

1 **Phytochemical study and antimicrobial activity of bark extracts of *Ceiba pentandra* (L.)**
2 **Gaertn. (Bombacaceae) from Côte d'Ivoire on resistant bacteria *Staphylococcus aureus***
3 **and *Pseudomonas aeruginosa***

4 **ABSTRACT**

5 One of the health problems in recent decades is the emergence of multi-antibiotic resistant
6 bacteria. Among there solution tracks of this concern, the secondary metabolites of medicinal
7 plants seem to be privileged. In this context, our work is devoted to the phytochemical study
8 and evaluation of antibacterial activity of bark extracts of *Ceiba pentandra* from Ivorian
9 Pharmacopoeia on *S. aureus* methicill in resistant and *P. aeruginosa* imipenem and
10 ceftazidime resistant. The phytochemical study shows that bark of *Ceiba pentandra* harvested
11 in center of Ivory Coast contain alkaloids, flavonoids, saponins, tannins, steroids, terpenoids
12 and cardiac glycosides. Microbiological testing showed that in liquid medium, ethylacetate
13 (AcOEt) extracted is active on all studied germs. MIC range from 0.78 to 6.25 mg /mL and
14 CMB from 1.04 to 8.33 mg /mL. Purification of AcOEt extract had no influence on his
15 activity against studied germs. In fact the most active fraction (F8) has MIC ranging from 0.52
16 to 6.25 mg/ mL and CMB 1.04 to 10.42 mg /mL. According to the ratio CMB /CMI which was
17 nearly 2, AcOEt extract and fraction F8 are bactericidal.

18 **Keywords:** *Ceiba pentandra*, Phytochemistry, *Staphylococcus aureus*, *Pseudomonas*
19 *aeruginosa*, resistant

20 **1. INTRODUCTION**

21 According to WHO [1], over 80% of the population in Africa still use traditional medicine to
22 meet their health care needs. This appears to be linked to the high cost of modern drugs,
23 distance and / or insufficiency of health units especially in rural areas. Moreover, in hospitals
24 control of bacterial infections is complex due to the emergence of bacteria resistant to many
25 conventional antibiotics. In the literature, many cases of multidrug-resistant bacteria are
26 reported as well in Ivory Coast [2, 3, 4] than in other countries in sub-Saharan Africa
27 [5]. *Staphylococcus aureus* and *Pseudomonas aeruginosa* are a major concern in hospitals
28 because of their epidemic spread [6] and unfortunately they are a part of microorganisms that
29 acquire the multidrug resistance. Furthermore, the conjunction of the lack of new efficient
30 antibiotics and the growth of multidrug resistance bacteria may lead, in the future, to increase
31 therapeutic impasses. So, it is more than appropriate to promote the development and
32 diffusion of treatments which are able to handle with these threats.

33 Cote d'Ivoire, with its 761 medicinal species and 1421 drug recipes that have been identified
34 [7] provides investigative options still poorly exploited. Thus, among the plants of the Ivorian
35 Pharmacopoeia, *Ceiba pentandra* (Bombacaceae) widely used at several levels of traditional
36 medicine. Indeed this plant is use to relieve painful conditions such as fevers, abscesses,
37 paronychia, mental illness, conjunctivitis, dizziness, headache etc [8], as well as in the
38 treatment of wounds and ulcers. In Nigeria, Adebayo-Tayo[9] confirmed the antimicrobial
39 activities of *Ceiba pentandra* local.

40 The present work aims to establish the phytochemical profile of *Ceiba pentandra* from
41 savannah of Côte d'Ivoire and to evaluate the antimicrobial activity of the ethyl acetate extract
42 of his bark against resistant strains of *Staphylococcus aureus* and *Pseudomonas aeruginosa*.

43 2. MATERIALS AND METHODS

44 Fresh bark of *Ceiba pentandra* were collected in December 2010 around the Institut National
45 Polytechnique Felix Houphouet-Boigny in Yamoussoukro (central Côte d'Ivoire). This plant
46 was identified by Mr. Amani N'Guessan botanist from the Institut National Polytechnique
47 Felix Houphouet-Boigny. A sample was deposited in the Herbarium of the Institute.

