

Role of soil nitrogen for the conifers of the boreal forest: a critical review

Lupi Carlo^{1*}, Morin Hubert¹, Deslauriers Annie¹, Rossi Sergio¹, Houle Daniel^{2,3}

¹555, boulevard de l'Université, Chicoutimi (QC), G7H 2B1, Canada, Université du Québec à Chicoutimi, Département des Sciences Fondamentales,

²Direction de la recherche forestière, Forêt Québec, Ministère des Ressources naturelles et de la Faune du Québec (QC), G1P 3W8, Canada

³Consortium sur la climatologie régionale et l'adaptation aux changements climatiques (Ouranos), Montréal (QC), H3A 1B9, Canada

ABSTRACT

Nitrogen is considered the most important element in plant nutrition and growth. However its role and availability for boreal forest conifers is still debated. Boreal conifers have adapted strategies to cope with the reduced availability of N. ECM fungi, associated with boreal conifer roots, increase soil exploration and N nutrition, especially where organic N predominates. Conifers usually take up ammonium at levels comparable to simple organic N, which probably grows in importance as organic matter accumulates with stand age, while estimates of nitrate uptake are generally lower. Conifers, especially slow growing species, may rely on internal N cycling to sustain the development of new tissues in spring. N increases photosynthesis and leaf area and thus increases growth and wood formation, leading to wider radial rings mostly because of increased earlywood production. N-depositions and disturbances (e.g. fire and harvest) may alter the soil N-cycle and affect boreal forest growth. N depositions are considered responsible for the increase in boreal forest growth during the last century. Intensive harvest and high N-depositions may shift limitation from N to another element (e.g. P, K, and B).

Climate change should affect the N cycle through complex mechanisms, including changes in the fire return interval, direct effects of warmer soils on N mineralization and stimulating plant growth modifying the balance between N stored in soils and in the living and dead (e.g. wood) biomass. Future research should try to improve our understanding of the possible outcomes of changes in disturbance regimes, N-depositions and climate, including the role of N fixation by mosses, canopy N uptake and the responses of conifers in relation to changes in microbial (symbiotic and not) communities.

Keywords ammonium; boreal conifers; growth; N limitation; nitrate; N uptake

1. OVERVIEW OF THE N CYCLE IN BOREAL FOREST SOILS

The boreal forests of the world are characterized by low evapotranspiration and decomposition, corresponding to slow nutrient cycles and accumulations of organic matter during the forest stand successions ([1],[2]). N is an essential element for plant nutrition and, together with P, is copiously required for all essential metabolic processes of the plants. The growth of the boreal forest is considered N limited, but wide ranges are generally observed in nutrient availability and interaction between elements ([3],[4], [5]), so it is important to thoroughly understand the characteristics of the N cycle in the boreal forest in order to identify the role of this fundamental element for forest productivity and C sequestration.

The major pathways of N input in forms available for plants in terrestrial ecosystems are biological N fixation (e.g. by cyanobacteria associated with mosses and lichens) and atmospheric N depositions, while N fixation through lightning is much less important (Table 1) ([6], [7], [8]). N losses may occur through leaching of dissolved N species. Nitrate is preferentially leached as compared to NH_4^+ but in unperturbed forests DON relatively gains importance over mineral N, with losses in the order of a few kg per ha per year. ([9], [10]). Losses of N also occur through volatilization (especially rapid volatilization through wildfire, while ammonium volatilization at high pH values is marginal in the typical acidic soils of the boreal forest), and denitrification (Table 1) ([11], [12]). Fires (wildfire and broadcast burning) may indeed represent an important punctual output of N (hundreds of kg N per ha) and the fire return interval is important in determining the long-term impact of N volatilization by fire on the long-term N balance of the ecosystem. Finally, N can be immobilized with polyphenols and other recalcitrant substances in microbial and plant biomass or in the soil after precipitation ([13], [14], [15], [16], [17]).

22 **Table 1 Characteristics of the soil N cycle in the boreal forest**

INPUT	Biological N fixation (BNF)	N-deposition	OUTPUT	Leaching	Volatilization	Denitrification
Characteristic	Energy intensive (symbiosis favored)	Probably low in remote and undisturbed areas		Flushes of NO_3^- associated with high water content (e.g. during snowmelt), especially in coarse-textured soils	N losses through wildfire may be locally important	Favored by limited availability of O_2 , high NO_3^- concentrations, high soil moisture, availability of soil carbohydrates, warm temperatures
	N-fixers have high requirements for P, Fe and Mo	Mainly as NH_4^+ and NO_3^- , through precipitations			Ammonia volatilization is insignificant in the boreal forest	
	Positive relation with evapotranspiration	Greater in southern boreal forest, near polluted areas		Higher potential for losses through Dissolved Organic Nitrogen (DON) with increasing organic matter accumulation	Difficult to quantify, especially for elevated punctual outputs associated with wildfires	Low in the boreal forest (since low NO_3^- concentrations)
	Favored where N supply is reduced	Probably greater in northern Europe than in Canada				Increase after freeze/thaw and wet/dry cycles
	Negative relation with N-depositions (e.g. for N-fixation in mosses) ([18])	71.8% of the boreal forest experience N_r deposition < 3 kg N ha ⁻¹ yr ⁻¹ suggesting bryophytes limiting woody plant acquisition of ambient anthropogenic N inputs through the majority of the boreal forest ([18])		Increased N losses following disturbances (insect outbreaks, harvesting and wildfires)	Fire return interval and fire severity, along with the moisture of the fuel and the type of fire (e.g. crown vs forest floor) affect N losses by volatilization and the long-term impact on the N balance of the ecosystem	

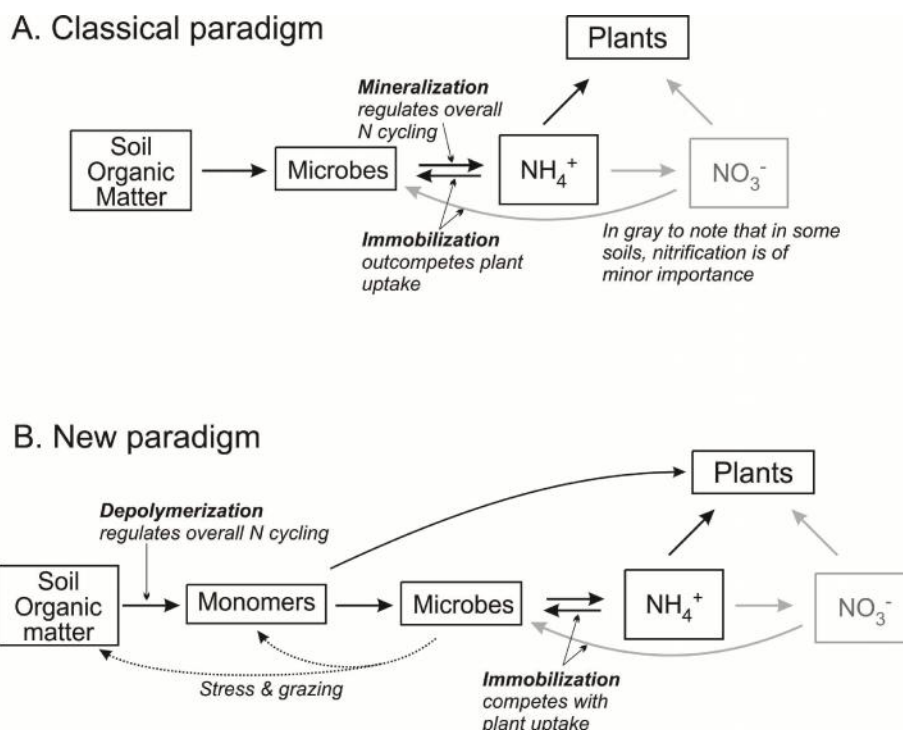
INPUT	Biological N fixation(BNF)	N-deposition	OUTPUT	Leaching	Volatilization	Denitrification
Estimate	<p>N-fixation incyanobacteria (e.g. <i>Nostoc sp.</i>) associated with mosses: 1-2 kg ha⁻¹ yr⁻¹ (comparable to low atmospheric N depositions 2-3 kg ha⁻¹ yr⁻¹)(it may be primary source of N in late-successional ecosystems;[19])</p> <p>BNF by <i>Ceanothus sp.</i> In the order of 20-100 kg N ha⁻¹ yr⁻¹ (in early to mid-successional ponderosa pine stands in the intermontane west) ([20])</p> <p>[21]reported N fixation by <i>Suillus tuberculatus</i> on lodgepole pine (<i>Pinus contorta</i>) in early-successional stands established after fire</p>	<p>Pristine watershed in eastern Canada: 1.7 kg ha⁻¹ yr⁻¹ as N-NO₃; 1.2 kg ha⁻¹ yr⁻¹ as N-NH₄; Total Dissolved Nitrogen (TDN) fluxes as deposition 3.1 kg ha⁻¹ yr⁻¹</p> <p>North-eastern North America: from <4 kg N ha⁻¹ yr⁻¹ in isolated zones to 10-12 kg N ha⁻¹ yr⁻¹ in polluted areas</p> <p>In eastern Canada (Quebec) N depositions, 2-11 kg ha⁻¹ yr⁻¹, on average 5.8 ± 1.8 kg ha⁻¹ yr⁻¹</p>		<p>Pristine watershed in eastern Canada: 0.25 kg ha⁻¹ yr⁻¹ exported via stream output N-NO₃; 0.05 kg ha⁻¹ yr⁻¹ exported via stream output N-NH₄; Total Dissolved Nitrogen (TDN) stream export 0.9 kg ha⁻¹ yr⁻¹; DON represent 67% of TDN in stream export</p>	<p>Over a fire return interval of 100-200 year, [22], estimated N losses by wildfire equal to 3-4 kg N ha⁻¹ yr⁻¹</p> <p>[23] estimated a change in the N balance associated with wildfires (not only volatilization losses) between -4 and +6 g N/m² but highly variable (standard deviation 19) with some samples showing an increase in N concentration after fire</p> <p>[24] observed effects of fire on forest floor and mineral soil N content (kg ha⁻¹) varying with the intensity of fire and significant losses at the level of the forest floor only with severe fires, with N content passing from 1364 to 764 kg N ha⁻¹ (losses amounted to only 17% of the total soil reserves)</p>	0.11 kg ha ⁻¹ yr ⁻¹ for coniferous forests (but no information for the boreal forest)
Source	[3], [6], [19], [21], [25],[26], [27], [28], [29]	[30], [31], [32], [33], [34], [35]		[36], [37], [33]	[6],[11], [12],[38], [39],[40], [41]	[42], [43], [44], [45], [46]

Even if inputs and outputs are important fluxes to and from the N cycle, they represent small amounts compared to the large pool stored in the soils of the boreal forest. [47] estimated N-accumulation in three northern Scandinavian forests stands demonstrating that, in these undisturbed forests, soils are the main reservoir of N, with humus showing contents from 3 to 24 times higher than those observed in vegetation. Rates of N accumulation in organic soil and vegetation were estimated to range between 0.30 and 0.35 g N m⁻² yr⁻¹, corresponding to about 85% of the N input to these forests. Consequently, a key element of the N cycle is soil organic matter, which is constituted by c.a. 50% of C and 5% of N, only partially (usually less than 5%) in available form ([11]).

N is cycled through plants and the relevant inputs to soils come from the turnover of below- and above-ground plant biomass, mostly leaves and fine roots. The turnover of roots, especially the fine roots and associated mycorrhizae, can return two to five times more organic matter and six times more N to the soil than the canopy litter ([48], [49], [50]). Different species may have an impact on the microbial composition of the soil and thus nutrient cycles through their different litter chemistry ([38], [51], [52], [53], [54], [55]). Binkley and Hogberg [56] reported lower soil pH in Norway spruce plantations than in plantations with hardwoods, because of the greater acid strength of the organic matter accumulated under the conifers.

Until recently, there were two different paradigms concerning N nutrition (Fig. 1). The first one asserted that plants prevalently used the inorganic N derived from N-inputs and mineralization. Mineralization, i.e. the breakdown of organic monomers performed by heterotrophic microbes releasing NH₄⁺ (ammonification), was considered the step regulating the availability of N for plants. NH₄⁺ was then used as energy source by oxidizing microbes producing NO₂⁻ readily converted to NO₃⁻ (nitrification) and also NO and N₂O (Fig. 1A). The second, more recent paradigm placed less emphasis on the mineralization as a limiting process and suggested that plants could access both the inorganic and organic pool of N ([16], [57]). Through depolymerization (i.e. soil organic matter breakdown operated by microorganisms and plants through the release of exoenzymes) (Fig. 1B), SOM may be decomposed in smaller and more readily available organic compounds (amino acids, nucleic acids, amino sugars). These organic compounds can be

49 mineralized releasing inorganic N (NH_4^+ and NO_3^-), which is more easily available for plants and
 50 microorganisms ([57]).



