

Original Research Article

Anti-venom Activity of *Mucuna pruriens* Leaves Extract Against

ABSTRACT

Aim: The study was done to investigate the anti-venom activity of *Mucuna pruriens* leaves extract against cobra snake (*Naja Hannah*) venom.

Study Design: The mice were randomly grouped into six groups (A, B, C, D, E, and F) of five rats each. Group A served as the normal control (no induction), and the mice in the group were given normal saline (1ml/kg/body weight). Group B served as the test control (snake venom was induced but no treatment administered), Group C served as the standard control (snake venom was induced and treated with antivenin, a standard drug), Group D, E and F were all induced with the cobra snake venom and treated with ethanolic extracts of the leaves of *Mucuna pruriens* for 14 days.

Methodology: The induction with cobra snake venom was done with 0.075mg/kg b.w of venom and thereafter the treatment with *Mucuna pruriens* extract for Group D, E and F were done with 40 mg/ kg, 60 mg/ kg and 80 mg/ kg respectively intraperitoneally in the mice. Serum blood of the animals was used to assay for total cholesterol, bilirubin, AST, ALT, GSH and catalase levels after 14 days.

Result: The injection of crude venom of cobra snake (*Naja hannah*) caused an increase in cholesterol, AST, ALT, bilirubin, catalase and glutathione in envenomated mice which significantly reduced ($p < 0.05$) compared to all the controls after 14 days of treatment with the extract.

Conclusion: The results suggest that 80 mg/ kg of the plant extract is more effective than the standard drug, therefore *Mucuna pruriens* leaves have a greater anti-venom potential for curing snake bite, than antivenin.

27 **Keywords:** Anti-venom, *Mucuna pruriens*, Antivenin, Cobra snake, Haemorrhage

28

29 1.0 INTRODUCTION

30 Each year in the world a lot of people receive venomous bites by snake and about 40,
31 000 of them die (Chippaux, 1998). *Echis carinatus* and *Naja hannah* are the major
32 causes of snakebite deaths in Plateau State, Nigeria. Snake venoms of Viperidae and
33 Elapidae are known to consist of a complex mixtures of toxins and enzymes which are
34 responsible for haemorrhage, myonecrosis, neurotoxicity and alteration of blood
35 coagulation (Markland, 1998; Warrell, 1989).

36 The only effective treatment of the modern medicine for serious snakebites is the use of
37 the antidote (antivenin), derived from antibodies, produced in horse's blood serum after
38 injecting the animal with snake venom. In humans, antivenin is administered either
39 through the veins or injected into muscle and acts by neutralizing the snake venom
40 which has entered the body. Because antivenin is obtained from horses, snakebite
41 victims sensitive to horse products must be very carefully treated. The danger is that
42 they could develop an adverse reaction or even an anaphylactic shock. Moreover the
43 efficacy of the serums is dependent on the rapidity with which the specificity treatment is
44 started and it is only specific for the venom used for the immunization. These represent
45 an important limitation of this kind of therapy.

46 The genus *Mucuna*, belonging to the Fabaceae family, sub-family Papilionaceae,
47 includes approximately 150 species of annual and perennial legumes. Among the
48 various under-utilized wild legumes, the velvet bean *Mucuna pruriens* ("Cowitch" and

“cowhage” are the common English names) is widespread in tropical and sub-tropical regions of the world. It is considered a viable source of dietary proteins (Janardhanan *et al.*, 2003; Pugalenti *et al.*, 2005) due to its high protein concentration (23–35%) in addition its digestibility, which is comparable to that of other pulses such as soybean, rice bean, and lima bean (Gurumoorthi *et al.*, 2003). It is therefore regarded as a good source of food.