48 The barks were dried for three weeks out of the sun and then crushed with an electric grinder.
49 The obtained powder was stored in polyethylene bags at 4°C until extraction.

50 The bacterial strains used for testing were provided by the unit Antibiotics, Natural
51 Substances and Surveillance of Resistance of microorganisms to anti-Infective (ASSURMI)
52 of Bacteriology Department at the Institut Pasteur in Côte d'Ivoire (IPCI). It consists of a
53 strain of:

- 54 - *Staphylococcus aureus* methicillin susceptible (*S. aureus* Meti S);
- 55 - *Staphylococcus aureus* resistant to methicillin (*S. aureus* Meti R);
- 56 - *Pseudomonas aeruginosa* ceftazidime and imipenem susceptible (*P. aeruginosa* Cefta
57 S / Imp S);
- 58 - *Pseudomonas aeruginosa* ceftazidime and imipenem resistant (*P. aeruginosa* Cefta R /
59 Imp R).

60 Referenced Strains of *Staphylococcus aureus*(ATCC 25923) and *Pseudomonas*
61 *aeruginosa*(ATCC 27853) were also tested.

62 Before testing, the different bacterial strains were cultured by the method of streaking and
63 incubated in an oven at 37 ° C for 18 to 24 hours to obtain young colonies.

64 **2.1. Extraction and purification of secondary metabolites**

65 In a flask of 3L capacity, protected from light, 1500 grams of crushed bark is macerated at
66 room temperature using 2.5L of hexane for 12 hours. This operation aimed to dewax the
67 powder and it have been repeated twice. After filtration the dried residual powder was
68 exhaustively extracted with ethyl acetate under the same conditions. At each cycle of
69 extraction, the ratio plant material - solvent was maintained at 1: 5 (w / v). The organic phases
70 were filtered through cotton wool and then on Whatman paper N°3. The filtrates were
71 evaporated to dryness at 40°C under reduced pressure using a rotavapor Buchi 161. A 12 g of
72 brown powder was obtained corresponding to 0.80% yield.

73 A mass of 10 g of extract in ethyl acetate was fractionated on a silica gel column [Merck-
74 Silica gel 60 (0.063-0.200 mm)] eluting with hexane, to ethyl acetate, and methanol in the
75 proportions 100/0/0; 50/50/0; 0/100/0; 0/50/50 (v / v / v). Fractions were collected following
76 the chromatographic profiles and after solvent evaporation, dry products were stored at 4°C
77 under nitrogen until achievement of phytochemicals and antimicrobial tests.

78 **Chemicals used:** All chemicals and drugs used were obtained commercially and of analytical
79 grade.

80 **2.2. Evaluation of antibacterial activity**

81 **2.2.1. Efficacy of substances**

82 Efficiency tests were used to detect the antimicrobial activity of the substances. For these tests,
83 the Mueller Hinton agar was the main culture medium [10, 11, 12]. For preparing extracts a
84 mixture of DMSO / distilled water in the ratio 1: 1 (v / v) was used. Non-impregnated discs of
85 6 mm of diameter, purchased from Polychimie were used. The tests were performed on
86 bacterial inoculums of $5 \cdot 10^6$ CFU/ mL.

87 Each disc was impregnated with 40µL of extract or fractions solutions at 200 mg/mL
88 concentration. The choice of 200mg/mL concentration for this test was literature guided.
89 After drying, the discs were placed on the agar previously seeded with micro bacterial strains
90 and incubated at 37 ° C for 18 to 24 hours [10]. The observation of an inhibition zone
91 reflected the existence of antimicrobial activity. Observation of an inhibition zone can be used
92 to judge the efficiency of substances in extract or fractions. Control tests were carried out
93 using discs impregnated with 40µL of appropriate solvent used to prepare extract or fractions.

94 To confirm the resistance of bacteria, tests on young colonies using oxacillin (OX-5 μ g) and
95 cefoxitin (FOX-30 μ g) for *S. aureus* and the ceftazidime (CAZ-30 μ g) and imipenem (IMP 10
96 μ g) for *P. aeruginosa* were made under the same conditions.