51

52 **Fig.1. The changing paradigm of the soil N cycle. (A) The dominant paradigm of N cycling up through the**
 53 **middle 1990s. (B) The paradigm as it developed in the late 1990s (from [57])**

54

55 In this paper the latter paradigm is preferred because of its flexibility: it does not refute the role of
 56 mineralization but, at the same time, allows the use of organic N in certain circumstances. This decision is
 57 supported by recent findings demonstrating that mycorrhizal and nonmycorrhizal plants can take up
 58 organic N ([58], [59], [60]). Moreover, with increasing time since fire, in old forest stands, lower soil pH and
 59 the accumulation of organic matter with high concentrations of polyphenols may limit N mineralization,
 60 especially nitrification, with proteolysis (i.e. depolymerization of proteins) potentially gaining more
 61 importance (Fig. 2; [2], [52], [61]). However, mineralization remains important in the boreal forest, since the
 62 low pH and temperature, the accumulation of phenolic-based allelopathic compounds, the wet and
 63 anaerobic conditions that inhibit nitrification, favor the formation and/or accumulation of NH_4^+ ([2], [17],
 64 [42], [62]). Boreal landscapes with complex topography can show a range of N forms ([63], [64], [65],

[66]).When soils extraction are carried out, NH_4^+ is generally the predominant inorganic form of N in mature conifer stands on less fertile sites, while NO_3^- tends to gain importance on the most productive forest ecosystems (e.g. toeslopes, fine-textured fluvial terraces), after disturbances (e.g. producing forest gaps, with temperature, pH and light conditions stimulating mineralization and nitrification) or in areas subject to high N depositions ([65], [67], [68]).An increase in the concentration of amino acid was reported with the accumulation of organic matter along a transect representing a primary succession (Fig. 2, Table 2) ([2], [69]).[70] suggested different sources and sinks of amino acids, to explain an increase with stand age in stands issued of secondary succession. Not only depolymerization of existing soil organic matter (SOM), but also increased direct input through needle litter, root exudates and root turnover, linked to greater aboveground and belowground plant biomass in mature stands, would be at the source of increased organic N inputs.However important differences may exist between primary and secondary succession, with dramatic species changes in the former (and complete changes in litter quality and microclimate) and the same species composition sometimes maintained in the latter (e.g. black spruce and balsam fire establishing immediately after a disturbance and persisting to the next stand-replacing event), probably affecting differently the N cycle ([71], [70]). During secondary succession, pioneer shrubs and trees might drop out over time, rather than dominate and being replaced.Certainly more papers are needed to clarify the differences in N cycle between primary and secondary succession.

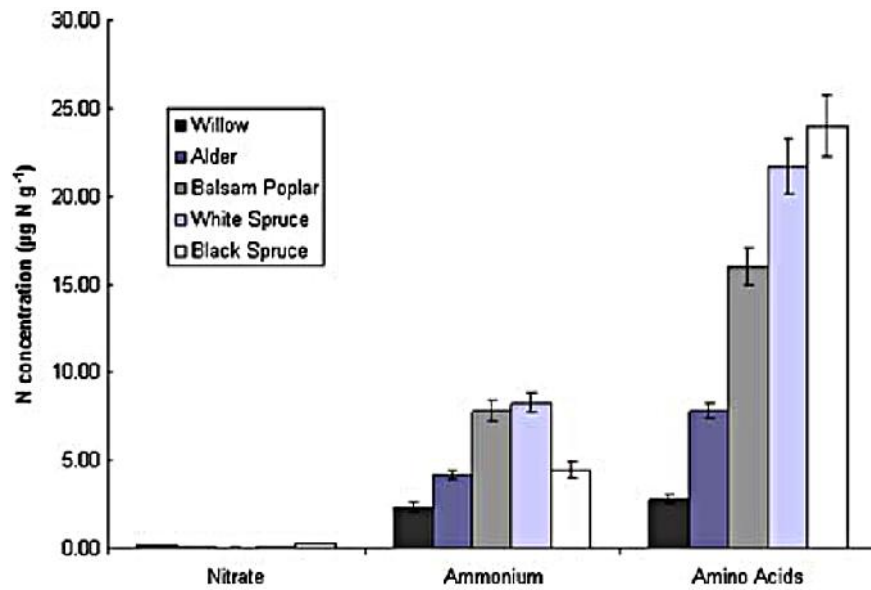


Fig. 2. Concentrations of soil nitrate, ammonium and free amino acids across a primary successional sequence on the Tanana River, interior Alaska. Values are seasonal average concentrations from monthly measurements in June-October. Mean \pm SE, n=3 (from [2])

87 Table 2. Characteristics and estimates of the different N forms in boreal soils

<i>N forms</i>		<i>Characteristic</i>	<i>Estimates</i>	<i>Some references</i>
Organic N	<i>Complex organic molecules</i>	Greatest concentrations in the organic surface horizons	DON 16-32 kg ha ⁻¹ (may be about an order of magnitude greater than DIN, but only a small fraction is considered easily available for plants)	[1], [2], [16], [19], [38], [42], [52], [61], [69]
		Quantitatively the most important in undisturbed northern ecosystems, increasingly supplemented by NH ₄ ⁺ , at first, and NO ₃ ⁻ , secondly, as productivity increases ([66])	Soluble proteins (0.5 mg g ⁻¹ soil, corresponding to c. 0.08 mg protein N g ⁻¹ soil)	
		Constituted by monomeric (e.g. amino acids) and polymeric organic compounds containing N		
	<i>(of which) Amino acids</i>	Major component of simple organic N pool: Glu, Asn, Gln, Asp, Ala and His	Amino acids concentrations in the bulk soil solution: Range 0.1-50 mM	
		At acid and subacid pH reduced mobility of basic amino acids (such as L-Arg and L-Lys) compared to neutral amino acids (e.g. Gly and L-Ala)		

<i>N forms</i>		<i>Characteristic</i>	<i>Estimates</i>	<i>Some references</i>
Inorganic N			<i>DIN</i> $\text{NH}_4^+ \text{NO}_3^-$, 0.9-1.5 kg ha^{-1}	[2],[42], [61], [62], [72]
	NH_4^+	Less mobile than NO_3^- (readily adsorbed to the cation exchange sites in the soil)	Diffusion coefficients: 10- to 100-fold less than NO_3^-	
		Reduced leaching losses compared to NO_3^-	Mean residence time in the FH horizon 0.30-0.86 days	
	NO_3^-	Mobile (due to soil overall negative charge)	Diffusion coefficient is ca. $1 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$	
		Easily lost through leaching	Mean residence time in the FH horizon 0.23-0.75 days	
		Mostly delivered to roots through diffusion and mass flow, guided by the transpirational water stream		

88

89

1.1. Hypotheses of N-limitation

Historically, growth in the northern temperate and boreal forests has been considered essentially N-limited, as N-addition often resulted in increases in productivity ([5], [73], [74], [75]). Vitousek and Howarth [3] proposed that N-limitation may occur through biogeochemical mechanisms (increased losses and immobilization) and limitation to N-fixation (decreased inputs).

The different mobility of N forms, in particular NO_3^- , can lead to greater losses than NH_4^+ , for example during snowmelt, when low soil temperature, high water fluxes, and the long period of tree dormancy limit N uptake ([3], [30]). However, in the boreal forest, high NO_3^- losses are probably rare, given the strong N sinks generally represented by soil and plants in this N-limited environment. Important N losses (e.g. N volatilization by fire and nitrate leaching following fire or clearcut) may be related to disturbances, especially when plant uptake is absent or reduced, while DON losses (which are generally greater than DIN losses in older undisturbed stands) proportionally gain importance during the later phases of stand succession, when organic matter accumulates ([10], [76], [77]). N losses in gaseous forms during fires can be in the order of hundreds of kg N ha^{-1} , but episodic and with different impacts on long-term N trends depending on fire return interval and fire severity ([53]).

The strong C-N bond of litter and soil organic matter was also proposed to explain the lower availability of N ([53], [78]). N would be chemically and physically protected by protein-precipitating and recalcitrant C compounds (polyphenols and tannin), which could slow down its decomposition. Besides, allelopathic effects of certain polyphenolic compounds have been described, which may inhibit N mineralization or N uptake by plants ([15], [17], [79], [80]). Moreover, N-limitation could result from strong competition between soil microorganisms and plants ([81]). So the increased growth of conifer seedlings after weed control is usually attributed to a reduced competition for N ([82]).

In nature, higher rates of N-fixation are observed with low N supply, thus it seems paradoxical that N-limited late-successional boreal forests do not contain N-fixing trees ([83]). However N-fixation may be constrained by the availability of other nutrients (as P, Mo or Fe), by the high energy cost of the symbiosis (high requirement for light) and other mechanisms including the preferential grazing of N-fixing plants by herbivores ([3], [78], [84]).

The more straightforward way to demonstrate the existence of N limitation of trees is to evaluate the response of growth to N additions. However disturbance of the ecosystem and site-specific conditions can complicate the interpretation of the results. Moreover, the absence of a growth response does not reject the hypothesis of limitation, since the added nutrient could be immobilized in another component of the ecosystem (e.g. soil or microbial immobilization or uptake by other plants), due to other strong N sinks ([53], [85]). Other approaches to evaluate limitation in N are the analyses of tissue (especially foliar) concentrations to calculate its critical levels and ratios with other elements (e.g. N:P, [53], [86]).

1.2. Aim of the review

Important reviews have been published on inorganic and organic N nutrition and on the relationships between mycorrhizal fungi and nutrient cycling ([1], [53], [61]). Nevertheless, for its particular features, a specific examination is required concerning the distinctive dynamics of the N cycle occurring in the boreal coniferous ecosystems. The aims of this paper are (1) to describe the importance of the different soil N forms in the nutrition of boreal conifers and the strategies of uptake developed under low N availability; (2) to briefly discuss the impact of present disturbances (namely fire and forest harvesting), increased N depositions and climate warming on the N cycle of the boreal conifer ecosystems; (3) to discuss the limitations of the past studies and provide some recommendations for future investigations.

In this review, N nutrition is defined as the process of uptake, assimilation and use of N and, unless otherwise indicated, the coniferous boreal forest will be taken specifically into account.

2. STRATEGIES FOR N-UP TAKE

Before its assimilation and use, N has to be taken up by trees. Fine roots are especially active sites of nutrient uptake and exchange with mycorrhizal fungi. In white spruce of interior Alaska, Ruess et al. [50] estimated that nearly every first-order fine root (i.e. the most distal, smallest diameter, fine roots) presented mycorrhizal associations. These fungi play an important role in enhancing N nutrition and can directly interact with other soil microorganisms. So, the influence of mycorrhizae in N-nutrition and the

mechanisms of uptake for the different N forms need to be taken into consideration when analyzing the role of N in tree growth.

2.1. Mechanisms of N-uptake

The net uptake of a nutrient results from the balance between influx and efflux and is a process dependent on concentration and regulated by plasma-membrane transporters ([61]). NO_3^- uptake requires active transport and the energy is supplied by ATP ([42]). The uptake of NH_4^+ , like that of K, occurs through channels mediated by the negative cell membrane potential, producing an acidification of the rhizosphere ([42]). The rates of NH_4^+ uptake by nonmycorrhized white spruce seedlings growing in hydroponics could be 20 times greater than that of NO_3^- ([87]). Grenon et al. [88] reported low NO_3^- uptake capacity in spruce suggesting that NO_3^- may be an important nutrient form for soil microbes. Compared to NH_4^+ , NO_3^- uptake and assimilation require 10-15 additional ATP ([89], [90]). Meyer et al. [91] estimates that C costs for the assimilation of NH_4^+ and simple organic N are half those for NO_3^- : 0.17 kg C/kg of NH_4^+ or of $\text{N}_{\text{organic}}$ vs 0.34 kg C/kg of NO_3^- .

Contrary to NO_3^- , NH_4^+ efflux probably takes place through an active antiport, and the associated high energy cost is sometimes considered to be one of the causes of ammonium toxicity in many species that are not adapted to soils with excess NH_4^+ ([87], [72]). Other features of NH_4^+ toxicity are the accumulation of amino acids in plant tissues. Britto and Kronzucker [72] speculated that the competitive exclusion of K^+ by NH_4^+ and the over-expression of K^+ channels at high external NH_4^+ concentrations could cause a high NH_4^+ influx, and consequently an excessive efflux to maintain the electrochemical balance of the cells, leading to NH_4^+ toxicity. Moreover, it has been observed that NH_4^+ toxicity is more pronounced at high light intensities and associated with reduced leaf moisture and water potentials ([42], [72]).

Kronzucker et al. [92] proposed that sensitivity to excesses of NH_4^+ may decrease between early- (trembling aspen and Douglas fir) and late-successional (white spruce) species because of the increased efficiency in controlling NH_4^+ fluxes through the plasmamembrane. Moreover, early successional deciduous species (e.g. aspen) cultivated in hydroponics showed a good capacity of absorbing NO_3^-

concurrently with NH_4^+ , contrary to white spruce and jack pine seedlings ([93]). However, all the species tested showed higher affinities and rates of uptake with NH_4^+ compared to NO_3^- ([93], [94]). Min et al. [95] confirmed this pattern, observing that, compared to NO_3^- , NH_4^+ uptake was 16-fold higher in lodgepole pine, while only 2-3-fold higher in trembling aspen. Similarly, rates of uptake of amino acids and NH_4^+ were 7-8 times higher than those of NO_3^- in a 100-day greenhouse experiment with potted seedlings of Norway spruce and Scots pine supplied with three concentrations of N (1, 3 and 10 mM of total N) in various forms and mixtures (NO_3^- , NH_4^+ , arginine, glycine), but reductions in growth and mortality of seedlings were observed at a high proportion of NH_4^+ in the fertilizer, probably because of NH_4^+ toxicity ([96]). Min et al. [95] observed that, compared to aspen, lodgepole pine and Douglas-fir had similar NH_4^+ utilization but lower capacity of utilizing NO_3^- with lower translocation to shoot and lower levels of nitrate reductase activity. So they proposed the hypothesis that the availability of different N forms may be implicated in niche separation among species, thus influencing the species distribution in temperate and boreal ecosystems.

Compared to inorganic N forms, amino acids and organic N could gain importance in plant nutrition with time since disturbance, given the accumulation of phenolic compounds that may reduce mineralization of soil organic matter ([2], [11], [97], [67], [70]). Plants possess the capacity to take up amino acids and can have mycorrhizal associations that enhance the nutrition of organic N ([58], [98], [99], [100]). A recent laboratory study showed that some nonmycorrhizal species can take up intact amino acids and proteins, but their ability to grow under N sources consisting of solely organic N was limited ([59]). In a field experiment, the uptake of NH_4^+ and amino acids was larger than that of NO_3^- for Norway spruce and common bilberry, while wavy hair-grass showed opposite results ([98]). In another experiment with Scots pine, amino acid uptake was similar to or larger than that of NH_4^+ , while NO_3^- uptake was low ([100]). So, in black spruce and Scots pine the rate of NO_3^- uptake is generally lower than that observed for NH_4^+ and amino acids ([96], [100]).