Mucuna prurien is a popular Indian medicinal plant, which has long been used in traditional Ayurvedic Indian medicine, for diseases including Parkinsonism (Sathyanarayanan *et al.*, 2007). The beans have also been employed as a powerful aphrodisiac (Amin, 1996) and have been used to treat nervous disorders and arthritis (Jeyaweera, 1981). The bean, if applied as a paste on scorpion stings, is thought to absorb the poison (Jeyaweera, 1981). The non-protein amino acid-derived L-dopa (3, 4-dihydroxy phenylalanine) found in this underutilized legume leaves resists attack from insects, and thus controls biological infestation during storage. According to D'Mello (1995), all anti-nutritional compounds confer insect and disease resistance to plants. Further, L-dopa has been extracted from the leaves to provide commercial drugs for the treatment of Parkinson's disease. L-Dopa is a potent neurotransmitter precursor that is believed, in part, to be responsible for the toxicity of the *Mucuna* leaves (Lorenzetti *et al.*, 1998). Antiepileptic and anti-neoplastic activity of ethanolic extract from *Mucuna prurien* had been reported (Gupta *et al.*, 1997).

An ethanolic extract of *Mucuna prurien* leaves has demonstrated significant in vitro anti-oxidant activity, and there are also indications that ethanolic extracts of *Mucuna puriens* may be a potential source of natural anti-oxidants and anti-microbial agents (Rajeshwar

et al., 2005). All parts of *Mucuna pruriens* possess valuable medicinal properties and it has been investigated in various contexts, for its anti-diabetic, aphrodisiac, anti-neoplastic, anti-epileptic, and anti-microbial activities (Sathyanarayanan et al., 2007). Its anti-venom activities have been investigated by (Guerranti et al. (2002) and its anti-helminthic activity has been demonstrated by Jalalpure (2007). *Mucuna pruriens* has also been shown to be neuroprotective (Misra and Wagner, 2007), and has demonstrated analgesic and anti-inflammatory activity (Hishika et al., 1981). The primary objective of the present study is to confirm the bioactivity of *Mucuna pruriens* leaves extract against cobra snake (*Naja Hannah*) venom.

81

82 **2.0 METHODOLOGY**

83 **2.1 Collection and Preparation of the Plant Sample**

84 The fresh leaves of *Mucuna pruriens* were collected from Kogi State, Nigeria. It was identified by Dr. S. M. Ayodele (Dept. of Botany, Kogi State University). The leaves were dried under room temperature for 5 days. The dried sample was ground powdery form, using the warring commercial blender.

88 **2.2 Extraction of *Mucuna pruriens* leaves**

89 The plant material (500g) was defatted with 400ml hexane (C₆ H₁₄) by using a Soxhlet apparatus for 5h. The defatted powder plant material was air-dried. The air-dried defatted powdered plant material was then extracted with 400 ml ethanol (C₂H₅OH) by using a Soxhlet apparatus for 8h. The residue was dried over night and extracted with

250 ml water (H₂O) by using a shaking water bath at 70⁰C for 2h. The extraction with water was repeated three time. The water filtrates were mixed together. The ethanol and water extract were filtered and evaporated by using a rotary evaporator and freeze dryer to give the crude-dried extract. The dried extracts were stored at -20⁰C until used.

Calculation:

$$\text{Percentage yield (\%)} = \frac{\text{weight of the plant extract}}{\text{Weight of the dried plant used}} \times 100$$

2.3 Proximate Analysis af *Mucuna prurien*

The moisture content, ash content, carbohydrate content, crude fibre and crude protein were determined using methods as described by AOAC (1990).

2.4 Phytochemical Screening

The ethanolic extract of *Mucuna prurien* leaves were screened for the presence of phytochemical compound as described by Treatise and Evans (1989) and Sofowora (1993).

2.5 Animals

Experimental mice were purchased from the animal house of NIPRID. The animals were housed in steel cages and kept at room temperature. The mice had no history of drug consumption that is; they had not been used for any investigation. The mice were put on standard mice pellet (feed) and pure drinking water and allowed to get acclimatized for 21 days before the start of the experiment.

111

112

113 2.6 Induction of Anti-Snake Venom

114 The cobra snake venom native preparations, from *Mucuna pruriens*, were given
 115 intraperitoneally (i.p) to the mice at a dose which were proportional to the weight of the
 116 animals. The volumes of preparation were identical and the same amounts were
 117 injected.