97 **2.2.2. Antimicrobial screening**

98 A concentration range of plant extract was prepared by the method of double dilution with
99 concentrations ranging from 100.00 to 0.39 mg / mL for ethyl acetate extract and from 50.00
100 to 0.19 mg/mL for its fractions.

101 The antimicrobial screening was performed using the method proposed by Oussou [13] and
102 Golly [12]. The tests were performed by introducing into a series of hemolysis tubes 1 mL of
103 the solution of plant extract and 1 mL of bacterial inoculums as described by Moroh [14]. At
104 the same time, in control tube, 1 mL of the solvent used to solve the extract (DMSO / distilled
105 water to 1: 13 v / v) and 1 mL of bacterial inoculums were introduced. All the tubes were
106 incubated at 37°C for 18 to 24 hours.

107 The results of antimicrobial screening were read looking through at daylight using human eye
108 [15, 16]. The transparency of the tubes indicated the antimicrobial effect of the tested extract,
109 while its turbidity shows its ineffectiveness (a sign of bacterial growth). The Minimum
110 Inhibitory Concentration (MIC) will correspond to the concentration of the extract in the first
111 tube with a clear content.

112 The minimum bactericidal concentration (MBC) is the lowest concentration of extract that
113 kills at least 99.99% of bacteria in culture. For its determination, the content of control tube
114 was diluted to 10^{-4} , corresponding to 0.01% of survival bacteria in culture. The experimental
115 tubes sowed antimicrobial effect from the CMI's one are transplanted by streaks of 5cm on
116 Mueller Hinton agar and incubated at 37°C for 24 hours. The first experimental tube in which
117 the number of determined germs is less or equal to the dilution concentration (10^{-4})
118 corresponds to the CMB.

119 **2.3. Phytochemical screening**

120 The phytochemical study of the barks of *Ceiba pentandra*, based on color and / or
121 precipitation tests was carried out on the powder of crushed bark, ethyl acetate extract and the
122 active fraction [17, 18]. The target molecules groups of this screening were saponins, tannins,
123 flavonoids, alkaloids, steroids, terpenoids and cardiac glycosides

124 **3. RESULTS**

125 The tests of efficiency conducted prior to the determination of microbiological parameters of
126 extract at 200 mg/mL give the results summarized in Table 1. The results of the effectiveness
127 tests confirm the resistant and sensitive characters received germs. Moreover, these tests
128 indicate that the extract of the bark of *Ceiba pentandra*, and its fractions exert antibacterial
129 actions on all *S. aureus strains*. According to the inhibitions diameters, fraction F8 appears
130 more effective than the crude extract, fractions F6 and F7 but still less effective than
131 antibiotics. However, no significant action is observed on *P. aeruginosa strains*.

132 These results highlight the presence in the crude extract and its fractions F6, F7 and F8 of
133 bioactive substances which are able to inhibit the growth of *S. aureus* at 200 mg / mL in a
134 solid medium.

135 Even if the efficacy tests showed a biological activity at the concentration of 200 mg / mL of
136 crude extracts and fractions only on *S. aureus*, antibacterial parameters were investigated
137 using all studied germs. However we limited the tests with AcOEt extract and fraction F8.

138 The results of this determination are summarized in Tables 2. This Table shows different
139 results from Table 1. In liquid medium crude ethyl acetate extract and fraction F8 have
140 antibacterial activity on all studied germs. This results are conformed to those we have
141 observed while testing leaves extracts of *Vernonia colorata* on the same strains [12]. The
142 results of this table shows that the purification of total extract does not affect positively the
143 antibacterial activity. Indeed, depending on the bacterial strain, the AcOEt extract exhibits
144 MIC values ranging from 0.78 to 6.25 mg / mL and those of CMB vary from 1.04 to 8.33 mg
145 / mL. The most active fraction (F8) after purification has a substantially similar activity: CMI
146 ranging between 0.52 to 6.25 mg / mL and CMB from 1.04 to 10.42 mg / mL.

