	[68]
<i>M. huakuii</i>	<i>Astragalus sinicus</i>	[69]
<i>M. ciceri</i>	<i>Cicer arietinum</i>	[70]
<i>M. tianshanense</i>	<i>Glycyrrhiza pallidiflora</i>	[63]

<i>M. mediterraneum</i>	<i>Cicer arietinum</i>	[71]
<i>M. plurifarium</i>	<i>Acacia, Chamaecrista, Leucaena, Prosopis,</i>	[72]
<i>M. amorphae</i>	<i>Amorpha fruticosa</i>	[73]
<i>M. chacoense</i>	<i>Prosopis alba</i>	[74]
<i>M. septentrionale</i>	<i>Astragalus adsurgens</i>	[75]
<i>M. temperatum</i>	<i>Astragalus adsurgens</i>	[75]
<i>M. thioganicum</i>	*	[75]
<i>M. albiziae</i>	<i>Albizia kalkora</i>	[75]
<i>M. caraganae</i>	<i>Caragana spp.</i>	[64]
<i>M. gobiense</i>	Wild legumes	[47]
<i>M. tarimense</i>	Wild legumes	[47]
<i>M. australicum</i>	<i>Biserrula pelecinus L.</i>	[76]
<i>M. opportunistum</i>	<i>Biserrula pelecinus L.</i>	[76]
<i>M. metallidurans</i>	<i>Anthyllis vulneraria</i>	[77]
<i>M. alhagi</i>	<i>Alhagi</i>	[78]
<i>M. camelthorni</i>	<i>Alhagi sparsifolia.</i>	[78]
<i>M. abyssinicae</i>	Different agroforestry legume trees	[79]
<i>M. muleiense</i>	<i>Cicer arietinum L.</i>	[80]
<i>M. hawassense</i>	Different agroforestry legume trees	[79]
<i>M. qingshengii</i>	<i>Astragalus sinicus</i>	[81]
<i>M. robiniae</i>	<i>Robinia pseudoacacia</i>	[82]
<i>M. shonense</i>	Different agroforestry legume trees	[79]
<i>M. shangrilense</i>	<i>Caragana species</i>	[83]
<i>M. silamurunense</i>	<i>Astragalus species</i>	[84]
<i>M. tamadayense</i>	<i>Anagyris latifolia, Lotus berthelotii</i>	[85]
Genus : <i>Phyllobacterium</i>		
<i>P. trifolii</i>	<i>Trifolium pratense</i>	[86]
Family : <i>Methylobacteriaceae</i>		
Genus : <i>Methylobacterium</i>		
<i>M. nodulans</i>	<i>Crotalaria spp</i>	[87]
Genus : <i>Microvirga</i>		
<i>M. lupini</i>	<i>Lupinus sp.</i>	[88]
<i>M. lotononidis</i>	Different legume host	[88]
<i>M. zambiensis</i>	Different legume host	[88]
Family : <i>Brucellaceae</i>		
Genus : <i>Ochrobactrum sp.</i>		
<i>Ochrobactrum cytisi</i>	<i>Cytisus</i>	[89]
<i>Ochrobactrum lupini</i>	<i>Lupinus albus</i>	[90]
Family: <i>Hyphomicrobiaceae</i>		
Genus: <i>Azorhizobium</i>		
<i>A. caulinodans</i>	<i>Sesbania rostrata</i>	[91]
<i>A. johannense</i>	<i>Sesbania virgata</i>	[91]
<i>Azorhizobium sp</i>	<i>Sesbania rostrata</i>	[91]