118 2.7 Experimental Design

119 White male albino mice of wister strain of body weight ranging between 15-30g were
 120 used for the research study. The mice were randomly grouped into six groups (A, B, C,
 121 D, E, and F) of five rats each. Group A served as the normal control (no induction), and
 122 the mice in the group were given normal saline (1ml/kg/body weight). Group B served as
 123 the test control (snake venom was induced but no treatment administered), Group C
 124 served as the standard control (snake venom was induced and treated with antivenin, a
 125 standard drug), Group D, E and F were all induced with the cobra snake venom and
 126 treated with ethanolic extracts of the leaves of *Mucuna pruriens* (Table 1).

127 **Table 1: Experimental Design**

Group	Name	Treatment for 14 days
A	Normal Control	No induction, No treatment
B	Test Control	Induced with 0.075mg/kg b.w of venom, but no treatment
C	Standard Control	Induced with 0.075mg/kg b.w of venom and treated with antivenin (a standard drug).
D	Group A	Induced with 0.075mg/kg b.w of venom and treated with 40mg/kg extract
E	Group B	Induced with 0.075mg/kg b.w of venom and treated with 60mg/kg extract

F	Group C	Induced with 0.075mg/kg b.w of venom and treated with 80mg/kg extract
---	---------	---

128

129 **2.8 Determination of Serum Cholesterol Level**

130 Total cholesterol was determined by the enzymatic endpoint method as described by
 131 Trinder (1969). In this method, cholesterol was determined after enzymatic hydrolysis
 132 and oxidation in a series of reaction. The indicator quinoneimine used in this method
 133 was formed from hydrogen peroxide and 4-aminoantipyrine in the presence of phenol
 134 and peroxidase.

135 **2.9 Determination of Serum Bilirubin Level (Vander Bergh's Reaction)**

136 The total bilirubin in serum or plasma is determined using the method of Jendrassik and
 137 Gróf (1938) by coupling with diazotized sulfanilic acid after the addition of caffeine,
 138 sodium benzoate and sodium acetate. A blue azobilirubin is formed in alkaline Fehling
 139 solution II. This blue compound can also be determined selectively in the presence of
 140 yellow by products (green mixed coloration) by photometry at 578 nm.

141

142 **2.10 Determination of Transaminases (ALT and AST)**

143 Alanine Aminotransferase was determined in a method as described by Reitman and
 144 Frankel (1957). In this method, ALT was measured by monitoring the concentration of
 145 pyruvate hydrazone formed with 2,4-dinitrophenylhydrazine in a reaction. Aspartate
 146 Aminotransferase was determined in a method as described by Reitman and Frankel

147 (1957). In this method, AST is measured by monitoring the concentration of
148 oxaloacetate hydrazone formed with 2,4-dinitrophenylhydrazine in a reaction.

149

150 **2.11 Determination of Serum Glutathione level**

151 The principle was based on the reduction of 5,5 dithiobis (2-nitrobenzoic acid) (DTNB)
152 with reduced glutathione (GSH) to produce a yellow compound. The reduced
153 chromogen was directly proportional to GSH concentration and its absorbance was
154 measured at 405 nm (Tietz, 1990).

155 **2.12 Determination of Serum Catalase Level**

156 Catalase was determined by the method described by Cohen *et al.* (1970). In this
157 method, catalase catalysed the conversion of hydrogen peroxide to oxygen and water in
158 a reaction.

159 **2.13 Statistical analysis**

160 Values are expressed as mean + S.E.M randomized complete block design analysis of
161 variance was used for statistical analysis. P values less than 0.05 was considered
162 significant.

163

164 **3.0 RESULTS**

165 **3.1 Percentage Yield of Extract**

166 The extract was thick and greenish in colour, with an ethanolic extraction of
167 *Mucuna pruriens* leaves which indicated the yield of 6.73%.