2.1.1. N-depositions and canopy N uptake: another important pathway?

Apart from N fixation (e.g. by cyanobacteria associated with mosses, especially important in less polluted areas) and the decomposition of organic matter, N enters ecosystems through N-depositions (dry, wet

and gaseous). An important part of the atmospheric inorganic N deposition may be retained within the canopy, particularly for NH_4^+ ([31], [101]). Canopies typically do not retain organic N, but they do retain inorganic N at a rate of $1\text{--}12 \text{ kg N ha}^{-1} \text{ year}^{-1}$, or 50-70% of deposited N at the sites of the study by [102]. Measuring the difference between throughfall and bulk precipitation, Houle et al. [31] estimated that for a coniferous boreal forest (spruce and fir) the uptake of NH_4^+ was significantly higher than that of a deciduous canopy, in agreement with previous studies ([103]), and reached 75% of incoming wet NH_4^+ deposition during the growing season. The uptake of NO_3^- and NH_4^+ was especially pronounced in October, for both canopies, while in January a net release was observed. In an overnight incubation of melting snow, epiphytic lichens effectively removed NO_3^- and NH_4^+ ([31]). Lichens are often present in significant amounts on the branches of coniferous stands and they may be responsible for a significant part of canopy N uptake([31], [104]). Because of this, there is a strong probability that the N uptake of trees themselves (calculated as the difference between total throughfall flux and wet deposition) is overestimated. ([31], [105], [85]). Indeed, in an experiment with balsam fir, the overall reactivity for both efflux and influx was in increasing order new needles, old needles, twigs and, finally, lichens ([104]). An observation of balsam fir tissues with a scanning electron microscope showed that bacterial and fungal coverage on twigs was greater than on needles and, between needles, older ones were more covered than younger ones. So, needle age, degree of microepiphyte cover and abundance of lichen cover are postulated as the three principal variables controlling ion loss or uptake in the canopy of balsam fir ([104]).

In a review on direct foliar uptake of N, Sparks [106] underlined the importance of considering foliar and soil pathways of N incorporation into biota separately. The mechanisms of foliar uptake are still unclear and estimates vary a lot with canopy uptake of reactive N varying between 0-50% of plant N demand. Ignatova and Dambrine [101], comparing throughfall under fake plastic trees and true canopies, estimated that in 8.5 months more than 4 kg ha^{-1} of inorganic N were taken up by the canopies of the different stands and that this represented between 10-30% contribution to annual requirement of N by foliage. However, in a recent labeling experiment with ^{15}N , less than 5% of the label was recovered in live foliage and wood after 2 years of N addition to the canopy ([107]). The majority of the label was in or on twig and branch materials. For these materials the authors weren't able to distinguish between bark and

wood and to establish if canopy N retention was due to physico-chemical interactions with plant surfaces or real uptake ([107]). They concluded that in the short term most of the N was retained on plant surfaces, branches and main-stem bark, with little being assimilated into woody material and little effect upon C sequestration.

Unlike more productive sites, where canopy “uptake” represents only a small fraction of the foliar increment and stand requirement, in low productivity sites, like spruce and spruce-fir sites, canopy “uptake” can be nearly equal to foliar increment ([32]). However, most of tree N requirement is still being met by root uptake and internal retranslocation (resorption). Moreover inorganic N uptake appears to be greatest in spruce-fir canopies with high epiphytic lichen biomass. Organic N is released from forest canopies, but the amount of release is generally less than the amount of inorganic N uptake so that total N is generally consumed ([32]). However, again, it is difficult to differentiate between uptake by epiphytes vs uptake by trees and/or physico-chemical interactions with canopy surfaces. No measurable canopy uptake is usually observed during winter months ([108]), thus pointing out the biological nature of this phenomenon. Indeed, this may be a function of the physiological state of the trees and epiphytes during these months **with** low photosynthesis, reduced growth and limited seasonal demand for N. Epiphytic lichens and mosses are a major component within the old-growth canopy and they definitely affect the flux of nutrients in the throughfall, with greater uptake of inorganic N in stands where epiphytes are abundant ([108]).

It is interesting to underline that it exists a link between N depositions and N fixation at the level of mosses. Some studies have proven that N fixation by microorganism associated with mosses usually decreases as N deposition increases or with higher N availability, early in the forest succession, following fire ([109], [27]). During secondary succession, the formation of a dense carpet of mosses usually starts when canopy closes, and increases with time since disturbance. The N fixed at the level of these mosses may represent an important potential input of N, as soil N availability may be progressively reduced by the accumulation of polyphenols and the decrease in the ratio of mineral N to DON in late-successional stands ([19], [97]). Nonetheless seedlings planted into these feather moss layers establish and grow poorly despite the ability of mosses to retain moisture, probably because mosses are strong sinks for

available nutrients and their litter decomposes slowly ([97]). A part of N arriving through depositions may be used by bryophytes, likely limiting acquisition by woody plants, while N fixation is downregulated ([18]). However at high N levels some species of mosses show reduced biomass and may even disappear.

Given the uncertainties linked to canopy N uptake and that the focus of this critical review on the importance of soil N for the conifers of the boreal forest, we suggest interested readers to consult other more specific reviews on canopy uptake.

2.2. Roots and competition for N

Soil N affects size, structure and distribution of the root system. In Norway spruce and silver birch, starch accumulates in leaves when N is limiting, and additional amounts of photosynthates are translocated to the roots, thus allowing an increase of the size of the root system ([42], [110]). It is assumed that, in roots and shoots, the balance between N and C influences the processes associated with C fixation or formation of new tissues and determines the allocation of resources between belowground and aboveground components ([110], [111]). Factors like soil temperature, moisture and nutrient availability play a role in the timing and duration of root growth, while root longevity is controlled by microsite conditions, patterns of development and length of the growing season ([112]).

Soil microorganisms play a key role in regulating the availability of nutrients through SOM depolymerization and mineralization. Except for symbionts (e.g. mycorrhizae), the other soil microorganisms are often C-limited resulting in a strong competition for energy sources, especially when availability of labile C substrates is limited by the accumulation of recalcitrant organic compounds (high lignin:N ratio, high content of polyphenols, tannins and lignin) ([17], [54], [113], [114], [115], [116]). The increased availability of labile C in the rhizosphere can strongly stimulate microbial activity through a priming effect ([11], [38], [113]). However, microorganisms can also be limited by the availability of mineral nutrients as N and P ([62], [114]).

Some experiments with isotopic tracers have found that plants are inferior to microbes in the uptake of inorganic and organic N in the short term (one to several days), but they acquire more and more of the

tracer over longer periods (weeks or months) ([61]). Various elements are important in the longterm, including microbial turnover, competition between mycorrhizae and microorganisms, lifespan of the tree tissues with great sink potential and strategies of effective nutrient conservation ([117], [118]).

Apart from the competition with microbes, the nutrition of conifers is also influenced by the competition with other plants. A reduced competition could explain the strong positive effects of weed-suppression on survival and growth of boreal conifer seedlings, leading to higher foliar concentrations of N and other nutrients ([93], [119]). Plants can compete in different ways. For example, sheep laurel is able to sequester soil N through the formation of tannin-protein complexes that are not easily accessible to black spruce, thus reducing its nutrition ([80]). Yamasaki et al. [79] observed that foliar N concentrations in black spruce were positively correlated with the height and biomass of seedlings growing away from sheep laurel, and proposed that the reduced mineralization alone is unable to explain this pattern. They proposed that sheep laurel could affect the growth of black spruce through four different mechanisms: (1) competition for nutrients; (2) direct allelopathic effects on black spruce, 3) allelopathic effects on mycorrhizal formation or maintenance, 4) increased susceptibility to root pathogens, as a consequence of the decreased mycorrhization. Some authors have suggested that polyphenols may inhibit microbial activity and thus reduce N mineralization, but results are still contradictory ([14], [19], [97]).

2.3. Mycorrhizal associations and N nutrition

Boreal forests are characterized by the prevalence of two groups of mycorrhizal fungi: ECM, associated with conifers, and ERM, associated with the ericaceous shrubs that spread in heathlands and forest understory, but arbuscular mycorrhizae associated with grasses are found after disturbances or in more fertile microsites ([120], [121], [65]). Mycorrhizal fungi are considered to play an important role in plant nutrition, especially when N is scarce. The abundance of different N forms and mycorrhizal associations and their host species are related. Along a short transect in a fennoscandian boreal forest, a series of studies ([63], [64], [65]) observed that a shift in vegetation and productivity corresponded to a change from inorganic to organic N forms in soils. In particular, NH_4^+ increased in the Norway spruce/short-herb type, and finally in the tall-herb type NO_3^- was as abundant as NH_4^+ and plants took up nearly equal amounts of the two mineral ions. The productivity increased along the gradient of concentration of soil N

and the change in plant community was associated with changes in the mycorrhizal community, from a dominance of ECM and ERM in the Scots pine/dwarf-shrub type to a prevalence of arbuscular mycorrhizal fungi in the tall-herb type ([63], [64]). C:N ratio and pH seem to be good predictors of changes in microbial community structure, with high and low ratios associated with fungi and bacteria, respectively. A negative correlation has also been reported between C:N ratio and N mineralization rates in some Scots pine stands in Sweden ([114], [122], [123]).

ECM are effective in the uptake of NH_4^+ and low concentrations of this ion in the soil solution of some boreal forests may be in part a consequence of a rapid uptake by mycorrhizal roots ([89]). For example, Nilsson and Wallander [124] excluded roots of Norway spruce colonized by ECM from a portion of soil (through plastic tubes), and observed that NH_4^+ and NO_3^- concentrations were higher in the portion where roots were excluded. In Northern Sweden, in a large tree-girdling experiment with Scots pine realized by killing trees by means of cutting the phloem and cambial tissues around the stem, soil respiration was reduced by ca. 50% after a few weeks and microbial biomass by one third after 1-3 months ([123], [125]). Analyzing phospholipid fatty acid, the 45% decrease in a fungal biomarker associated with ECM suggested that the decrease in microbial biomass was mainly due to loss of ECM fungi. Moreover, the higher N content and growth of dwarf shrubs after tree-girdling revealed that the conifers associated with ECM fungi were efficient competitors for N ([123]). These findings clearly demonstrate the peculiar importance of ECM associations in soils of the boreal forest.

Compared with ERM and saprotrophic fungi, ECM show a reduced capacity to take up N from protein-polyphenol complex ([14]). Considering different $\delta^{15}\text{N}$ as an indication of niche separation and access to different N pools, Schulze et al. [126] suggested that ECM of white spruce would use NH_4^+ , NO_3^- and organic N from litter, while ERM of *Vaccinium* spp. would break down more complex slowly-decomposing organic matter. Read et al. [1] proposed that proximity, achieved through an extensive colonization of organic horizons containing important quantities of N and P (especially the FH layer), is an effective strategy to gain access to these nutrients before the formation of the polyphenol complexes that reduce the availability of N for the trees associated with ECM. In the organic horizons, microbial biomass and

mesofauna could also be an important source of uncomplexed organic N, as nutrients become available with microbial turnover or through direct predation of certain mesofauna (e.g. nematodes; [117], [127]).

Recently, Paul et al. [21] have observed the association of N-fixing bacteria with the ECM *Suillus tomentosus* on lodgepole pine in tubercles growing inside rotting wood, possibly acting as a reservoir for moisture during periods of drought. They also report that N-fixation was higher in young stands and during the dry summers (10-15% of the values observed in alder). Moreover, associations between N-fixing bacteria and mycorrhizae were observed in Douglas fir ([25]). Given the relevance of these findings for the N-limited boreal forests, it could be interesting to explore if the presence is limited to the youngest stands or if they are also associated with adult and older trees.

Recent studies pointed out that N-fixation by algae living on mosses can equal inputs from atmospheric N depositions, especially in the late-successional stages of forest development at sites with low N depositions ([26],[27], [109]). Mycorrhizal fungi are able to efficiently colonize dead and senescing bryophytes, but it is not clear to what extent they are able to mobilize both N and P from living tissues of mosses ([128], [129]). So, the role of these widespread mosses and their interactions with mycorrhizal fungi in providing an additional supply of N to the boreal forest deserves further investigation ([26], [28], [130]).

2.4. Key-concepts of N uptake

The boreal conifers have adapted strategies to cope with the limited availability of N. Even if canopy N uptake may contribute, especially in low productive sites, to N nutrition in conifers, a lot of uncertainties remain on the subject, not allowing evaluating the real importance of these mechanisms. In response to the reduced mobility (low diffusion coefficients, see table 2) of certain soil N forms, especially of the heavier organic ones, root proliferation to increase interception seems to gain importance over diffusion in the soil solution and the plasticity (capacity of exploration and rapid colonization of fertile patches) of the rooting system become important when competing for N resources. ECM confer an advantage to trees through the extensive absorbing surface, the increased exploration of the soil micropores and colonization of fertile patches, the wider enzymatic capabilities compared to plant roots alone and the improved access to a wide variety of sources of nutrients. Conifers usually take up ammonium at

levels comparable to simple organic N, which probably grows in importance as organic matter accumulates with stand age, while estimates of nitrate uptake are generally lower in boreal conifer seedlings, but relatively higher in grasses and broadleaves.

3. N METABOLISM AND USES

After assimilation, N is transported towards organs and tissues where it is stored or used for different functions, depending on plant requirements. Transport of amino acids occurs both in xylem and phloem ([42]). The reserves of proteins and amino acids in stem, roots and older leaves are rapidly remobilized in spring to sustain shoot elongation, bud burst and development of needles ([131], [132]). Some experiments in conifer seedlings have found that N retranslocation is independent of current N supply and that needle development relies mostly on N from pre-existing shoots ([131], [133], [134], [135]). However, some differences between species have been observed. For example, the first phases of leaf growth of Scots pine and silver birch were sustained concurrently by root uptake and remobilization, while *Sorbus aucuparia* remobilized half the N from storage before additional N was taken up by roots. Black spruce seedlings receiving a high nutrient fertilization before plantation showed greater height and biomass growth than unfertilized seedlings ([136]), pointing again to the importance of internal nutrient reserves. It has been estimated that in some mature conifers, the internal cycling may provide between 30 and 60% of the N contained in the new foliage ([133]).