Genus: Devosia		
<i>Devosia neptuniae</i>	<i>Neptunia natans</i>	[92]
Family : Bradyrhizobiaceae		
Genus : Bradyrhizobium		
<i>B. Japonicum</i>	<i>Glycine max, Glycine soja</i>	[26 ; 93]
<i>B. Elkanii</i>	<i>Glycine max</i>	[94]
<i>B. Liaoningense</i>	<i>Glycine max</i>	[95]
<i>B. Yuanmingense</i>	<i>Lespedeza</i>	[96]
<i>B. Betae</i>	<i>Betae vulgaris</i>	[97]
<i>B. Canariense</i>	<i>Genisteae et Loteae</i>	[98]
<i>B. iriomotense</i>	<i>Entada koshunensis</i>	[99]
<i>B. jicamae</i>	<i>Pachyrhizus erosus</i>	[49]
<i>B. lablabi</i>	<i>Lablab purpureus</i>	[99]
<i>B. huanghuaihaiense</i>	Soybean	[100]
<i>B.cytisi</i>	<i>Cytisus villosus</i>	[101]
<i>B.daqingense</i>	Soybean	[102]
<i>B. denitrificans</i>	<i>Aeschynomene</i>	[103]
<i>B.oligotrophicum</i>		[104]
<i>B.pachyrhizi</i>	<i>Pachyrhizus erosus</i>	[105]
Class : Beta Proeobacteria		
Order : Burkholderiales		
Family: Burkholderiaceae		
Genus : Burkholderia		[106]
<i>B. caribensis</i>	<i>Vertisol microaggregates</i>	[107]
<i>B. cepacia</i>	<i>Alysicarpus glumaceus</i>	[4]
<i>B.tuberum</i>	<i>Aspalatus carnosa</i>	[108]
<i>B. phymatum</i>	<i>Aspalatus carnosa</i>	[108]
<i>B.nodosa</i>	<i>Mimosa bimucronata, Mimosa scabrella</i>	[109]
<i>B. sabiae</i>	<i>Mimosa caesalpiniiifolia</i>	[110]
<i>B.mimosarum</i>	<i>Mimosa spp.</i>	[111]
<i>B.rhizoxinica</i>	<i>Rhizopus microsporus</i>	[112]
<i>B.diazotrophica</i>	<i>Mimosa spp.</i>	[113]
<i>B.endofungorum</i>	<i>Rhizopus microsporus</i>	[112]
<i>B.heleia</i>	<i>Eleocharis dulcis</i>	[114]
<i>B.symbiotica</i>	<i>Mimosa spp.</i>	[115]
Genus : Cupriavidus	<i>Aspalatus carnosa</i>	
<i>C. taiwanensis</i>	<i>Mimosa sp.</i>	[116]
Class: Gamma-Proteobacteria		
Order: Pseudomonadales		
Family: Pseudomonaceae		
<i>Pseudomonas sp.</i>	black locust	[5]

*Species with no described nodulation ability included in traditionally considered rhizobial genera.