168 3.2 Phytochemical Screening

169 The preliminary phytochemicals test reveals that the major phytochemical constituents
170 in *Mucuna pruriens* leaves are alkaloids, flavonoids, tannins, saponins, steroids,
171 terpenoids, cardiac glycosides and anthraquinones (Table 2).

172

173 Table 2: Phytochemical composition of *Mucuna pruriens* leaves

Phytochemicals	Presence in <i>Mucuna pruriens</i> leaves
Alkaloids	++
Flavonoids	++
Tannins	+
Saponins	++
Steroids	++
Terpenoids	++
Cardiac glycosides	+
Anthraquinones	+

174

175 3.3 Proximate Analysis

176 The proximate analysis of the leaves in Table 3 has a moisture content of 11.37%,
177 crude fiber 31.91%, crude fat 2.97%, carbohydrate 45.65%, Ash content 3.00%.

178

179 Table 3: Proximate composition of *Mucuna pruriens* leaves

PARAMETER	COMPOSITION
Moisture content	11.37 %
Crude protein	31.91%
Crude fat	2.97 %
Carbohydrate	45.65 %

Ash	3.00 %
-----	--------

3.4 Effect of Venom Induction and Extract on Biochemical Parameters

The results revealed in Table 4a and 4b that, the injection of crude venom of cobra snake (*Naja hannah*) caused an increase in cholesterol, AST, ALT, bilirubin, catalase and glutathione in envenomated mice compared to the normal control mice. There was no significant difference in the levels of cholesterol, AST, ALT, bilirubin, catalase and glutathione of the treated groups when compared to the test control group.

Table 4a: Result obtained on the effect of extract on Cholesterol, AST and ALT at day one after venom induction

Treatment	Cholesterol (mg/dl)	AST (U/l)	ALT (U/l)
Normal control	55.94 ± 0.07	28.93 ± 0.14	22.60 ± 0.49
Test control	127.41 ± 0.13 ^a	74.34 ± 2.24 ^a	36.64 ± 0.64 ^a
Standard control	125.91 ± 0.06 ^a	74.96 ± 0.18 ^a	34.96 ± 0.35 ^a
Group A 40mg/kg	127.48 ± 0.05 ^a	75.67 ± 0.21 ^a	36.20 ± 0.42 ^a
Group B 60mg/kg	128.30 ± 0.04 ^a	73.61 ± 0.14 ^a	37.10 ± 0.21 ^a
Group C 80mg/kg	128.11 ± 0.03 ^a	73.93 ± 0.07 ^a	37.04 ± 0.14 ^a

Values are expressed as means ± SEM. ^a indicate values that are significantly different when compared to the normal control at (p < 0.05).

Table 4b: Result obtained on the effect of extract on Bilirubin, Catalase and Glutathione at day one after venom induction

Treatment	Bilirubin (mg/dl)	Catalase (mg/dl)	Glutathione (U/L)
Normal control	0.42 ± 0.04	17.19 ± 0.05	4.69 ± 0.51
Test control	0.95 ± 0.07 ^a	25.45 ± 0.93 ^a	8.47 ± 1.21 ^a
Standard control	0.90 ± 0.02 ^a	26.06 ± 0.06 ^a	8.21 ± 0.68 ^a
Group A 40mg/kg	0.94 ± 0.05 ^a	25.82 ± 0.74 ^a	8.37 ± 0.74 ^a
Group B 60mg/kg	0.91 ± 0.03 ^a	27.48 ± 0.62 ^a	7.95 ± 0.55 ^a
Group C 80mg/kg	0.95 ± 0.02 ^a	25.59 ± 0.50 ^a	8.11 ± 0.48 ^a

Values are expressed as means ± SEM. ^a indicate values that are significantly different when compared to the normal control at (p < 0.05).

The levels of cholesterol, AST, ALT, bilirubin, catalase and glutathione were significantly reduced when compared to the test control after 14 days of treatment with the extract (Table 5a and 5b).