147

148 **Table 1 Results of effectiveness tests of the extract, fractions and antibiotics**

Bacterial strains	Tested substances and observed inhibition diameter (mm)													
	Extrait AcOEt	F1	F2	F3	F4	F5	F6	F7	F8	F9	OX (5µg)	FOX (30µg)	CAZ (30µg)	IPM (10µg)
<i>S. aureus Méti S</i>	13,00±1,00	0	0	0	0	0	10,00±1,00	10,67 ±0,58	14,33 ±0,58	0	43	32	-	-
<i>S. aureus Méti R</i>	12,33±0,58	0	0	0	0	0	10,33±0,58	0,00 ±0,00	13,67 ±0,58	0	0	0	-	-
<i>S. aureus ATCC 25923</i>	12,00±1,00	0	0	0	0	0	11,00 ±00	0,67± 0,58	16,00 ±1,00	0	28	30	-	-
<i>P. aeruginosa Cefta S/ImpS</i>	0	0	0	0	0	0	0	0	0	0	-	-	30	30
<i>P. aeruginosa Cefta R/ImpR</i>	0	0	0	0	0	0	0	0	0	0	-	-	0	0
<i>P. aeruginosa ATCC 27853</i>	0	0	0	0	0	0	0	0	0	0	-	-	26	25

150 *Not tested (-); Oxacillin (Ox); Cefoxitin (Fox); Ceftazidime (Caz); Imipenem (Imp).*

151

152 These results could be due to a specific nature, concentration of the actives ingredients, modes
 153 of action and the test mode [19]. In addition, the antibacterial activity observed in the liquid
 154 medium seems to indicate a poor dissemination of the extract in solid medium [20].
 155 The ratio CMB / CMI of all substances is less than or equal to two (≤ 2). According
 156 Marmonier [21], such of ratio indicate that the substances are bactericide. We can therefore
 157 conclude that bark extracts of *Ceiba pentandra* have bactericidal activity on all studied germs.
 158

159 Table 2: Antimicrobial parameters of AcOEt extract and fraction F8

Bacterial strains	Crude ethyl acetat extract (AcOEt)			Fraction F8		
	CMI (mg/mL)	CMB (mg/mL)	CMB/CMI	CMI (mg/mL)	CMB (mg/mL)	CMB/CMI
<i>S.aureus Méti S</i>	1,04±0,45	1,04±0,45	1	1,04 ± 0,45	1,56 ± 00	2
<i>S.a ureus Méti R</i>	0,78±00	1,56±00	2	0,78 ± 00	1,56 ± 00	2
<i>S.aureus ATCC 25923</i>	1,04±0,45	1,04±0,45	1	0,52 ± 0,23	1,04 ± 0,45	2
<i>P.aeruginosa Cefta S</i>	6,25±00	8,33±3,61	1	6,25 ± 00	10,42 ± 3,61	2
<i>P. aeruginosa Cefta R</i>	6,25±00	8,33±3,61	1	6,25 ± 00	10,42 ± 3,61	2
<i>P. aeruginosa ATCC 27853</i>	6,25±00	8,33±3,61	1	6,25 ± 00	10,42 ± 3,61	2

160 To establish the molecular family of active substance against bacterial strains phytochemical
 161 screening was undertaken. The results of these analyzes of the crushed dried bark, AcOEt
 162 extract and fraction F8 are summarized in Table 3. From this table it is apparent that the bark
 163 of this plant contain alkaloids, tannins, flavonoids, saponins, steroids and terpenoids and
 164 cardiac glycosides. These results are almost consistent with those reported by Adebayo-Tayo
 165 [9]. However, no tannin were found in Nigeria's *Ceiba pentandra*. The presence of these
 166 secondary metabolites in bark of *Ceiba pentandra* could justify its use in relieving ailments
 167 such as fevers, abscesses, paronychia, mental illness, conjunctivitis, vertigo, headaches,
 168 wounds and other skin diseases [8, 9].