A strong positive relationship is observed between N concentration in leaves and photosynthetic capacity, the latter varying with P availability and across biomes with the highest slope and lowest intercept at the lowest N:P ratio, corresponding to the arctic and boreal ecosystems ([137]). N fertilization increases the photosynthetic capacity but also stimulates foliage production ([138]). Evans et al. [139] observed that by increasing N supply, balsam fir accumulated N in the foliar tissues even if this did not translate into increased growth, but the results were not confirmed for heart-leaf paper birch. In a mature black spruce site, 3-year-long N additions increased N, Ca, Mg and Mn foliar concentrations without affecting growth ([77]). It has been suggested that slow growing species could respond to nutrient stress by adopting strategies to maintain adequate internal concentrations of N and other nutrients through luxury

consumption (and storage) during the periods when resources are more available (e.g. at the beginning of the growing season) ([140], [141]). Indeed, compared to white spruce, growth of black spruce was less reduced in response to low N conditions, while the absorption rate was higher at high N conditions, even if growth was similar ([141]). This was interpreted as a better adaptation of black spruce to low nutrient availability.

In a greenhouse experiment with boreal tree species, Reich et al. [142] measured low rates of N uptake in conifers, while broadleaf species showed high rates of N uptake. In the species tested, the rates of N uptake were related to the relative growth rates and interacted with light availability. At low light, the effect of increased soil fertility was less expressed ([143]). Wood properties are also likely to be influenced by N ([144], [145], [146]) as N is implicated in cell lignification ([147]), stimulates photosynthesis ([148]), and affects growth rate (in height and diameter, e.g. [149], [150]). Indeed, many studies report increases in radial growth after N-fertilization ([144], [151]). Fertilization also affects wood structure. In Norway spruce increases in ring-width are often the result of increased proportions of earlywood, since earlywood has usually lower density than latewood, wood density decreases ([144]).

3.1. Key-concepts of N metabolism and uses

N taken up from the soil is generally assimilated into organic N compounds (mainly amino acids) before it can be used in plant metabolism. Following assimilation, N is transported towards the developing organs to sustain growth or stored in the form of reserve. N is an important constituent of photosynthetic proteins and enzymes, so increases in N-availability result in increased net photosynthesis and foliage production. There is a positive relation between N uptake and relative growth and N fertilization increases biomass production. Boreal conifers, especially the slow growing species, seem to rely much on the stored N and its internal cycling, so growth is partially independent of current N supply in the soil, especially at the beginning of the growing season.

4. ANTHROPIC AND NATURAL FACTORS AFFECTING THE N-CYCLE

4.1. N-depositions and boreal forests: a brief account

N-cycle and plant nutrition have been greatly altered in the last two hundred years by anthropogenic N depositions ([6], [152], [153]). This additional input is due to N pollution linked to fossil fuel consumption and it is usually accompanied by changes in precipitation pH, increased S depositions and, in the long-term, may result in changes in the abundance and composition of the soil N pool and of other nutrients (e.g. increased loss of base cations, due to the acidification of the soil by acid rain). In boreal and temperate forests, considered N-limited ecosystems, N addition through wet and dry depositions are very likely responsible of increased forest growth and C sequestration observed in the 20th century ([153], [154], [155]). Magnani et al. [155] showed that the increase in C sequestration in the last century was mainly determined by the additional input of N induced by human activities. Even if the estimates proposed by Magnani et al. [155] are among the highest, a recent review on the subject confirmed the stimulating effect of N-deposition on forest growth ([75]). De Vries et al. [75] reported positive forest growth under low to moderate N additions, with increase in site productivity for Scots pine and Norway spruce between 1 and 4% in response to a fertilizing effect of N deposition of 1 kg of N ha⁻¹ year⁻¹.

It has been proposed that in areas exposed to high N-depositions, tree growth may shift from limitation in N to limitation in other nutrients, as base cations (Ca, Mg, K) or P ([56], [156], [157], [158]). Addition of NPK fertilizers, increased the response of C-sequestration to N alone at N rich sites (soil C:N ratio below 25), possibly pointing out a limitation of P and K for tree growth ([75]). In a survey on the response of Swedish forests to increased N depositions, Binkley and Hogberg [56] reported that growth increased by about 30% from the 1950s to 1990s, and only stands that received heavy N fertilizations responded to fertilization with P or base cations or trace amounts of boron. However Houle and Moore [77] found no response to N-additions in a balsam fir and black spruce dominated site of eastern Canada where N depositions were low (5.7 kg ha⁻¹ yr⁻¹) but concentrations of foliar N were already high in the control trees (1.5% of dry weight). They proposed that this could be due to the high retention by other components of the ecosystem (e.g. soil immobilization) or, alternatively, to a limitation by other nutrients. Similarly, after three years of simulated increased N-depositions, in a balsam fir and in a black spruce stands no

significant change in soil N or in tree growth were observed, underlining the fact that strong N sinks may be present in these boreal coniferous forests ([105], [85])

In an experiment combining heating and fertilization, stem growth of Norway spruce increased by 84 and 25% in heated and in heated plus fertilized plots, respectively ([159]). In various experiments, the lower fertilizer applications or lower N depositions were always more efficient in stimulating growth than greater ones ([73], [153]). C accumulation in response to N addition was 25 kg C/kg N and 11 kg C/kg N added for tree and soil, respectively. The application of NPK fertilization improved tree C accumulation to 38 kg C/kg N, suggesting the occurrence of co-limitation by P and K in these sites. Comparing the sites from northern and southern Sweden, higher N-depositions in the southern Norway spruce stands were estimated to be responsible for an increase of 2.0 kg m⁻² and 1.3 kg m⁻² of C in tree and in soil organic carbon, respectively ([73]). In a 40-yr old Norway spruce stand, long-term N-addition resulted in 10-fold additional carbon storage (3.0 Mg C ha⁻¹ yr⁻¹) compared to the effect on the humus layer (0.3 Mg C ha⁻¹ yr⁻¹). Even if production and litterfall were increased, no increase was observed in soil heterotrophic activity, suggesting a possible negative effect of N on SOM decomposition ([160]). In fact, some studies have found that moderate or high N additions (>5-10 kg N ha⁻¹ yr⁻¹) inhibit the decomposition of low quality litter with a repression of the lignolytic enzymes of decomposing fungi ([38], [161]).

The presence of mycorrhizal fungi usually decreases with N depositions ([162], [163]). This may be the consequence of direct changes in the soil conditions or in belowground/aboveground allocation by trees. In a Norway spruce stand, fine root mortality increased by 191% in fertilized plots ([164]). Other studies employing N-fertilization found contrasting results on the diversity of fungal species ([165], [166], [167]). For example, Rossi et al. [167], found an increase in vital root tips, in root tips showing ectomycorrhizae and in number of morphotypes in their high N treatment (+30 kg N ha⁻¹ yr⁻¹, representing 10 times the current N deposition), relative to the control (no N added), after 8 years of ammonium nitrate addition in a natural black spruce stand of the Canadian boreal forest. The observed contrasting results could be related to the different host species and their fungal partners, the quantity of N added (i.e. high N loads may be deleterious), the form of N added (e.g. organic N may increase ECM; see [165]), the composition of the microbial community and the plant response to N addition (e.g. positive N fertilization

effects on tree growth in N poor sites would result in increased C fixation and availability of photosynthates for the fungal partner; ([168], [167]). Moreover, there are reports that some mycorrhizal species may be well adapted to soils with high rates of N mineralization and the optimum N concentration could evidently differ between the fungal species ([162], [166], [167]).

4.2. Forest harvesting, fire and climate change: the impact of different anthropic and natural disturbances on the N cycle

Disturbances affect the N cycle, but according to the type and severity of disturbance, their effects may differ. Some studies have investigated the effect of forest management (e.g. clearcut and variable-retention harvesting) and wildfires, which are probably the most common anthropic and natural disturbances in boreal forests. In a recent meta-analysis, [68] reported increased soil nitrogen concentration as nitrate, N concentration in leachates and nitrification rates and pH, but no changes in soil inorganic nitrogen concentration, ammonification and N mineralization rates after clearcut. The effect of clearcut was delayed in coniferous stands relative to deciduous stands but persisted for several years. In various studies, the removal of group of trees (gap creation) produced bigger changes in nitrates than single-tree selection, when roughly the same number of trees was removed ([169], [68]). [169] observed similar rates of decomposition of litter and forest floor in gaps of all sizes (similar or slower to that of uncut forests) and that the nature of the forest floor or soil had a greater influence on nitrate concentrations than changes in environmental conditions in the gaps. [170] reported significantly lower amount of soil organic N in young wildfire-burned stands, whereas young clearcut and mature stands had similar quantities of soil N. They also observed that the most pronounced difference between disturbance types was for net nitrification and concluded that, given the different mobility of nitrate and ammonium, differences between clearcutting and wildfires can have important consequences for plant nutrition and leaching losses following disturbance. [171] concluded that fire had no significant influence on soil N amount or concentration across all studies “screened” in their meta-analysis. They also concluded that the soil layer that is most influenced by fire is limited to the upper several centimeters.

Post-disturbance increases in the concentration and production of mineral N are generally rapid and short lived (from some months to a few years, [171], [170], [67]). After an initial “spike” following fire, increases in N availability are often followed by a decline ([67]). [70], studying chronosequences spanning from 4 to 60 years since stand-destroying fire, observed an increase in amino acid concentration and a parabolic trend in mineral N with stand age, with mineral N decreasing after an initial high concentration and then increasing again. Similarly, in jack pine forests, N mineralization rates decreased to low levels by 14 years following fire and increased again through 70 years, maybe because of increasing stocks on soil organic matter ([172]). However, other studies found that net N mineralization rates may decline with stand age ([71], [19]). Vitousek et al. [71] suggested that evidence is insufficient to decide which pattern is more widespread and speculated that climate may affect the direction (increase or decrease) in N availability late in secondary succession.

Climate change may also impact N cycles through direct effects on N mineralization via changes in temperature and precipitations (e.g. soil moisture), and indirect effects on species composition and growth, as well as impacts on natural disturbance regimes. Through model simulations, Smithwick et al. [173] observed an increase in forest production and net N mineralization under future climate scenarios relative to current climate. Other syntheses and meta-analyses also reported increases in N mineralization with increase in soil temperatures ([174], [55]). Soil organic matter C and N were resilient to changes in fire return intervals and densities. For example, given historical fire return intervals between 100 and 300 years and that the N stocks were recovered less than 100 years following disturbance, [173] concluded that fire return intervals would need to decrease dramatically to affect long-term N and C storage at Yellowstone (USA), due to low aboveground N losses via combustion, the large soil N pool and relatively fast recovery after fire. Similarly, for sub-boreal spruce zone of the central interior British Columbia, Canada, [175] observed that a period of 14 years after a forest fire was sufficient to restore the pre-fire level of total N in soils. However complex feedbacks and an incomplete understanding of many mechanisms and processes limit our predictive capabilities.

Chen et al. [176] suggested that, as an effect of climate change, an increase in Net Primary Production (NPP) of the order of 30% could be anticipated in Canada's boreal forests. Modeling the response of

forest C dynamics in the boreal forest, [177] pointed out that it is not likely that all boreal forests will exhibit enhanced growth as an outcome of global change. The boreal forest may become a C sink or source according to responses in plant growth, decomposition and disturbance regimes. According to [178], there will be a doubling in the area burned annually by wildfire in Canada. However, a thorough consideration of climate change effects on the N cycle is not an easy task and is beyond the scope of our review. We suggest the interested reader to consult modeling studies published in the years to come. The integration of C-N feedbacks, also taking in consideration anthropic and natural disturbance regimes, in models predicting global change is at the frontier of current research and scientific efforts in this sense are encouraged.

5. LIMITATIONS AND KEY-FINDINGS OF PAST STUDIES

5.1. Different N forms and plant nutrition

There is evidence that in boreal soils, NO_3^- levels are often very low (except in the more fertile soils, in particular topographical situations and in the first years following disturbance) while NH_4^+ and organic N usually represent the majority of the N pool, with an increase in the ratio of organic to inorganic N, late in the secondary succession, with the accumulation of organic matter, or along a decreasing gradient of site productivity ([2], [66], [97]). NO_3^- was thus not considered the most important source of N for conifers of the majority of the boreal forest. However, the rate of flux through the different N pools could be more important than their size ([76]). Moreover in experiments with lysimeters, which represent concentrations in the soil solution (sometimes considered more easily available for plants) rather than total nitrogen present in the soil (soil extractions), NO_3^- is usually well represented ([36]). So these conclusions, about different soil N forms, have to be taken with caution because we don't know yet which measure better represent available N for plant growth. Hydroponics experiments do not take into account the mobility of the different N forms in the soil, and thus fail to quantify the real availability and plant preference for these forms. Nonetheless, various experiments performed in conifers have confirmed that uptake of NO_3^- is slower than that of NH_4^+ and amino acid, thus limiting the importance of NO_3^- as a primary source of N for conifer nutrition ([58], [100]). Instead, NO_3^- probably acts as a signal to trigger the proliferation of roots towards fertile patches of soil ([57]).

5.2. Mycorrhizal fungi and N nutrition

ECM fungi enhance conifer nutrition through several mechanisms. One of the most important and controversial issues is their ability to use organic N. Jones et al. [76] proposed that DON is abundant in boreal soils because it has a reduced availability for plant and microbial nutrition, and thus accumulates. Moreover, if the trees associated with ECM were able to use this N source, why should N-limitation persist? The answer may be in the cost required by mycorrhizal associations for the use of organic N ([179]). For breaking-down organic matter in order to absorb low molecular weight organic compounds, mycorrhizal fungi and plants have to release exoenzymes ([180]). Since N is required to produce enzymes, the return on investment may be low if organic N decomposition is inhibited ([179]). This might explain why productivity is lower than that measured in soils where inorganic N is more abundant ([63], [64]). Polyphenols and other recalcitrant substances may reduce the ability of ECM exoenzymes to breakdown organic matter ([14]). However, it is still unclear to what extent polyphenols directly reduce the uptake by ECM trees ([19], [79]). Bending and Read [14] report that polyphenols may inhibit ECM when they are in solution, but not when they are precipitated with proteins.