Table 5a: Result obtained on the effect of extract on Cholesterol, AST and ALT at day fourteen

Treatment	Cholesterol (mg/dl)	AST (U/l)	ALT (U/l)
Normal Control	50.00 ± 0.07	25.54 ± 0.07	17.46 ± 0.20
Test control	113.45 ± 0.17	62.25 ± 1.24	38.88 ± 0.44
Standard control	60.00 ± 0.06 ^{ab}	30.37 ± 0.19 ^{ab}	19.71 ± 0.28 ^{ab}
Group A 40mg/kg	96.67 ± 0.05 ^{abc}	40.45 ± 0.21 ^{abc}	26.24 ± 0.35 ^{abc}
Group B 60mg/kg	83.33 ± 0.04 ^{abc}	36.39 ± 0.14 ^{abc}	21.46 ± 0.21 ^{ab}
Group C 80mg/kg	55.67 ± 0.03 ^{abc}	28.82 ± 0.09 ^{abc}	18.41 ± 0.17 ^b

Values are expressed as means \pm SEM. ^a indicate values that are significantly different when compared to the normal control at ($p < 0.05$), ^b indicate values that are significantly different when compared to the test control at ($p < 0.05$) and ^c indicate values that are significantly different when compared to the standard control at ($p < 0.05$).

Table 5b: Result obtained on the effect of extract on Bilirubin, Catalase and Glutathione at day fourteen

Treatment	Bilirubin (mg/dl)	Catalase (mg/dl)	Glutathione (U/L)
Normal Control	0.34 \pm 0.02	14.16 \pm 0.32	2.59 \pm 0.31
Test control	0.98 \pm 0.07	20.45 \pm 1.30	6.47 \pm 1.21
Standard control	0.47 \pm 0.03 ^{ab}	13.06 \pm 0.56 ^b	1.83 \pm 0.33 ^{ab}
Group A 40mg/kg	0.76 \pm 0.04 ^{abc}	16.67 \pm 0.95 ^{abc}	4.32 \pm 0.49 ^{abc}
Group B 60mg/kg	0.54 \pm 0.03 ^{abc}	15.89 \pm 0.84 ^{bc}	3.97 \pm 0.22 ^{abc}
Group C 80mg/kg	0.41 \pm 0.01 ^{abc}	12.95 \pm 0.72 ^{abc}	1.82 \pm 0.12 ^{ab}

Values are expressed as means \pm SEM. ^a indicate values that are significantly different when compared to the normal control at ($p < 0.05$), ^b indicate values that are significantly different when compared to the test control at ($p < 0.05$) and ^c indicate values that are significantly different when compared to the standard control at ($p < 0.05$).

4.0 DISCUSSION

Nevertheless, in *in vivo* studies tannins may interact with plasma proteins from the blood circulation. The anti-venom activity observed in the mice treated with the extract may be attributed to the presence of any of these compounds alkanoids, tannins, flavonoids, steroids and terpenoid (Rajendran *et. al*, 2010). The proximate analysis of the leaves in Table 3 has a moisture content of 11.37%, crude fiber 31.91%, crude fat 2.97%, carbohydrate 45.65%, Ash content 3.00%. The Ash content is known to enhance digestibility, slow down the release of glucose into the blood stream and reduces blood cholesterol level.

The present study revealed in Table 4a and 4b that, the injection of crude venom of cobra snake (*Naja hannah*) caused an increase in cholesterol, AST, ALT, bilirubin, catalase and glutathione in envenomated mice which significantly reduced after 14 days of treatment with the extract as shown in Table 5a and 5b. These findings are in agreement with other investigators who reported that the reduction in cholesterol, AST, ALT, bilirubin and catalase in envenomated mice was observed in laboratory animals treated with the extracts of *mucuna pruriens* leaves Abdul-Nabi *et al.* (1997). It might be assumed that, the increased levels of these serum constituents could be due to disturbance in renal functions as well as haemorrhages in some internal organs when challenged with a snake venom. In addition, the increasing in vascular permeability and haemorrhages in vital organs due to the toxic action of various snake venoms were described by (Meier and Stocker 1999; Meier and Theakston; 1986). Also, the reduction in serum cholesterol, AST, ALT, catalase, glutathione albumin and total bilirubin levels in the envenomated mice could be attributed to the anti-venom potentials of the extract of *mucuna pruriens* administered.