169 If the ethyl acetate extract contains flavonoids, tannins, terpenoids and steroids,
 170 the most active fraction (F8) contains only flavonoids, tannins, steroids and terpenes.
 171 Therefore, we may apply the antimicrobial activity on the microorganisms studied flavonoids,

172 tannins and terpenes and steroids. That support that the active substance is quite flavonoid,
 173 terpene or annin
 174

175 Table 3: Phytochemical screening of crushed bark, extract and active fraction of *Ceiba*
 176 *pentandra*

Groupes chimiques	Crushed powder	(AcOEt) extract	Fraction F8
Alcaloïdes	+	-	-
Flavonoïdes	+	+	+
Tanins	+	+	+
Saponines	+	-	-
Stéroïdes et Terpénoïdes	+	+	+
Glycosides cardiaques	+	-	-

177 *Presence (+); Absence (-)*

178

179 4. CONCLUSION

180 This microbiological and phytochemical study revealed that the bark of *Ceiba pentandra* of
 181 Côte d'Ivoire have a slightly different composition secondary metabolites that of the Nigerian
 182 plant.

183 Ratio values of CMB / CMI indicate that the ethyl acetate extract and its fraction F8 have
 184 bactericidal properties on the studies strains. This activity could be attributed to flavonoids,
 185 tannins, terpenes or steroids. In future, we plan to identify the actives components of this
 186 plant.

187

188 REFERENCENCESLITERATURE

- 189 1. Organisation Mondiale de la Santé (OMS). 2002. Stratégie de l'OMS pour la médecine
 190 traditionnelle pour 2002-2005. WHO/EDM/TRM/2002, Genève, 65 p.
- 191 2. Akoua Koffi C., Guessennd N., Gbonon V., Faye-Kette A. Y. H., Dosso M. 2004.
 192 Methicillinresistant of *staphylococcus aureus* in Abidjan (1998-2001): A new hospital
 193 problem. *Medicines et maladies infectieuses* 34 (3) pp. 132-136.

- 194 3. Kacou-N'Douba A., Bouzid S. A., Guessenned K. N., Kouassi M., Bengue A. A., Faye-
195 Kette A. Y. H., Dosso M. 2001. Antimicrobial resistance of nasopharyngeal isolates of
196 *Streptococcus pneumoniae* in health carriers/ report of a study in 5-year-olds in marcorey;
197 Abidjan Côte d'Ivoire. *Annals of Tropical Paediatrics: International child health*. 21 (2)
198 pp. 149-154.
- 199 4. Benbachir M., Benredjeb S., Boye C. S., Dosso M., Belabbes H., Kamoun A., Kane O.,
200 Elmdaghri N. 2001. Two-year surveillance of antibiotic resistance in *streptococcus*
201 *pneumonia* in four African cities. *Antimicrobial Agents and Chemotherapy* 45 (2) pp. 627-
202 629.
- 203 5. Akinyemi K. O., Oladapo O., Okwara C. E., Ibe C. C., Fasure K. A. 2005. Screening of
204 crude extracts of six medicinal plants used in South-West Nigerian unorthodox medicine
205 for antimethicillin resistant *Staphylococcus aureus* activity. *BMC Complementary and*
206 *Alternative Medecine*, 5 pp. 1-6.
- 207 6. Mirabaud, M.I., (2003).Entérobactéries à BLES en pédiatrie en 1996. Thèse de doctorat
208 d'Etat en Médecine n°10303. Université de Genève (Suisse) 96p.
- 209 7. Aké, A. (1991).Rapport du colloque international sur la médecine traditionnelle et
210 pharmacopée africaine à Abidjan, Côte d'Ivoire. Bull. Pharm. Afri.4. Paris n°2. 203p.
- 211 8. Arbonnier M. 2000. Arbres, arbustes et lianes des zones sèches d'Afrique de l'Ouest.
212 CIRAD-MNHN-UICN, France, 541 p.
- 213 9. Adebayo-Tayo BC, Adegoke AA, Okoh AI, Ajibesin KK. 2010.Rationalizing some
214 medicinal plants used in treatment of skin diseases. *African Journal of Microbiology*
215 *Research_Vol. 4(10)*, pp. 958-963.
- 216 10. Zakaria A. Z., Zaiton H., Henie P. F. T., Mat Jais M. A., Kasthuri D., Thenamutha M.,
217 Othman W. F., Nazaratulmawarina R., Fatimah A. C. The *in vitro* Antibacterial Activity
218 of *Corchorus olitorius* and *Muntingia calabura* Extracts. *Journal of pharmacology and*
219 *toxicology*, (2006 a), 1(2): 108-114.
- 220 11. SoroD.,Kone M. W.,Kamanzi K. ;Evaluation des activités Antimicrobiennes et Anti-
221 Radicaux Libres de Quelques Taxons Bioactifs de Côte D'Ivoire ; *European Journal of*
222 *Scientific Research* 1450-216X Vol.40 No.2 (2010), pp.307-317
- 223 12. Golly K. J., Siaka S., Guessennnd N., Soro Y., Djama A. J., Dosso M.Phytochemical
224 assessment and antimicrobial activity of leaves extract of *Vernonia colorata* (Wild.) Drake
225 on Resistant Germs of *Staphylococcus aureus* and *Pseudomonas aeruginosa*. *Journal of*
226 *Chemical and Pharmaceutical Research*, 2012, 4(5):2490-2494.