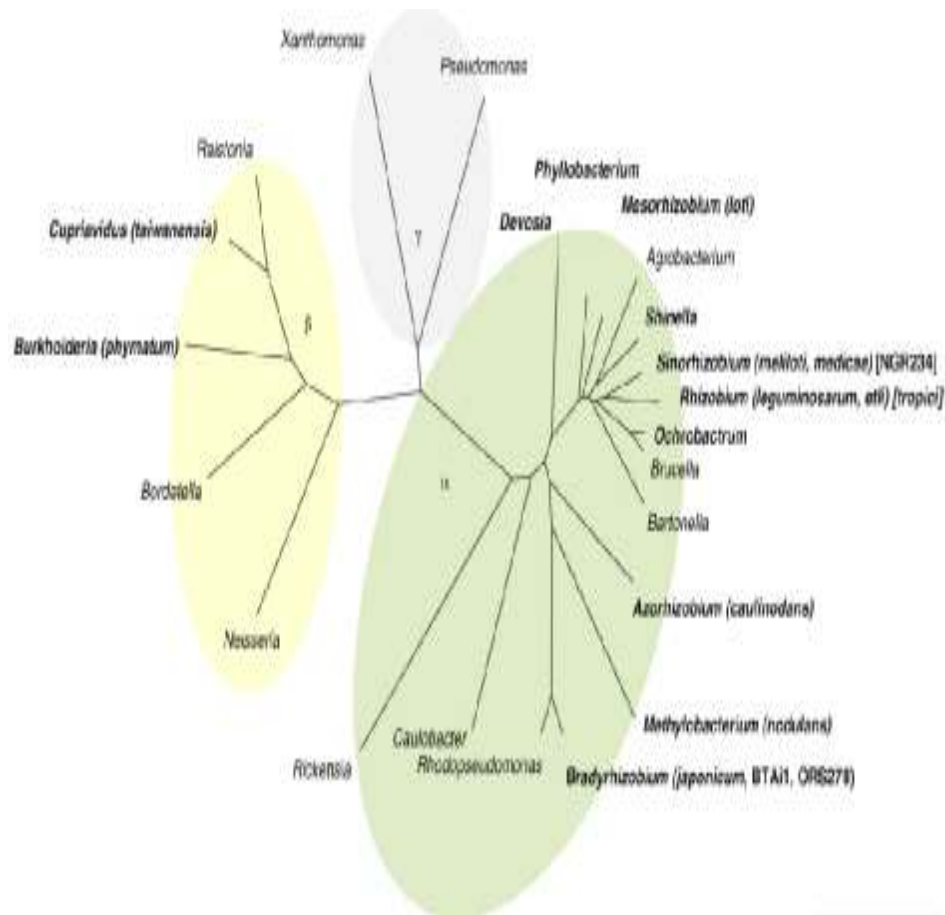


Figure 3: Simplified phylogenetic tree of the 16S rRNA of proteobacteria. The genus shown in bold contain rhizobia [117].

4.1. Alpha-subclass of Proteobacteria

4.1.1. Rhizobium /Ensifer/ Shinella branch

This branch is subdivided in **three** sub branches in the genus: *Rhizobium* [25], *Ensifer* (formerly *Sinorhizobium*) [37; 59], and *Shinella* [50]. These genera are grouped with:

- *Agrobacterium*, non-symbiotic bacterial genus involved in pathogenic interactions with plants;
- Bacteria ameliorative plant growth (PGPR: Plant growth-promoting rhizobacteria) as *Azospirillum*,

- Human and animal pathogens including *Brucella*, *Ochrobactrum*, *Bartonella* or soil bacteria as *Mycoplana*.

The first sub-branch corresponds to genus *Rhizobium*, defined as a group of thirty species, from various hosts, and includes *R. leguminosarum*, *R. tropici* [28], *R. etli* [32], *R. gallicum* [36], *R. mongolense* [39], *R. undicola* [35] and *Agrobacterium*. The members of this genus are short Gram-negative rods, of a 0.5 to 0.9 µm width and a 1.2 to 3 µm length, often with an uncolored region due to the presence of polymer β-hydroxybutyrate (PHB). Rhizobia do not form endospores but are motile with polar or peritrichous flagella (4 to 6), aerobic, chemo-organotrophs. They become pleomorphic under adverse conditions [118]. The paragraph below is not properly connected to preceding paragraph

There are a number of species present in these rhizobial genera that have not been observed to form nodules, and therefore do not fit the functional definition of rhizobia. These include many of the species that were formerly known as *Agrobacterium* (e.g. *R. larrymoorei*, *R. rubi*, and *R. vitis*) [55]. However, there is recent evidence that other species formerly classified as *Agrobacterium* are capable of nodulation. For example *R. radiobacter* nodulates *Phaseolus vulgaris*, *Campylotropis* spp., *Cassia* spp. [119], and *Wisteria sinensis* [120]. Both nodules and tumours were formed on *Phaseolus vulgaris* by *R. rhizogenes* strains containing a Sym plasmid [121].

The second sub-branch corresponds to genus *Ensifer* (formerly *Sinorhizobium*) and includes the species *E. fredii*, *E. meliloti*, *E. teranga* and *E. sahelense* [59; 122], *E. medicae* [61], *E. kostense* and *E. arboris* [62]. Rods of this genus are 0.5 to 0.9 µm wide and 1.2 to 3 µm length, motile by polar flagellum or some peritrichous flagella, Gram-negative, aerobic accumulating PHB. Optimal growth of this genus is at 25-30°C (10-35°C) and at pH 6-8 (5 -10.5) and tolerates 10 g/l of NaCl [123].

Sinorhizobium was proposed as the new genus of the fast-growing soybean rhizobia. This genus is separated from *Rhizobium* and *Bradyrhizobium* based on GC content, DNA-DNA hybridization data, numerical taxonomy, serological analysis data, composition of extracellular gum, bacteriophage typing data and soluble protein pattern. However, *Rhizobium* and *Sinorhizobium* are phylogenetically fairly closely related to each other. Recently, the Judicial Commission of the International Committee on Systematics of Prokaryotes decided to transfer some species of the genus *Sinorhizobium* to the genus *Ensifer* [124]. Check the fact. The use of *Sinorhizobium* as a genus nomenclature has been proposed to be discontinued.