Furthermore, acute renal damage together with glomerular, tubular and vascular lesions following various snake bites have been reported (Sitprija *et al.*, 1982; Sani and Purandare, 1972; Aung-Khin, 1978) with additional, increased vascular permeability and hemorrhages in various vital organs. Another factor is the increase vascular permeability due to toxic action of the venom which could contribute to the low level of protein from plasma and tissue (Olajide *et al.*, 1999).

In Table 4a, elevation of ALT and AST in the mice administered with venom as observed in the serum have serious implication on health of the animals. Such elevations are found in cases of both liver damage and myocardial infarction (Gray and Howorth, 1982). The elevation of AST and ALT makes the liver a target of suspicion as this is usual in cases of hepatotoxicity caused by toxic agent (Rosalki, 1974). From the experiment it was observed that some of the mice died after 30 minute of induction with the snake venom except the (Normal control) and the ones treated with different dose of the venom/extract (Groups A-C) which survived till the end of the experiment. There was significant difference between the time of death in the extract treated group and those treated with venom only showing that the plant extract had effect on the activity of the venom. Thus, it was obvious that *Mucuna pruriens* leaves did show greater anti-venom activity, the extract of *Mucuna pruriens* showed a better anti-snake activity compared with antivenin. This can be attributed to the fact that tannins are able to non-specifically bind to *Naja hannah* venom proteins and precipitate them, thus provoking the anti-lethal effects.

CONCLUSION

In conclusion, this investigation revealed that the ethanolic extract of *Mucuna pruriens* leaves has the following phytochemical components of saponin, terpenoid, flavonoids, steroids, Alkaloids, Anthraquinones. It was observed that 80 mg/ kg of the plant extract is more effective than the standard drug, therefore *Mucuna pruriens* leaves has a greater medicinal plant for curing snake bite, than anti-venin.

283

284

285

286 **COMPETING INTERESTS**

287 Authors have declared that no competing interests exist.

288 **CONSENT**

289 Not applicable

290 **ETHICAL APPROVAL**

291 All authors hereby declare that all experiments have been examined and approved by
292 the appropriate ethics committee and have therefore been performed in accordance
293 with the ethical standards laid down in the 1964 Declaration of Helsinki.

294

295

296 **REFERENCES**

297 AOAC. Official Methods of analysis. 15th Edn. Association of Official Agricultural
298 Chemists, Washington, DC. 1990.

299 Abdul-Nabi IM, Raafat A, El-Shany H. Biological effect of intraperitoneal injection of rats
300 with the venom of the snake *Echis carnetus*. Egypt J. Zool. 1997;29:195-205.

- 301 Amin KMY, Khan MN, Zillur-Rehman S, Khan NA. Sexual function *Mucuna pruriens*
302 roving effect of *Mucuna pruriens* in sexually normal male rats. Fitoterapia. 1996; 67, 53-
303 56.
- 304 Aguiyi JC, Uguru MO, Johnson PB, Obi CI. Effect of *Mucuna pruriens* leaves extract on
305 smooth and skeletal muscle preparation. Fitoterapia.1997; 4: 366-70.
- 306 Aung-Khin M. Histological and ultrastructural changes of the kidney in renal failure after
307 viper envenomation. Toxicon. 1978;16: 71-75.
- 308 Chippaux JP. Snake Bite: Appraisal of the Global Situation. Bulletin WHO 1998, 76:515-
309 524.
- 310 Chippaux JP, Goyffon M. Venoms, antivenoms and immunotherapy. Toxicon;36:823-46.
- 311 Cohen, G., Dembiec, D. and Marcus, J. (1970). Measurement of catalase activity in
312 tissues extracts. Anal. Biochem. 1998;34: 30-38.
- 313 D'Mello JPF. Leguminous leaf meals in non-ruminant nutrition. In: Tropical Legumes in
314 Animal Nutrition. J.P.F. D'Mello and C. Devendra, Eds. CAB International, Wallingford,
315 UK. 1995.
- 316 Markland FS. Snake Venoms, Drugs. 1997;54 (Suppl.3):1-10.
- 317 Gray C, Howarth PJN. clinical Chemical Pathology. 9th ed. The ELBS, Edward Arnold
318 Publishers Ltd., 1982.