- 227 13. Oussou K. R., Yolou S., Boti J. B., Kouadio Guessennd N., Kanko C., Ahibo C. et
228 Casanova J. ; Etude Chimique et Activité Antidiarrheique des Huiles essentielles de deux
229 Plantes aromatiques de la pharmacopée ivoirienne; *European Journal of Scientific*
230 *Research*1450-216X Vol.24 No.1 (2008), pp.94-103.
- 231 14. Moroh J. L. A., Bahi C., Dje K., Loukou Y. G., Guédé-Guina F. (2008). Étude de l'activité
232 antibactérienne de l'extrait acétatique (EAC) de *Morinda morindoides* (Baker) milne-
233 redheat (rubiaceae) sur la croissance *in-vitro* des souches *d'Escherichia coli*. *Bull Soc R*
234 *Sci Liege*, **77**: 44-61.
- 235 15. Oussou K. R.; C. Kanko; N. Guessend; S. Yolou; G. Koukoua; M. Dosso; Y. T.
236 N'guessan; G. Figueredo; Jean-Claude CHALCHAT, 2004; « Activités antibactériennes
237 des huiles essentielles de trois plantes de Côte d'Ivoire », C.R. Chimie 7) pp1081-1086.
- 238 16. Koné, W. M., Kamanzi Atindehou, K., Kacou-n'douba, A., dosso, M., (2007). «Evaluation
239 of 17 medicinal plants from Northern Côte d'Ivoire for their in vitro activity against
240 *Streptococcus pneumoniae*». *African Journal of Traditional, Complementary and*
241 *Alternative Medicines* 4 (1):17-22
- 242 17. Oyetayo F. L., Oyetayo V O., Ajewole V. Phytochemical profile and antibacterial
243 properties of the seed and leaf of the Luffa plant (*Luffa cylindrical*). *Journal of*
244 *pharmacology and toxicology.*, 2007, 2(6): 586-589.
- 245 18. Reza H. S. M., Mandal C., Alam A. K., Salam A., Rahman A. M., Amin R. M., Huda N.
246 M., Ghosh C. N., Ali R. M., Ahmed F. Phytochemical, antibacterial and antinociceptive
247 studies of *Hoya parasitica*. *Journal of pharmacology and toxicology*, (2007), 2(8): 753-
248 756.
- 249 19. Thagara J. H. S., Adjei O., Allen B. W., Portaels F. *et al.* (2000) *In vitro* activity of
250 ciprofloxacin, Sparfloxacin, Ofloxacin, Amikacin and Rifampicin against Ghanian isolates
251 of *Mycobacterium ulcerans* ; *J. Antimicrob. Agents Chemother*, 45(2), 2000, 231-233.
- 252 20. Zakaria A. Z., Mat Jais M. A., Sulaiman R. M., Mohamed Isa P. S. S., Riffin S. The *in*
253 *vitro* Antibacterial Activity of Methanol and Ethanol Extracts of *Carica papaya* flowers
254 and *Mangifera indica* Leaves. *Journal of pharmacology and toxicology*, (2006 b), 1(3):
255 278-283.
- 256 21. Marmonier A. A. (1990)Introduction aux techniques d'étude des antibiotiques.
257 Bactériologie Médicale, technique usuelles, 227-236.