The third sub-branch corresponds to genus *Shinella* and include one specie: *S. kummerowiae* [50], a symbiotic bacterium nodulating *Kummerowia stipulacea*. Cells of the strain are Gram-negative, strictly aerobic, non-spore-forming, motile short rods [125].

In 2001, Young et al. had revised taxonomic data and showed that the discriminatory phenotypic characteristics to distinguish between these types are not convincing, and that the phylogenetic relationships between genera deduced from the comparative analysis of 16S rDNA sequences differ depending on the chosen algorithm and more especially on the chosen sequences [55]. Therefore, despite phenotypic differences, these authors suggested to combine the three genera *Rhizobium*, *Agrobacterium* and *Allorhizobium* in a single genus sfe: *Rhizobium*, including all species of

Agrobacterium and *Allorhizobium* as new combinations: *R. radiobacter*, *R. rhizogenes*, *R. rubi*, *R. undicola* and *R. vitis*.

4.1.2. Mesorhizobium branch

A new genus proposed in 1997 to include the species of intermediate growth named *Mesorhizobium* [126] was included in the family *Phyllobacteriaceae* that also included the former genus *Phyllobacterium* (with one specie: *Phyllobacterium trifolii* isolated from *Trifolium pratense* nodules) [86].

In this branch *Mesorhizobium* [126] contains, *M. loti* [68], *M. huakuii* [69], *M. ciceri* [70], *M. mediterraneum* [71], *M. tianshanense* [63], *M. plurifarum* [72] and *M. amorphae* [127]. They are phylogenetically related and distinct from the large phylogenetic grouping that includes *Rhizobium*, *Agrobacterium* and *Sinorhizobium* [126]. They are characterized by a growth rate intermediate between the fast- and slowgrowing rhizobia. They are Gram-negative rods, motile by polar or subpolar flagellum, aerobic, accumulating PHB. They assimilate glucose, rhamnose, and sucrose with acid production [123].

4.1.3. Azorhizobium branch

This family regroups only one genus and only one species of symbiotic bacterium: *Azorhizobium caulinodans*. They are short rods very similar to *Rhizobium*, who present a polar and peritrichous ciliation but produce an alkaline reaction on glucose as *Bradyrhizobium*. Only this sugar is used. Strains also degrade fatty acids, organic acid and alcohols and are the only ones to use alcohols. Their growth is intermediate between the two previous genus, Generation time = 7-9 h. No strain is denitrifying. They grow up to 43 °C. They have only the dihydrolase arginine and lysine decarboxylase. They do not use mannitol, conventional substrate of the other two genera, but lactate gives good growth yields. They seem more sensitive to antibiotics [123]. In addition, it doesn't only form nodules on root parts but also on aerial parts of *Sesbania rostrata* [91]. A second species *Azorhizobium johannae* was proposed [92]. However, this species is characterized by a low level of DNA / DNA hybridization compared to the typic strain of the genus.

The analysis of the 16S rRNA sequences showed that the typic strain is mixed with species of the genus *Xanthobacter* and *Aquabacters*. The combination of these last two genres with *Azorhizobium* was investigated without being suggested (Do you mean that the proposal to merge genera *Xanthobacter*, *Aquabacter* and *Azorhizobium* was not accepted) because of the great dissimilarity of phenotypic traits [128; 129].

4.1.4. Bradyrhizobium branch

This genus has been defined to include all slow-growing rhizobia [26]. These are rods with a single polar or subpolar flagellum. The colonies do not exceed 1 mm diameter in agar + mannitol. These symbiotic bacteria use many sugars and organic acids but prefer pentoses, with production of polysaccharidic mucus. Some species can grow as chimolithotrophic in the presence of H₂, CO₂ and

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low content in O₂ due to the presence of an hydrogenase, Generation time = 10-12h. Some strains can fix N₂ *in vitro*. It's more resistant to antibiotics than *Rhizobium* [123].