- 319 Gupta M, Mazumder UK, Chakraborti S, Bhattacharya S, Rath N, Bhawal SR.
320 Antiepileptic and anticancer activity of some indigenous plants. Indian J. Physiol. Allied
321 Sci. 1997;51, 53-56.
- 322 Guerranti R, Aguiyi JC, Errico E, Pagani R, Marinello E. Effects of *Mucuna pruriens*
323 extract on activation of prothrombin by Echis carinatus venom. J Ethnopharmacol.
324 2001;75, 175-180.
- 325 Gurumoorthi P, Pugalenti M, Janardhanan K. Nutritional potential of five accessions of
326 a south Indian tribal pulse *Mucuna pruriens* var. utilis; II. Investigation on total free
327 phenolics, tannins, trypsin and chymotrypsin inhibitors, phytohaemagglutinins, and in
328 vitro protein digestibility. Trop. Subtrop. Agroecosys. 2003;1,153-158.
- 329 Hishika R, Shastry S, Shinde S, Gupta SS. Preliminary, phytochemical and anti-
330 inflammatory activity of seeds of *Mucuna pruriens*. Indian J. pharmacol. 1981;13 (1), 97-
331 98.
- 332 Jalalpure SS, Alagawadi KR, Mahajanashell CS. In vitro antihelmintic property of
333 various seed oils against Pheritima posthuma. Ind. Pharm. Sci. 2007;69, 158-160.
- 334 Janardhanan K, Gurumoorthi P, Pugalenti M. Nutritional potential of five accessions of
335 a South Indian tribal pulse, *Mucuna pruriens* var. utilis. Part I. The effect of processing
336 methods on the contents of L-Dopa, phytic acid, and oligosaccharides. J Trop and
337 Subtrop Agro-ecosys. 2003;1, 141-152.
- 338 Jendrassik L, Grof P. Colorimetric Method of Determination of bilirubin. Biochem Z.
339 1938; 297:81-82

- 340 Jeyaweera, DMA. Madicinal plants used in Ceylon. Colombo, Sri Lanka; National
341 Science Council of Sri Lanka, 1981.
- 342 Lorenzetti E, MacIsaac S, Arnason JT, Awang DVC, Buckles D. The phytochemistry,
343 toxicology and food potential of velvet bean (*Mucuna adans* spp. Fabaceae). In D.
344 Buckles, O. Osiname, M. Galiba, & G. Galiano, Cover crops of West Africa; contributing
345 to sustainable agriculture. IDRC, Ottawa, Canada & IITA, Ibadan, Nigeria, 1998.
- 346 Maria DA, Vassa RC, Ruiz IRG. Haematopoietic effects induced in mice by the snake
347 venom toxin jarahagin. *Toxicon*, 1998;42: 579-585.
- 348 Markland F. Snake venoms and the hemostatic system. *Toxicon*. 1998;36, 1749–1800.
- 349 Meier J, Stocker K. Effect of venoms on homeostasis. *Toxicol*. 1999;21 (3): 1711-1820.
- 350 Meier J, Theakston RDG. Approximate LD50 determinations of snake venoms using
351 eight to ten experimental animals. *Toxicon* 1986;24, 395 – 401.
- 352 Misra L, Wagner H. Extraction of bioactive principles from *Mucuna pruriens* seeds.
353 *Indian J. Biochem. Biophys.* 2007;44, 56-60.
- 354 Olajide OA, Makinde JM, Awe SO. Effect of aqueous extract of mucuna pruriens stem
355 bark on corragenan induced oedema and granuloma tissue formation in rats and mice.
356 *J. Ethnopharmacol.* 1999;66(1): 113-177
- 357 Pugalenti M, Vadivel V, Siddhuraju P. Alternative Food/Feed Perspectives of an
358 Underutilized Legume *Mucuna pruriens* var. Utilis—A Review. *Plant Foods Hum. Nutr.*
359 2005;60: 201–218.