It should be stressed that early studies have often been conducted in greenhouses and in vitro, sometimes using hydroponics or agar-cultures ([14], [61]). Another potential issue with controlled studies is that the role of ectomycorrhizae is largely generic, with no ability to control the assemblage of fungal species on a root system, even if it has been observed a considerable diversity in functional attributes among ectomycorrhizal species ([181]). Moreover, the majority of the field experiments use seedlings, so it is difficult to correctly evaluate how these results can be scaled up to adult or mature trees with an extensive network of ECM roots and important internal storage of nutrients. For example, root exudates (substances released in the rooting zone) vary with species and age, underlining the problem of extrapolating results from studies on seedlings to older plants ([49]). Besides, in a natural forest, the competition with the surrounding vegetation (e.g. ericaceous species) and other microbial communities adds complexity to the interpretation of results. The chemical composition of root exudates also varies in the presence of microorganisms (e.g. mycorrhizae) that can cycle them through their metabolism ([49]). As pointed out by Nasholm and Persson [118], a major problem in studying the competition between plants and soil microorganisms is to distinguish among the different microorganisms in the field (e.g.

mycorrhizae vs. other microorganisms), thus short-term studies can underestimate N acquisition by trees. However, recently, the analysis of phospholipid fatty acids (with biomarkers for ECM, saprotrophic and bacterial communities) has been used profitably to distinguish between the different microbial groups ([123]).

Many experiments testing the importance of different N forms in the field have often added N at unrealistic rates, thus impeding an evaluation of the effective nutrition in natural conditions, but rather simulating a saturating condition ([61], [105], [85]). Some experiments have observed that growth may be reduced at high concentrations of certain amino acids, especially for those compounds that are generally present at low endogenous concentrations in plants ([61]). Given that the uptake of organic N differs between amino acids, the widespread use of glycine in many studies is subject to criticism, since this amino acid can be rare in boreal soils (3% of total free amino acid concentration) while alanine or glutamine could be used as substitutes ([69], [100]). Finally, experiments focus essentially on uptake, so the contribution of the organic N taken up, to the total N plant, is not known ([61], [76]). Jones et al. [76] report that, in some grass species, the contribution of the organic N could be low, but it should be noted that, contrary to arbuscular mycorrhizal fungi associated with grasses, ECM always show greater capacities to use organic N ([179]).

Some doubts still remain concerning the regulation of exchanges and transfer of nutrients between ECM and their hosts ([179]). The studies with excised roots do not account for the effect of exudation and transfer of C to the mycorrhizae. These processes may be key elements in defining N dynamics in the soil and could affect N uptake and transfer ([123]). In both lab and field studies, mycorrhizal plants are depleted in $\delta^{15}\text{N}$, while mycorrhizal fungi are enriched relative to the supplied N sources or soil N ([182]), suggesting that N taken up by mycorrhizae is “selectively filtered” (i.e. metabolized) by the fungi before being transferred to the host tree. Moreover, a delay could occur between mycorrhizal uptake and transfer to the host, so mycorrhizal fungi may store N and make it temporarily unavailable to the partner ([183]).

Future studies with dual labeled amino acids (i.e. with both C and N isotopes) and labeled inorganic N could allow a better understanding of the uptake and metabolism of these compounds ([61]). By using

small amounts of highly enriched tracers in long-term studies it should be possible to study uptake and assimilation at more realistic concentrations and to characterize how the partitioning of N varies with time according to plant requirements. Finally, it will be important to better quantify how much of total plant N is represented by the labeled N compounds taken up.

5.3. N uses in conifers

Some studies report that uptake and assimilation of NH_4^+ are less energy-expensive compared to those of NO_3^- ([76]). However, NO_3^- is transported without the toxicity problems of NH_4^+ . Thus, NH_4^+ has to be assimilated in the organic compounds before it can be transported to those organs where N is needed and, together with amino acids, its translocation is slower than that of NO_3^- ([100]). This may explain the importance of internal N stores in conifers, which may rely less on current N supply in the soil for the formation of new tissues in spring ([131]). Moreover, many boreal conifers, especially some slow growing species, show luxury consumption of N and may use the stored N to sustain growth or survive during periods of reduced N supply ([140]). This can be seen as a strategy for nutrient conservation in poor soils where N is not readily available. Various traits of conifers (low tissue N, long leaf lifespan, high concentrations of phenols and other defense compounds) are also interpreted as strategies for nutrient conservation and adaptations to nutrient-poor environments ([184], [185], [186]).

Reich et al. [186] proposed that the adaptation to the nutrient-poor environment typical of conifer species of native habitats have generated a series of correlated leaf traits, a “syndrome” of slow growing species. This may have implications for competition and forest succession, since these traits are also involved in the low responsiveness of conifers to environmental changes when compared to deciduous species ([184]). One of the trade-offs of longer leaf lifespan could be lower photosynthesis, probably due to diffusional constraints and storage of N in Rubisco ([187], [188]). Slow growing species may be more adapted to conditions typical of late successions while fast-growing species, with their higher photosynthesis rates, usually have higher rates of nutrient uptake that cannot be sustained where nutrient availability is limited ([142]). On the contrary, where resources are not limiting, slow growing species may be less competitive and thus be excluded.

N is also implicated in the biosynthesis of lignin and the changes in photosynthesis and growth following N additions may affect wood properties ([144], [146]). However it is not clear how N supplies vary at stand level and determine differences among groups of trees under otherwise common environmental conditions ([146]), even if, at a broader scale, a positive relationship exists between mineralization and productivity ([64], [148]). In a recent publication, Coates et al. ([189]) demonstrated that the effect and importance of soil fertility in sub-boreal forests on radial growth of different tree species was context-dependent, affected by competition in species-specific manner and varying with the composition of local neighbourhood of the tree.

5.4. Disturbances, N-depositions and stand development: implications for N cycling

Fertilization tests have found that the majority of N is retained by the soil rather than vegetation, confirming that boreal soils are strong sinks for N. The capacity of N retention in soils depends on the history of the past depositions and disturbances (e.g. fire frequency and intensity), soil chemistry (e.g. C:N ratio) and physical properties (e.g. texture) ([4]). The accumulation of organic matter, which has a high exchange capacity at low pH, can increase NH_4^+ retention. Following N-depositions, NO_3^- leaching has been observed especially in systems approaching saturation (which are currently rare in boreal forests) or during winter, when N uptake is reduced ([31], [190], [191]). Apart from the excessive quantity added, one of the problems of many N-addition experiments is the low frequency of applications. In nature, gradual changes and lower deposition levels are more common, so these unnaturally-high and punctual additions can alter the structure and competition of the microbial community ([123], [125]). Nonetheless, these experiments are a realistic approximation of the situation following disturbances (especially in more polluted areas), and their results should be interpreted and applied in this sense. Usually, N-additions have stimulated aboveground rather than belowground growth, indicating that, at high N availability, trees invest fewer resources for soil exploration and N uptake ([110], [111]). Future efforts should be directed towards reducing the quantities and increasing the frequency of N-additions, using highly enriched isotopic tracers and taking into account the specific effects of the different N forms in plant nutrition.

The clear responses to N fertilization have confirmed that N-limitation is widespread in the boreal forest ([73], [75]). The changes in N cycle following fire may favor fast-growing species, resulting in a temporary release from N-limitation. Nonetheless, it is reasonable to expect that frequent fires, leading to continuous N losses, can reduce N availability. Moreover, **wildfires and harvesting** seem to affect differently the N cycle: for example even if harvesting and wildfires increased initial N losses, DON exports were higher in harvested areas compared to burnt areas, where inorganic and organic N exports were equally important ([37]). Other disturbances, like insect outbreaks, may also increase N losses and alter the N cycle (e.g. nitrification), thus affecting forest growth ([36]). Harvesting, especially that of whole trees, removes important quantities of certain nutrients, such as Ca, Mg and P, from the ecosystem, so shifting the growth limitation from N towards other elements, especially in stands chronically subject to high N depositions ([37], [157], [158], [192], [193]). It should be underlined that the occurrence of co-limitation is probably restricted to limited areas, where acidic depositions and harvesting have been intense for a long period of time ([56], [158]).

During stand development after a major disturbance, the forest becomes **denser**, the competition for N increases, and the availability of mobile forms (NO_3^-) is reduced ([67], [194]), gradually leading trees to invest more resources for N scavenging and favoring the belowground allocation towards mycorrhizae, which are particularly adapted for this function ([1], [19]). N-limitation can also be related to the competition with ericaceous species and soil microorganisms, as demonstrated by weed-control and experiments excluding roots of competing plants that resulted in an improved nutrition of conifer seedlings ([81], [82]). The reduced decomposition at high latitudes could be an effect of low temperatures on the forest floors when the canopy closes and organic matter insulation increases, the prevalence of low quality litter, low pH, and the particular microbial communities ([19], [28], [52]). The accumulation of organic matter, increased proportions of recalcitrant compounds, and increased soil acidity could affect the availability of mineral N, **in mature stands**.

6. CONCLUDING REMARKS AND RESEARCH DIRECTIONS

The majority of the studies support the hypothesis that the growth of the boreal forest is limited by the availability of N. Nonetheless, the availability of N may change and be more limited in the late stages of stand development, when low rates of decomposition and mineralization cause an accumulation of organic matter on the soil. Organic N is less available to conifers than inorganic N, which predominates after disturbances (even in forest gaps, if above a certain size, [68]). Consequently, conifers of older forests probably rely more on organic N than earlier in the stand development, closer to the disturbance at the origin of the new stand.

Read and Perez-Moreno [121] proposed that the role of mycorrhizal associations gains importance for N nutrition moving from temperate towards boreal forest and tundra, with ERM especially important in tundra, where the decomposition of organic matter is extremely reduced due to the low temperatures. We refine their hypothesis, based on our conclusion that N limitation could vary overtime according to the stand age and the history of past disturbances (e.g. changes in N-depositions), and that the composition of soil microbial communities varies with N cycle (e.g. C:N ratio, pH and variability of N forms). We deduce that young stands of secondary successions may be less N limited since the disturbances, resetting the succession, tends to increase mineralization and to release the N stored in the soils of the mature and old stands. Future studies should test this hypothesis using chronosequences or manipulating the N cycle and testing how competitive relations may change by measuring the increase in growth after fertilization performed at low rates and concentrations, simulating natural conditions.

Changes in species composition and plant productivity are paralleled by changes in N cycle: the less available and less mobile forms of N are, the higher the dependence on symbiotic fungi for nutrition is. The importance of the processes operated by bacteria decreases in some less productive conifer stands. Boreal conifers of the late-successional stages seem adapted to these conditions and show low rates of N uptake, associated with low rates of growth. Consequently, the majority of these late-successional species are classified as slow growing. However, a doubt arises that some young conifers may show rapid juvenile growth after a major disturbance that modifies the nutrient cycles ([195]). We propose that the rate of growth, which is related to the nutrient uptake, the capacity to adjust growth to

699 changes in availability of N and nutrients, and the presence of different species (conifers and broadleaves)
700 may drastically alter the competitive relations between organisms and define the development of the
701 ecosystem following disturbance. Of course, we acknowledge that soil nutrient availability is not the only
702 factor affecting the rate of growth or forest succession. Light and climatic factors play a fundamental role
703 and must be considered in conjunction with biotic and soil factors, like competition with neighboring plants
704 and availability of adequate substrate for seedling establishment and growth ([196], [189]).

705 The increase in anthropogenic N depositions in the last century seems to have alleviated N limitation and
706 stimulated tree growth in certain regions of the boreal forest. The increase in N depositions, coupled with
707 climate change, may affect the N cycles, increasing the decomposition of soil organic matter and the
708 availability of inorganic N, thus reducing N limitation in the long term, but also altering competitive
709 relations among species. Fast-growing early-successional broadleaf species could be more responsive to
710 these changes because of their closer relationship between N and photosynthesis and their higher rates
711 of N uptake and growth, and could gain advantage over conifers. It is urgent to test this hypothesis
712 through manipulative experiments that simulate scenarios of N enrichment coupled with climate
713 warming to understand the potential responses of the boreal ecosystems and their consequent future
714 evolution.

715 This paper focused the role of soil N in the nutrition of boreal conifers, but N depositions may also affect
716 N nutrition and, as a result, impact soil N cycle through direct canopy uptake. This process is insufficiently
717 understood (a brief account of current knowledge has been given in the text), so we suggest that future
718 research should aim to verify and define its importance in respect to N uptake and study its impact on the
719 microbial, and in particular mycorrhizal, communities of the boreal ecosystems. Indeed, future research
720 should try to improve our understanding of the possible outcomes of changes in disturbance regimes, N-
721 depositions and climate, including the role of N fixation by mosses, canopy N uptake and the responses
722 of conifers in relation to changes in microbial (symbiotic and not) communities. Finally it is important to
723 point out that the review of the literature has showed that studies tend to be concentrated in certain areas.
724 It is surprising the small number of studies available regarding Siberia and the Russian part of the boreal
725 forest. We encourage researchers to publish (in English, the international scientific language) more on

726 this area, since the relative importance of different drivers of the N cycle (e.g. N depositions, harvesting,
727 fire regime) may change in vast and less populated areas.

728

729 **ACKNOWLEDGMENTS**

730 This work was funded by the Natural Sciences and Engineering Research Council of Canada, the
731 Consortium Ouranos and the Consortium de Recherche sur la Forêt Boréale Commerciale

732 The authors thank B. Côté and M. Vincent for their critical suggestions on an initial version of the
733 manuscript and A. Garside for checking the English text. Thanks to J.-F. Boucher, M. Gelinias-Pouliot and
734 A. Turcotte for suggestions on the canopy uptake and N-deposition sections. We also want to thank two
735 anonymous reviewers for their comments which helped in greatly improving an earlier version of the
736 manuscript.