This branch has included, for a long time, a single species: *Bradyrhizobium japonicum* that regrouped all strains nodulating the soybean (*Glycine max*). Very heterogeneous, it was then divided into three groups (I, Ia and II), based on homologies obtained by DNA/DNA hybridization [130]. A new species: *Bradyrhizobium elkanii* was created in the Group II, which differs from the species *Bradyrhizobium japonicum* [94]. For other very slow growing strains, isolated from nodules of soybean *Glycine max* and *Glycine soya* in China, *Bradyrhizobium liaoningense* species has been proposed [95]. The *Bradyrhizobium* genus includes strains for which the taxonomic position is not clearly defined at the species level. The discrepancy between *Bradyrhizobium japonicum* and *Bradyrhizobium elkanii* is clear. It is more important than the divergence between *Bradyrhizobium* and non-symbiotic species as: *Aifipia* (animal bacterial pathogens), *Nitrobacter* (nitrifying bacteria in the soil), *Rhodopseudomonas palustris* (photosynthetic bacteria) and *Blastobacter denitrificans* [106].

In recent years, the characterization of different strains of *Bradyrhizobium* by comparing several molecular techniques revealed the existence of 11 different genotypes, three of which correspond to known strains, while 8 genotypes are very distinct.

A new species *B. yuanmingense* was isolated from the genus *Lespedeza* [96; 131]. In the same year, a strain of *Bradyrhizobium* nodulating a wild plant of the genus *Phaseolus* was reported [132]. Other recognized species of this group are *Bradyrhizobium betae* from the roots of *Beta vulgaris* afflicted with tumor-like deformations [97] and *Bradyrhizobium canariense* from genistoid legumes from the Canary Islands [98]. Many other slow-growing rhizobia were isolated from other legume hosts and are commonly referred to as *Bradyrhizobium* sp., followed by the name of the legume host: *B. lablabi* was isolated from the genus *Lablab purpureus* [128], *B. cytisi* has been isolated from the genus *Cytisus villosus* [101] and *B. pachyrhizi* was isolated from effective nodules of *Pachyrhizus erosus* [105].

Based on ITS sequence data, the photosynthetic bradyrhizobia isolated from stem-nodules of *Aeschynomene*, form a distinct group closely related to *Blastobacter denitrificans* [106]. As a result of a comprehensive study of both groups, van Berkum *et al.* (2006) recently proposed to transfer *Blastobacter denitrificans* to *Bradyrhizobium* and unite the species with the isolates from *Aeschynomene indica* as the species *Bradyrhizobium denitrificans* [103].

Recently, A new species *B. Canariense* from *Genisteae Loteae* [98], *B. Iriomotense* from *Entada koshunensis* [99], *B. jicamae* from *Pachyrhizus erosus* [85], *B. daqingense* and *B. huanghuaihaiense* from soybean was reported [102].

4.1.5. Methylobacterium branch

The *Methylobacterium* genus [133], has included a single species: *M. nodulans* that regrouped all strains nodulating the *Crotalaria*. Bacteria of this genus have the ability to grow on single-carbon compounds such as formate, formaldehyde, methylamine and methanol as only source of carbon and energy. These are chemo-organotrophic and optional methylotrophic, able to use multi-carbon compounds for growth. The cells of this genus present the rods form (0.8 - 1.0 x 1.0 to 3.0 mm), most frequently isolated and occasionally in rosette. They are often branched or pleomorphic, especially in

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cultures in late stationary phase. They are strictly aerobic bacteria, Gram-negative, oxidase and catalase positive and have a typical pink pigmentation due to the presence of carotenoids [134]. *Methylobacterium* strains are commonly called "pink-pigmented facultative Methylophs" (PPFM). However, at least one case, *M. nodulans* is not pigmented [100]. They are mesophilic and optimal temperature is between 25 and 30°C. All these strains are motile with a single polar flagellum. Often, cells contain inclusions of poly-β-hydroxybutyrate and sometimes inclusions of polyphosphates [135].