- 360 Rajendran K, Annie S, Maneesh MR, Vijaya B. Invitro and In vivo anti snake venom
361 (Daboia russelli) studies on various leaf extracts of *Acalpha indica* Linn. Int J Phytomed.
362 2010;2: 217-220
- 363 Rajeshwar, Y., Kumar, S.G.P., Gupta, M., Mazumder, K.U., (2005). Studies on in vitro
364 antioxidant activities of mhetanol extract of *Mucuna prurien* (Fabaceae) seeds. Euro
365 Bull Drug Res., 13, 31-39.
- 366 Reitman S, Frankel S. A colourimetric method for the determination of serum glutamate-
367 oxaloacetate and pyruvate transaminases. Am J Clin Pathol. 1957;28: 56-63.
- 368 Roldan E, Rodriguez,C, Naves G, Parra C, Brieva JA. VLA-4fibronectin interaction is
369 required for the terminal differentiation of human bone marrow capable of spontaneous
370 and high rate immunoglobulin secretion. J Exp Med. 1992;175: 1739-1747.
- 371 Rosalki SB. Enzyme profiles as indicators of susceptibility to environmental toxic
372 agents. J Proc Soc Med. 1974;67:633 – 636.
- 373 Sani SM, Purandare NM. Autopsy study of cases of snake bite with special reference to
374 renal lesions. J. Postgrad Med. 1972;18: 181-188.
- 375 Santosh R, Fattepur, Shivaji P, Gawade. Preliminary screening of herbal plant extracts
376 for antivenom activity against common sea snake (*Enhydrina schistose*) Poisoning.
377 Pharmacog, Magazing. 2004;16: 56-60.
- 378 Siddhuraju P, Becker K, Makkar HPS. Studies on the nutritional composition and
379 antinutritional factors of three different seed material of an under-utilised tropical
380 legume, *Mucuna prurien* var. utilis . J. Agric. Food Chem. 2000;48, 6048-6060.

- 381 Sitprija V, Suvanpha R, Pochanugool C, Chusil S, Tungsanga K. (1982). Acute
382 intestinal nephritis snakebite. Am J Trop Med and Hyg. 2000; 31:408-410.
- 383 Sofowora. A Medicinal Plants and Traditional Medicine in Africa. Spectrum Books Ltd,
384 Ibadan, 1993.
- 385 Sathyanarayan G, Garg PK, Prasad H, Tandon RK. Elevated level of interleukin-6
386 predicts organ failure and severe disease in patients with acute pancreatitis. J
387 Gastroenterol Hepatol. 2007;22:550-4.
- 388 Tietz, N. W. Clinical Guide to Laboratory Tests, Second Edition W. B. Saunders
389 Company, Philadephia, USA,1990.
- 390 Treatise GE, Evans WC. Pharmacognosy.11th edn. Brailliar Tiridel Can. Macmillan
391 publishers. Ltd, Ibadan, 1989.
- 392 Tripathi YB, Updhyay AK. Antioxidant property of *Mucuna prurien* Linn. Curr. Sci. 2001;
393 80, 1377-1378.
- 394 Trinder P. Determination of glucose in blood using glucose oxidase with an alternative
395 oxygen acceptor. Anal Clin Biochem. 1969;6: 24-27.
- 396 Warrell DA. Snake venoms in science and clinical medicine. Russell's viper: biology,
397 venom and treatment of bites. Trans Roy Soc Trop Med Hyg. 1989;83, 732–740.