737

DEFINITIONS, ACRONYMS, ABBREVIATIONS

DIN:dissolved inorganic nitrogen, the inorganic N in the soil solution, whose main components are NO₃⁻ and NH₄⁺ ([61], [77])

DON: dissolved organic nitrogen, usually calculated by subtracting inorganic N (NH₄⁺ and NO₃⁻) from total dissolved N. ([33]. DON is a heterogeneous mixture of organic compounds that can be divided in two pools, one highly labile and another more recalcitrant. In boreal soils, free amino acids represent 10-20% of DON ([10]).

ECM:ectomycorrhizae, mycorrhizal fungi associated with trees forming sheathing mantles of fungal tissues over the exterior of the root surfaces and among the root cells ([1]).

ERM:ericoid mycorrhizae of fungi penetrating within the epidermal cells of roots. The mycelium does not extend widely beyond the individual roots but remains a few millimetres from the cortical cells ([120]).

L layer:litter layer of the organic soil horizon at the soil surface in forest floors, with slightly decomposed organic matter but still recognizable organic debris ([62], [197])

FH layers:fermentation-humification horizons on the top of the soil profile ([1]). Layers of the ectorganic soil horizon of forest floors with intermediate to high degree of organic matter decomposition. The original form of most plant and animal matter cannot be recognized with the naked eye ([62], [197]).

RUBISCOphotosynthetic enzyme associated with C fixation; ribulose 1,5-bisphosphate carboxylase ([198])

SOM:Soil organic matter mainly composed of dead and chemically transformed material of biological origin as well as living microorganisms biomass, ([11])

Throughfall:Incident precipitation that had interacted with the forest canopy before reaching the forest floor ([31]).

REFERENCES

1. Read DJ, Leake JR, and Perez-Moreno J. Mycorrhizal fungi as drivers of ecosystem processes in heathland and boreal forest biomes. *Can J Bot.* 2004; 82(8):1243-1263 DOI: 10.1139/b04-123.
2. Kielland K, McFarland JW, Ruess RW, and Olson K. Rapid cycling of organic nitrogen in taiga forest ecosystems. *Ecosys.* 2007; 10(3):360-368 DOI: 10.1007/s10021-007-9037-8.
3. Vitousek PM and Howarth RW. Nitrogen limitation on land and in the sea - How can it occur. *Biogeochemistry.* 1991; 13(2):87-115
4. Vitousek PM, Aber JD, Howarth RW, Likens GE, Matson PA, Schindler DW, et al. Human alteration of the global nitrogen cycle: Sources and consequences. *Ecol Appl.* 1997; 7(3):737-750
5. Vitousek PM and Farrington H. Nutrient limitation and soil development: Experimental test of a biogeochemical theory. *Biogeochemistry.* 1997; 37(1):63-75
6. Galloway JN, Dentener FJ, Capone DG, Boyer EW, Howarth RW, Seitzinger SP, et al. Nitrogen cycles: past, present, and future. *Biogeochemistry.* 2004; 70(2):153-226
7. Luo Y, Su B, Currie WS, Dukes JS, Finzi A, Hartwig U, et al. Progressive nitrogen limitation of ecosystem responses to rising atmospheric carbon dioxide. *Bioscience.* 2004; 54(8):731-739
8. Schlesinger WH. On the fate of anthropogenic nitrogen. *Proc Natl Acad Sci U S A.* 2009; 106(1):203-208 DOI: 10.1073/pnas.0810193105.
9. Perakis SS and Hedin LO. Nitrogen loss from unpolluted South American forests mainly via dissolved organic compounds. *Nature.* 2002; 415(6870):416-419
10. Neff JC, Chapin FS, and Vitousek PM. Breaks in the cycle: dissolved organic nitrogen in terrestrial ecosystems. *Front Ecol Environ.* 2003; 1(4):205-211
11. Jackson LE, Burger M, and Cavagnaro TR. Roots nitrogen transformations, and ecosystem services. *Annu Rev Plant Biol.* 2008; 59:341-363 DOI: 10.1146/annurev.arplant.59.032607.092932.
12. Sutton MA, Erisman JW, Dentener F, and Moller D. Ammonia in the environment: From ancient times to the present. *Environ Pollut.* 2008; 156(3):583-604 DOI: 10.1016/j.envpol.2008.03.013.
13. Gallet C and Lebreton P. Evolution of phenolic patterns in plants and associated litters and humus of a mountain forest ecosystem. *Soil Biol Biochem.* 1995; 27(2):157-165
14. Bending GD and Read DJ. Nitrogen mobilization from protein-polyphenol complex by ericoid and ectomycorrhizal fungi. *Soil Biol Biochem.* 1996; 28(12):1603-1612
15. Hattenschwiler S and Vitousek PM. The role of polyphenols in terrestrial ecosystem nutrient cycling. *Trends Ecol Evol.* 2000; 15(6):238-243
16. Lindahl BO, Taylor AFS, and Finlay RD. Defining nutritional constraints on carbon cycling in boreal forests - towards a less 'phytcentric' perspective. *Plant Soil.* 2002; 242(1):123-135
17. Ushio M, Miki T, and Kitayama K. Phenolic Control of Plant Nitrogen Acquisition through the Inhibition of Soil Microbial Decomposition Processes: A Plant-Microbe Competition Model. *Microbes Environ.* 2009; 24(2):180-187 DOI: 10.1264/jsme2.ME09107.
18. Gundale MJ, DeLuca TH, and Nordin A. Bryophytes attenuate anthropogenic nitrogen inputs in boreal forests. *Glob Chang Biol.* 2011; 17:2743-2753
19. DeLuca TH, Nilsson MC, and Zackrisson O. Nitrogen mineralization and phenol accumulation along a fire chronosequence in northern Sweden. *Oecologia.* 2002; 133(2):206-214 DOI: 10.1007/s00442-002-1025-2.
20. Busse MD. Suitability and use of the 15N-isotope dilution method to estimate nitrogen fixation by actinorhizal shrubs. *For Ecol Manage.* 2000; 136:85-95
21. Paul LR, Chapman BK, and Chanway CP. Nitrogen fixation associated with *Suillus tomentosus* tuberculate ectomycorrhizae on *Pinus contorta* var. *latifolia*. *Ann Bot.* 2007; 99(6):1101-1109 DOI: 10.1093/aob/mcm061.

22. Thornley JHM and Cannell MGR. Long-term effects of fire frequency on carbon storage and productivity of boreal forests: a modeling study. *Tree Physiol.* 2004; 24:765-773
23. Harden JW, Neff JC, Sandberg DV, Turetsky MR, Ottmar R, Gleixner G, et al. Chemistry of burning the forest floor during the FROSTFIRE experimental burn, interior Alaska, 1999. *Global Biogeochem Cycles.* 2004; 18 DOI: doi:10.1029/2003GB002194.
24. Brais S, Pare D, and Ouimet R. Impacts of wild fire severity and salvage harvesting on the nutrient balance of jack pine and black spruce boreal stands. *For Ecol Manage.* 2000; 137(1-3):231-243
25. Garbaye J. Helper bacteria - a new dimension to the mycorrhizal symbiosis. *New Phytol.* 1994; 128(2):197-210
26. Houle D, Gauthier SB, Paquet S, Planas D, and Warren A. Identification of two genera of N-2-fixing cyanobacteria growing on three feather moss species in boreal forests of Quebec, Canada. *Can J Bot.* 2006; 84(6):1025-1029 DOI: 10.1139/b06-059.
27. Zackrisson O, DeLuca TH, Gentili F, Sellstedt A, and Jaderlund A. Nitrogen fixation in mixed *Hylocomium splendens* moss communities. *Oecologia.* 2009; 160(2):309-319 DOI: 10.1007/s00442-009-1299-8.
28. Lagerstrom A, Nilsson MC, Zackrisson O, and Wardle DA. Ecosystem input of nitrogen through biological fixation in feather mosses during ecosystem retrogression. *Funct Ecol.* 2007; 21(6):1027-1033 DOI: 10.1111/j.1365-2435.2007.01331.x.
29. Vitousek PM, Cassman K, Cleveland C, Crews T, Field CB, Grimm NB, et al. Towards an ecological understanding of biological nitrogen fixation. *Biogeochemistry.* 2002; 57(1):1-45
30. Houle D, Paquin R, Camire C, Ouimet R, and Duchesne L. Response of the Lake Clair Watershed (Duchesnay, Quebec) to changes in precipitation chemistry (1988-1994). *Can J For Res.* 1997; 27(11):1813-1821
31. Houle D, Ouimet R, Paquin R, and Laflamme JG. Interactions of atmospheric deposition with a mixed hardwood and a coniferous forest canopy at the Lake Clair Watershed (Duchesnay, Quebec). *Can J For Res.* 1999; 29(12):1944-1957
32. Lovett GM and Lindberg SE. Atmospheric deposition and canopy interactions of nitrogen in forests. *Can J For Res.* 1993; 23(8):1603-1616
33. Duchesne L and Houle D. Base cation cycling in a pristine watershed of the Canadian boreal forest. *Biogeochemistry.* 2006; 78(2):195-216 DOI: 10.1007/s10533-005-4174-7.
34. Aber JD, Goodale CL, Ollinger SV, Smith ML, Magill AH, Martin ME, et al. Is nitrogen deposition altering the nitrogen status of northeastern forests? *Bioscience.* 2003; 53(4):375-389
35. Ouimet R and Duchesne L. Dépôts atmosphériques dans les forêts au Québec - Retombées actuelles et tendances au cours des 20 à 30 dernières années. *Le naturaliste canadien.* 2009; 133(1):56-64
36. Houle D, Duchesne L, and Boutin R. Effects of a spruce budworm outbreak on element export below the rooting zone: a case study for a balsam fir forest. *Ann For Sci.* 2009; 66(7) DOI: 10.1051/forest/2009057.
37. Lamontagne S, Carignan R, D'Arcy P, Prairie YT, and Pare D. Element export in runoff from eastern Canadian Boreal Shield drainage basins following forest harvesting and wildfires. *Can J Fish Aquat Sci.* 2000; 57(Suppl. 2):118-128
38. Prescott CE, Maynard DG, and Laiho R. Humus in northern forests: friend or foe? *For Ecol Manage.* 2000; 133(1-2):23-36
39. Turnbull MH, Schmidt S, Erskine PD, Richards S, and Stewart GR. Root adaptation and nitrogen source acquisition in natural ecosystems. *Tree Physiol.* 1996; 16(11-12):941-948
40. Galloway JN, Aber JD, Erisman JW, Seitzinger SP, Howarth RW, Cowling EB, et al. The nitrogen cascade. *Bioscience.* 2003; 53(4):341-356

41. Certini G. Effects of fire on properties of forest soils: a review. *Oecologia*. 2005; 143(1):1-10 DOI: 10.1007/s00442-004-1788-8.
42. Miller AJ and Cramer MD. Root nitrogen acquisition and assimilation. *Plant Soil*. 2004; 274(1-2):1-36 DOI: 10.1007/s11104-004-0965-1.
43. Hobbie EA and Ouimette AP. Controls of nitrogen isotope patterns in soil profiles. *Biogeochemistry*. 2009; 95(2-3):355-371 DOI: 10.1007/s10533-009-9328-6.
44. Matzner E and Borken W. Do freeze-thaw events enhance C and N losses from soils of different ecosystems? A review. *Eur J Soil Sci*. 2008; 59(2):274-284 DOI: 10.1111/j.1365-2389.2007.00992.x.
45. Rixen C, Freppaz M, Stoeckli V, Huovinen C, Huovinen K, and Wipf S. Altered snow density and chemistry change soil nitrogen mineralization and plant growth. *Arct Antarct Alp Res*. 2008; 40(3):568-575 DOI: 10.1657/1523-0430(07-044)[rixen]2.0.co;2.
46. Niboyet A, Barthes L, Hungate BA, Le Roux X, Bloor JMG, Ambroise A, et al. Responses of soil nitrogen cycling to the interactive effects of elevated CO₂ and inorganic N supply. *Plant Soil*. 2010; 327(1-2):35-47 DOI: 10.1007/s11104-009-0029-7.
47. Berg B and Dise N. Calculating the long-term stable nitrogen sink in northern European forests. *Acta Oecol - Int J Ecol*. 2004; 26(1):15-21 DOI: 10.1016/j.actao.2004.03.003.
48. Nambiar EKS and Fife DN. Nutrient retranslocation in temperate conifers. *Tree Physiol*. 1991; 9(1-2):185-207
49. Grayston SJ, Vaughan D, and Jones D. Rhizosphere carbon flow in trees, in comparison with annual plants: The importance of root exudation and its impact on microbial activity and nutrient availability. *Appl Soil Ecol*. 1996; 5(1):29-56
50. Ruess RW, Hendrick RL, Burton AJ, Pregitzer KS, Sveinbjornsson B, Allen ME, et al. Coupling fine root dynamics with ecosystem carbon cycling in black spruce forests of interior Alaska. *Ecol Monogr*. 2003; 73(4):643-662
51. Aerts R. Nutrient resorption from senescing leaves of perennials: Are there general patterns? *J Ecol*. 1996; 84(4):597-608
52. Wardle DA, Zackrisson O, Hornberg G, and Gallet C. The influence of island area on ecosystem properties. *Science*. 1997; 277(5330):1296-1299
53. Aerts R and Chapin FS. The mineral nutrition of wild plants revisited: A re-evaluation of processes and patterns. in *Advances in Ecological Research*, Vol 30, Academic Press Inc: San Diego. p. 1-67; 2000
54. Prescott CE. The influence of the forest canopy on nutrient cycling. *Tree Physiol*. 2002; 22(15-16):1193-1200
55. Campbell JL, Rustad LE, Boyer EW, Christopher SF, Driscoll CT, Fernandez IJ, et al. Consequences of climate change for biogeochemical cycling in forests of northeastern North America. *Can J For Res*. 2009; 39(2):264-284 DOI: 10.1139/x08-104.
56. Binkley D and Hogberg P. Does atmospheric deposition of nitrogen threaten Swedish forests? *For Ecol Manage*. 1997; 92(1-3):119-152
57. Schimel JP and Bennett J. Nitrogen mineralization: Challenges of a changing paradigm. *Ecology*. 2004; 85(3):591-602
58. Nasholm T, Ekblad A, Nordin A, Giesler R, Hogberg M, and Hogberg P. Boreal forest plants take up organic nitrogen. *Nature*. 1998; 392(6679):914-916
59. Paungfoo-Lonhienne C, Lonhienne TGA, Rentsch D, Robinson N, Christie M, Webb RI, et al. Plants can use protein as a nitrogen source without assistance from other organisms. *Proc Natl Acad Sci U S A*. 2008; 105(11):4524-4529 DOI: 10.1073/pnas.0712078105.