4.2. Beta-subclass of Proteobacteria:

4.2.1. Burkholderia branch :

The *Burkholderia* [136] genus has been proposed in 1992 by Yabuuchi and al (cite reference properly). The definition of this was based primarily on genomic considerations and on cellular lipid's composition. Regarding the main phenotypic characteristics, they are right bacilli, Gram negative, usually accumulating granules of poly-beta-hydroxybutyrate, motile with one or more polar flagella (*Burkholderia mallei* is however lacking flagella and stationary), strictly aerobic, catalase positive, oxidase variable depending on the species, can grow using as sole carbon source glucose, glycerol, inositol, galactose, sorbitol, mannose (unlike *Ralstonia sp.* that do not use this sugar) and mannitol. The different species of the genus can be differentiated by the results of auxanogrammes (137 substrates were used in the study of Gillis et al. 1995) [137].

This genus contains 39 species including rhizospheric bacteria, human pathogenic bacteria, as well as nitrogen fixing bacteria as *Burkholderia vietnamiensis* and *Burkholderia kururiensis*. Nevertheless, the genus also includes important species important in environmental management to the ecology of environment. In particular *Burkholderia xenovorans* (formerly known as *Pseudomonas cepacia*, then *Burkholderia cepacia* and *Burkholderia fungorum*) was studied for its ability/faculties to of degrade terioration-of the organochlorine links in pesticides and polychlorobiphenyl (PCB).

Following the discoveries of Moulin et al. (2001) and Chen et al. (2003) [4; 111], showing the ability of bacteria to the subclass β-Proteobacteria (*Burkholderia* and *Ralstonia*) to induce nodulation in legumes, bacteria of the subclass γ-Proteobacteria (*Pseudomonas sp.*) were also isolated from legume nodules [24]. Based on these findings, it is not excluded to find other bacteria able to nodulate legumes, even outside the Proteobacteria phylum.

4.3. Other nitrogen-fixing legume symbionts

Recently, a number of isolates have been reported from legume nodules, capable of nitrogen fixation but phylogenetically located outside the traditional groups of rhizobia in the α-Proteobacteria. New genus/lines that contain nitrogen-fixing legume symbionts include *Devosia*, *Ochrobactrum* and *Microvirga* in the α-Proteobacteria and *Cupriavidus* in the β-Proteobacteria.

In the α-Proteobacteria, *Devosia neptuniae* was proposed for strains from *Neptunia natans* from India [82]. *Ochrobactrum lupini* was described for nodule isolates from *Lupinus Albus* [90] and *Ochrobactrum cytisi* was described for nodule isolates from *Cytisus* [89]. *Microvirga lupini* was described for nodule isolates from *Lupinus sp.*, *M. lotononidis* and *M. zambiensis* were described for nodule isolates from different legume host [88].

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In 2003 the nodulation of *Mimosa* by *Ralstonia taiwanensis* or *Wautersia taiwanensis* was published [86]. This species was isolated from nodules of *Mimosa pudica* and *Mimosa diplotricha*. In this case the strain was erroneously classified as *Ralstonia* and it has been later named *Cupriavidus taiwanensis* [116]. At present, *Cupriavidus* is a β - Proteobacteria, belonging to family *Burkholderiaceae* of order *Burkholderiales*. Several species of this genus were isolated from soil and human clinical specimens but *C. taiwanensis* was the only species able to form effective nodules and fixes atmospheric nitrogen in legumes.

5. Conclusion

From one genus and 4 species in 1981, the rhizobia taxonomic studies have led to a total of 13 genera and 98 recognized rhizobial species. This is in constant progress due to various studies in the world. Progress in taxonomy is also due to increasing numbers of effective techniques available in the characterization of bacteria, more accessible to laboratories.

So far only 23% of the total number of legumes species (between 16 500 and 19 700) were characterized for their microsymbionts. The majority (88%) of the studied legumes proved to be able of forming nodules. Specifically, tropical rhizobia are still poorly documented compared to what can be expected based on recent data, which show a great diversity in China, Brazil, Senegal, Sudan, Morocco.

In 2001, Sy et al. reported that some rhizobia isolated from *Crotalaria* sp. in Senegal were phylogenetically separated from all known rhizobia. They are a new species of the genus *Methylobacterium*, for which the name of *M. nodulans* was proposed. These finding suggests that several other new groups of symbiotic leguminous plant's bacteria (genera, species) may emerge in the future.

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