60. Gardenas AI, Agren GI, Bird JA, Clarholm M, Hallin S, Ineson P, et al. Knowledge gaps in soil carbon and nitrogen interactions - From molecular to global scale. *Soil Biol Biochem.* 2010; 10.1016/j.soilbio.2010.04.006:1-16 DOI: 10.1016/j.soilbio.2010.04.006.
61. Nasholm T, Kielland K, and Ganeteg U. Uptake of organic nitrogen by plants. *New Phytol.* 2009; 182(1):31-48 DOI: 10.1111/j.1469-8137.2008.02751.x.
62. Ste-Marie C and Houle D. Forest floor gross and net nitrogen mineralization in three forest types in Quebec, Canada. *Soil Biol Biochem.* 2006; 38(8):2135-2143 DOI: 10.1016/j.soilbio.2006.01.017.
63. Giesler R, Hogberg M, and Hogberg P. Soil chemistry and plants in Fennoscandian boreal forest as exemplified by a local gradient. *Ecology.* 1998; 79(1):119-137
64. Nordin A, Hogberg P, and Nasholm T. Soil nitrogen form and plant nitrogen uptake along a boreal forest productivity gradient. *Oecologia.* 2001; 129(1):125-132
65. Hogberg M, Myrold DD, Giesler R, and Hogberg P. Contrasting patterns of soil N-cycling in model ecosystems of Fennoscandian Boreal Forests. *Oecologia.* 2006; 147(1):96-107
66. Kranabetter JM, Dawson CR, and Dunn DE. Indices of dissolved organic nitrogen, ammonium and nitrate across productivity gradients of boreal forests. *Soil Biol Biochem.* 2008; 39:3147-3158
67. Smithwick EAH, Kashian DM, Ryan MG, and Turner MG. Long-Term Nitrogen Storage and Soil Nitrogen Availability in Post-Fire Lodgepole Pine Ecosystems. *Ecosys.* 2009; 12(5):792-806
68. Jerabkova L, Prescott CE, Titus BD, Hope GD, and Walters MB. A meta-analysis of the effects of clearcut and variable-retention harvesting on soil nitrogen fluxes in boreal and temperate forests. *Can J For Res.* 2011; 41:1852-1870
69. Werdin-Pfisterer NR, Kielland K, and Boone RD. Soil amino acid composition across a boreal forest successional sequence. *Soil Biol Biochem.* 2009; 41(6):1210-1220 DOI: 10.1016/j.soilbio.2009.03.001.
70. LeDuc SD and Rothstein DE. Plant-available organic and mineral nitrogen shift in dominance with forest stand age. *Ecology.* 2010; 91(3):708-720
71. Vitousek PM, Matson PA, and Van Cleve K. Nitrogen availability and nitrification during succession: Primary, secondary, and old-field seres. *Plant Soil.* 1989; 115:229-239
72. Britto DT and Kronzucker HJ. NH₄⁺ toxicity in higher plants: a critical review. *J Plant Physiol.* 2002; 159(6):567-584
73. Hyvonen R, Persson T, Andersson S, Olsson B, Agren GI, and Linder S. Impact of long-term nitrogen addition on carbon stocks in trees and soils in northern Europe. *Biogeochemistry.* 2008; 89(1):121-137 DOI: 10.1007/s10533-007-9121-3.
74. LeBauer DS and Treseder KK. Nitrogen limitation of net primary productivity in terrestrial ecosystems is globally distributed. *Ecology.* 2008; 89(2):371-379
75. de Vries W, Solberg S, Dobbertin M, Sterba H, Laubhann D, van Oijen M, et al. The impact of nitrogen deposition on carbon sequestration by European forests and heathlands. *For Ecol Manage.* 2009; 258(8):1814-1823 DOI: 10.1016/j.foreco.2009.02.034.
76. Jones DL, Healey JR, Willett VB, Farrar JF, and Hodge A. Dissolved organic nitrogen uptake by plants - an important N uptake pathway? *Soil Biol Biochem.* 2005; 37(3):413-423 DOI: 10.1016/j.soilbio.2004.08.008.
77. Houle D and Moore JD. Soil solution, foliar concentrations and tree growth response to 3-year of ammonium-nitrate addition in two boreal forests of Quebec, Canada. *For Ecol Manage.* 2008; 255(7):2049-2060 DOI: 10.1016/j.foreco.2007.10.056.
78. Vitousek PM, Hattenschwiler S, Olander L, and Allison S. Nitrogen and nature. *Ambio.* 2002; 31(2):97-101

79. Yamasaki SH, Fyles JW, Egger KN, and Titus BD. The effect of *Kalmia angustifolia* on the growth, nutrition, and ectomycorrhizal symbiont community of black spruce. *For Ecol Manage.* 1998; 105(1-3):197-207
80. Joannis GD, Bradley RL, Preston CM, and Bending GD. Sequestration of soil nitrogen as tannin-protein complexes may improve the competitive ability of sheep laurel (*Kalmia angustifolia*) relative to black spruce (*Picea mariana*). *New Phytol.* 2009; 181(1):187-198 DOI: 10.1111/j.1469-8137.2008.02622.x.
81. Jaderlund A, Zackrisson O, Dahlberg A, and Nilsson MC. Interference of *Vaccinium myrtillus* on establishment, growth, and nutrition of *Picea abies* seedlings in a northern boreal site. *Can J For Res.* 1997; 27(12):2017-2025
82. Imo M and Timmer VR. Vector competition analysis of black spruce seedling responses to nutrient loading and vegetation control. *Can J For Res.* 1999; 29(4):474-486
83. Menge DNL, Levin SA, and Hedin LO. Evolutionary tradeoffs can select against nitrogen fixation and thereby maintain nitrogen limitation. *Proc Natl Acad Sci U S A.* 2008; 105(5):1573-1578 DOI: 10.1073/pnas.0711411105.
84. Finzi AC and Rodgers VL. Bottom-up rather than top-down processes regulate the abundance and activity of nitrogen fixing plants in two Connecticut old-field ecosystems. *Biogeochemistry.* 2009; 95(2-3):309-321 DOI: 10.1007/s10533-009-9338-4.
85. D'Orangeville L, Houle D, Côté B, Duchesne L, and Morin H. Three years of increased soil temperature and atmospheric N deposition have no effect on the N status and growth of a mature balsam fir forest. *Biogeosci. Discuss.* 2013; 10:1313-1343 DOI: doi:10.5194/bgd-10-1313-2013.
86. Turner MG, Smithwick EAH, Tinker DB, and Romme WH. Variation in foliar nitrogen and aboveground net primary production in young postfire lodgepole pine. *Can J For Res.* 2009; 39(5):1024-1035 DOI: 10.1139/x09-029.
87. Kronzucker HJ, Siddiqi MY, and Glass ADM. Conifer root discrimination against soil nitrate and the ecology of forest succession. *Nature.* 1997; 385(6611):59-61
88. Grenon F, Bradley RL, Jones MD, Shipley B, and Peat H. Soil factors controlling mineral N uptake by *Picea engelmannii* seedlings: the importance of gross NH_4^+ production rates. *New Phytol.* 2005; 165(3):791-800 DOI: 10.1111/j.1469-8137.2004.01289.x.
89. Leake JR, Johnson D, Donnelly DP, Muckle GE, Boddy L, and Read DJ. Networks of power and influence: the role of mycorrhizal mycelium in controlling plant communities and agroecosystem functioning. *Can J Bot.* 2004; 82(8):1016-1045 DOI: 10.1139/b04-060.
90. Guo SW, Zhou Y, Gao YX, Li Y, and Shen QR. New insights into the nitrogen form effect on photosynthesis and photorespiration. *Pedosphere.* 2007; 17(5):601-610
91. Meyer A, Grote R, Polle A, and Butterbach-Bahl K. Simulating mycorrhiza contribution to forest C- and N cycling-the MYCOFON model. *Plant Soil.* 2010; 327(1-2):493-517 DOI: 10.1007/s11104-009-0017-y.
92. Kronzucker HJ, Siddiqi MY, Glass ADM, and Britto DT. Root ammonium transport efficiency as a determinant in forest colonization patterns: an hypothesis. *Physiol Plant.* 2003; 117(2):164-170
93. Hangs RD, Knight JD, and Van Rees KCJ. Nitrogen uptake characteristics for roots of conifer seedlings and common boreal forest competitor species. *Can J For Res.* 2003; 33(1):156-163 DOI: 10.1139/x02-169.
94. Rennenberg H, Schneider S, and Weber P. Analysis of uptake and allocation of nitrogen and sulphur compounds by trees in the field. *J Exp Bot.* 1996; 47(303):1491-1498
95. Min X, Siddiqi MY, Guy RD, Glass ADM, and Kronzucker HJ. A comparative study of fluxes and compartmentation of nitrate and ammonium in early-successional tree species. *Plant Cell Environ.* 1999; 22(7):821-830

997 96. Ohlund J and Nasholm T. Growth of conifer seedlings on organic and inorganic nitrogen sources.
998 Tree Physiol. 2001; 21(18):1319-1326
999 97. Nilsson MC and Wardle DA. Understory Vegetation as a Forest Ecosystem Driver: Evidence from
1000 the Northern Swedish Boreal Forest. Front Ecol Environ. 2005; 3(8):421-428
1001 98. Persson J, Hogberg P, Ekblad A, Hogberg MN, Nordgren A, and Nasholm T. Nitrogen acquisition
1002 from inorganic and organic sources by boreal forest plants in the field. Oecologia. 2003;
1003 137(2):252-257 DOI: 10.1007/s00442-003-1334-0.
1004 99. Ohlund J and Nasholm T. Regulation of organic and inorganic nitrogen uptake in Scots pine
1005 (*Pinus sylvestris*) seedlings. Tree Physiol. 2004; 24(12):1397-1402
1006 100. Persson J, Gardestrom P, and Nasholm T. Uptake, metabolism and distribution of organic and
1007 inorganic nitrogen sources by *Pinus sylvestris*. J Exp Bot. 2006; 57(11):2651-2659 DOI:
1008 10.1093/jxb/erl028.
1009 101. Ignatova N and Dambrine E. Canopy uptake of N deposition in spruce (*Picea abies* L Karst)
1010 stands. Ann For Sci. 2000; 57(2):113-120
1011 102. Nave LE, Vogel CS, Gough CM, and Curtis PS. Contribution of atmospheric nitrogen deposition to
1012 net primary productivity in a northern hardwood forest. Can J For Res. 2009; 39(6):1108-1118
1013 DOI: 10.1139/x09-038.
1014 103. Olson RK, Reiners WA, Cronan CS, and Lang GE. The chemistry and flux of throughfall and
1015 stemflow in subalpine balsam fir forests. Holarctic Ecology. 1981; 4(4):291-300
1016 104. Reiners WA and Olson RK. Effects of canopy components on throughfall chemistry - An
1017 experimental analysis. Oecologia. 1984; 63(3):320-330
1018 105. Lupi C, Morin H, Deslauriers A, Rossi S, and Houle D. Increasing nitrogen availability and soil
1019 temperature: effects on xylem phenology and anatomy of mature black spruce. Can J For Res.
1020 2012; 42(7):1277-1288
1021 106. Sparks JP. Ecological ramifications of the direct foliar uptake of nitrogen. Oecologia. 2009;
1022 159(1):1-13 DOI: 10.1007/s00442-008-1188-6.
1023 107. Dail DB, Hollinger DY, Davidson EA, Fernandez I, Sievering HC, Scott NA, et al. Distribution of
1024 nitrogen-15 tracers applied to the canopy of a mature spruce-hemlock stand, Howland, Maine,
1025 USA. Oecologia. 2009; 160(3):589-599 DOI: 10.1007/s00442-009-1325-x.
1026 108. Klopatek JM, Barry MJ, and Johnson DW. Potential canopy interception of nitrogen in the Pacific
1027 Northwest, USA. For Ecol Manage. 2006; 234(1-3):344-354 DOI: 10.1016/j.foreco.2006.07.019.
1028 109. DeLuca TH, Zackrisson O, Gentili F, Sellstedt A, and Nilsson MC. Ecosystem controls on nitrogen
1029 fixation in boreal feather moss communities. Oecologia. 2007; 152(1):121-130 DOI:
1030 10.1007/s00442-006-0626-6.
1031 110. Ericsson T. Growth and shoot-root ratio of seedlings in relation to nutrient availability. Plant Soil.
1032 1995; 168:205-214
1033 111. Treseder KK and Allen MF. Direct nitrogen and phosphorus limitation of arbuscular mycorrhizal
1034 fungi: a model and field test. New Phytol. 2002; 155(3):507-515
1035 112. Gill RA and Jackson RB. Global patterns of root turnover for terrestrial ecosystems. New Phytol.
1036 2000; 147(1):13-31
1037 113. De Nobili M, Contin M, Mondini C, and Brookes PC. Soil microbial biomass is triggered into
1038 activity by trace amounts of substrate. Soil Biol Biochem. 2001; 33(9):1163-1170
1039 114. Ekblad A and Nordgren A. Is growth of soil microorganisms in boreal forests limited by carbon or
1040 nitrogen availability? Plant Soil. 2002; 242(1):115-122
1041 115. Freppaz M, Williams BL, Edwards AC, Scalenghe R, and Zanini E. Labile nitrogen, carbon, and
1042 phosphorus pools and nitrogen mineralization and immobilization rates at low temperatures in
1043 seasonally snow-covered soils. Biol Fertil Soils. 2007; 43(5):519-529 DOI: 10.1007/s00374-006-
1044 0130-5.

1045 116. Lindahl BD, Ihrmark K, Boberg J, Trumbore SE, Hogberg P, Stenlid J, et al. Spatial separation of
1046 litter decomposition and mycorrhizal nitrogen uptake in a boreal forest. *New Phytol.* 2007;
1047 173(3):611-620 DOI: 10.1111/j.1469-8137.2006.01936.x.

1048 117. Kaye JP and Hart SC. Competition for nitrogen between plants and soil microorganisms. *Trends*
1049 *Ecol Evol.* 1997; 12(4):139-143

1050 118. Nasholm T and Persson J. Plant acquisition of organic nitrogen in boreal forests. *Physiol Plant.*
1051 2001; 111(4):419-426

1052 119. Zackrisson O, Nilsson MC, Dahlberg A, and Jaderlund A. Interference mechanisms in conifer-
1053 Ericaceae-feathermoss communities. *Oikos.* 1997; 78(2):209-220

1054 120. Read DJ. Mycorrhizas in ecosystems. *Experientia.* 1991; 47(4):376-391

1055 121. Read DJ and Perez-Moreno J. Mycorrhizas and nutrient cycling in ecosystems - a journey
1056 towards relevance? *New Phytol.* 2003; 157(3):475-492

1057 122. Hogberg MN, Chen Y, and Hogberg P. Gross nitrogen mineralisation and fungi-to-bacteria ratios
1058 are negatively correlated in boreal forests. *Biol Fertil Soils.* 2007; 44(2):363-366 DOI:
1059 10.1007/s00374-007-0215-9.

1060 123. Hogberg MN, Hogberg P, and Myrold DD. Is microbial community composition in boreal forest
1061 soils determined by pH, C-to-N ratio, the trees, or all three? *Oecologia.* 2007; 150(4):590-601
1062 DOI: 10.1007/s00442-006-0562-5.

1063 124. Nilsson LO and Wallander H. Production of external mycelium by ectomycorrhizal fungi in a
1064 Norway spruce forest was reduced in response to nitrogen fertilization. *New Phytol.* 2003;
1065 158(2):409-416 DOI: 10.1046/j.1469-8137.2003.00728.x.

1066 125. Yarwood SA, Myrold DD, and Hogberg MN. Termination of belowground C allocation by trees
1067 alters soil fungal and bacterial communities in a boreal forest. *FEMS Microbiol Ecol.* 2009;
1068 70(1):151-162 DOI: 10.1111/j.1574-6941.2009.00733.x.

1069 126. Schulze ED, Chapin FS, and Gebauer G. Nitrogen nutrition and isotope differences among life
1070 forms at the northern treeline of Alaska. *Oecologia.* 1994; 100(4):406-412

1071 127. Klironomos JN and Hart MM. Food-web dynamics - Animal nitrogen swap for plant carbon.
1072 *Nature.* 2001; 410(6829):651-652

1073 128. Davey ML and Currah RS. Interactions between mosses (Bryophyta) and fungi. *Can J Bot.* 2006;
1074 84(10):1509-1519 DOI: 10.1139/b06-120.

1075 129. Kauserud H, Mathiesen C, and Ohlson M. High diversity of fungi associated with living parts of
1076 boreal forest bryophytes. *Botany.* 2008; 86(11):1326-1333 DOI: 10.1139/b08-102.

1077 130. Gundale MJ, Gustafsson H, and Nilsson MC. The sensitivity of nitrogen fixation by a
1078 feathermoss-cyanobacteria association to litter and moisture variability in young and old boreal
1079 forests. *Can J For Res.* 2009; 39(12):2542-2549 DOI: 10.1139/x09-160.

1080 131. Nasholm T and Ericsson A. Seasonal changes in amino-acids, protein and total nitrogen in
1081 needles of fertilized Scots pine trees. *Tree Physiol.* 1990; 6(3):267-281

1082 132. Gezelius K. Free amino-acids and total nitrogen during shoot development in Scots pine
1083 seedlings. *Physiol Plant.* 1986; 67(3):435-441

1084 133. Millard P and Proe MF. Storage and internal cycling of nitrogen in relation to seasonal growth of
1085 Sitka spruce. *Tree Physiol.* 1992; 10(1):33-43

1086 134. Proe MF and Millard P. Relationships between nutrient supply, nitrogen partitioning and growth
1087 in young Sitka spruce (*Picea sitchensis*). *Tree Physiol.* 1994; 14(1):75-88

1088 135. Millard P. Ecophysiology of the internal cycling of nitrogen for tree growth. *J Plant Nutr Soil Sci.*
1089 1996; 159(1):1-10

1090 136. Malik V and Timmer VR. Biomass partitioning and nitrogen retranslocation in black spruce
1091 seedlings on competitive mixedwood sites: a bioassay study. *Can J For Res.* 1998; 28(2):206-215

- 1092 137. Reich PB, Oleksyn J, and Wright IJ. Leaf phosphorus influences the photosynthesis-nitrogen
1093 relation: a cross-biome analysis of 314 species. *Oecologia*. 2009; 160(2):207-212 DOI:
1094 10.1007/s00442-009-1291-3.
- 1095 138. Brix H. Effects of nitrogen fertilization on photosynthesis and respiration in Douglas-fir. *For Sci*.
1096 1971; 17(4):407-&
- 1097 139. Evans CA, Miller EK, and Friedland AJ. Effect of nitrogen and light on nutrient concentrations and
1098 associated physiological responses in birch and fir seedlings. *Plant Soil*. 2001; 236(2):197-207
- 1099 140. Gezelius K and Nasholm T. Free amino-acids and protein in Scots pine seedlings cultivated at
1100 different nutrient availabilities. *Tree Physiol*. 1993; 13(1):71-86
- 1101 141. Patterson TB, Guy RD, and Dang QL. Whole-plant nitrogen- and water-relations traits, and their
1102 associated trade-offs, in adjacent muskeg and upland boreal spruce species. *Oecologia*. 1997;
1103 110(2):160-168
- 1104 142. Reich PB, Walters MB, Tjoelker MG, Vanderklein D, and Buschena C. Photosynthesis and
1105 respiration rates depend on leaf and root morphology and nitrogen concentration in nine boreal
1106 tree species differing in relative growth rate. *Funct Ecol*. 1998; 12(3):395-405
- 1107 143. Lilles EB and Astrup R. Multiple resource limitation and ontogeny combined: a growth rate
1108 comparison of three co-occurring conifers. *Can J For Res*. 2012; 42:99-110
- 1109 144. Makinen H, Saranpaa P, and Linder S. Wood-density variation of Norway spruce in relation to
1110 nutrient optimization and fibre dimensions. *Can J For Res*. 2002; 32(2):185-194 DOI:
1111 10.1139/x01-186.
- 1112 145. Saren MP, Serimaa R, Andersson S, Saranpaa P, Keckes J, and Fratzl P. Effect of growth rate on
1113 mean microfibril angle and cross-sectional shape of tracheids of Norway spruce. *Trees*. 2004;
1114 18(3):354-362 DOI: 10.1007/s00468-003-0313-8.
- 1115 146. Meyer FD, Paulsen J, and Korner C. Windthrow damage in *Picea abies* is associated with physical
1116 and chemical stem wood properties. *Trees*. 2008; 22(4):463-473 DOI: 10.1007/s00468-007-
1117 0206-3.
- 1118 147. Canovas FM, Avila C, Canton FR, Canas RA, and de la Torre F. Ammonium assimilation and amino
1119 acid metabolism in conifers. *J Exp Bot*. 2007; 58(9):2307-2318 DOI: 10.1093/jxb/erm051.
- 1120 148. Reich PB, Grigal DF, Aber JD, and Gower ST. Nitrogen mineralization and productivity in 50
1121 hardwood and conifer stands on diverse soils. *Ecology*. 1997; 78(2):335-347
- 1122 149. Alcubilla M, Aufsess HV, and Rehfuess KE. Nitrogen-fertilization experiments in a Norway Spruce
1123 stand (*Picea abies* Karst.) of stagnant growth on devastated marly rendzina - effects on nutrient
1124 contents of spruce tissues and height increment. *Eur J For Res*. 1976; 95(5-6):306-323
- 1125 150. Kaakinen S, Saranpaa P, and Vapaavuori E. Effects of growth differences due to geographic
1126 location and N-fertilisation on wood chemistry of Norway spruce. *Trees*. 2007; 21(2):131-139
1127 DOI: 10.1007/s00468-006-0103-1.
- 1128 151. Anttonen S, Manninen AM, Saranpaa P, Kainulainen P, Linder S, and Vapaavuori E. Effects of
1129 long-term nutrient optimisation on stem wood chemistry in *Picea abies*. *Trees*. 2002; 16(6):386-
1130 394 DOI: 10.1007/s00468.002.0181.7.
- 1131 152. Kielland K, McFarland J, and Olson K. Amino acid uptake in deciduous and coniferous taiga
1132 ecosystems. *Plant Soil*. 2006; 288(1-2):297-307 DOI: 10.1007/s11104-006-9117-0.
- 1133 153. Thomas RQ, Canham CD, Weathers KC, and Goodale CL. Increased tree carbon storage in
1134 response to nitrogen deposition in the US. *Nat Geosci*. 2010; 3(1):13-17 DOI: 10.1038/ngeo721.
- 1135 154. Boisvenue C and Running SW. Impacts of climate change on natural forest productivity -
1136 evidence since the middle of the 20th century. *Glob Chang Biol*. 2006; 12(5):862-882 DOI:
1137 10.1111/j.1365-2486.2006.01134.x.

- 1138 155. Magnani F, Mencuccini M, Borghetti M, Berbigier P, Berninger F, Delzon S, et al. The human
1139 footprint in the carbon cycle of temperate and boreal forests. *Nature*. 2007; 447(7146):848-850
1140 DOI: 10.1038/nature05847.
- 1141 156. Gress SE, Nichols TD, Northcraft CC, and Peterjohn WT. Nutrient limitation in soils exhibiting
1142 differing nitrogen availabilities: What lies beyond nitrogen saturation? *Ecology*. 2007; 88(1):119-
1143 130
- 1144 157. Akselsson C, Westling O, Alveteg M, Thelin G, Fransson AM, and Hellsten S. The influence of N
1145 load and harvest intensity on the risk of P limitation in Swedish forest soils. *Sci Total Environ*.
1146 2008; 404(2-3):284-289 DOI: 10.1016/j.scitotenv.2007.11.017.
- 1147 158. Braun S, Thomas VFD, Quiring R, and Fluckiger W. Does nitrogen deposition increase forest
1148 production? The role of phosphorus. *Environ Pollut*. 2010; 158(6):2043-2052 DOI:
1149 10.1016/j.envpol.2009.11.030.
- 1150 159. Stromgren M and Linder S. Effects of nutrition and soil warming on stemwood production in a
1151 boreal Norway spruce stand. *Glob Chang Biol*. 2002; 8(12):1195-1204
- 1152 160. Olsson P, Linder S, Giesler R, and Hogberg P. Fertilization of boreal forest reduces both
1153 autotrophic and heterotrophic soil respiration. *Glob Chang Biol*. 2005; 11(10):1745-1753 DOI:
1154 10.1111/j.1365-2486.2005.001033.x.
- 1155 161. Knorr M, Frey SD, and Curtis PS. Nitrogen additions and litter decomposition: A meta-analysis.
1156 *Ecology*. 2005; 86(12):3252-3257
- 1157 162. Wallenda T and Kottke I. Nitrogen deposition and ectomycorrhizas. *New Phytol*. 1998;
1158 139(1):169-187
- 1159 163. Treseder KK. A meta-analysis of mycorrhizal responses to nitrogen, phosphorus, and
1160 atmospheric CO₂ in field studies. *New Phytol*. 2004; 164(2):347-355 DOI: 10.1111/j.1469-
1161 8137.2004.01159.x.
- 1162 164. Majdi H and Ohrvik J. Interactive effects of soil warming and fertilization on root production,
1163 mortality, and longevity in a Norway spruce stand in Northern Sweden. *Glob Chang Biol*. 2004;
1164 10(2):182-188 DOI: 10.1111/j.1529-8817.2003.00733.x.
- 1165 165. Avolio ML, Tuininga AR, Lewis JD, and Marchese M. Ectomycorrhizal responses to organic and
1166 inorganic nitrogen sources when associating with two host species. *Mycol Res*. 2009; 113:897-
1167 907 DOI: 10.1016/j.mycres.2009.05.001.
- 1168 166. Kranabetter JM, Durall DM, and MacKenzie WH. Diversity and species distribution of
1169 ectomycorrhizal fungi along productivity gradients of a southern boreal forest. . *Mycorrhiza*.
1170 2009; 19(2):99-111 DOI: 10.1007/s00572-008-0208-z.
- 1171 167. Rossi S, Bordeleau A, Houle D, and Morin H. Effect of chronic ammonium nitrate addition on the
1172 ectomycorrhizal community in a black spruce stand. *Can J Bot*. 2012; 42(7):1204-1212
- 1173 168. Clemmensen KE, Michelsen A, Jonasson S, and Shaver GR. Increased ectomycorrhizal fungal
1174 abundance after long-term fertilization and warming of two arctic tundra ecosystems. *New*
1175 *Phytol*. 2006; 171(2):391-404
- 1176 169. Prescott CE, Hope GD, and Blevins LL. Effect of gap size on litter decomposition and soil nitrate
1177 concentrations in a high-elevation spruce-fir forest. *Can J For Res*. 2003; 33:2210-2220
- 1178 170. LeDuc SD and Rothstein DE. Initial recovery of soil carbon and nitrogen pools and dynamics
1179 following disturbance in jack pine forests: A comparison of wildfire and clearcut harvesting. *Soil*
1180 *Biol Biochem*. 2007; 39:2865-2876
- 1181 171. Wan S, Hui D, and Luo Y. Fire effects on nitrogen pools and dynamics in terrestrial ecosystems: a
1182 meta-analysis. *Ecol Appl*. 2001; 11(5):1349-1365
- 1183 172. Yermakov Z and Rothstein DE. Changes in soil carbon and nitrogen cycling along a 72-year
1184 wildfire chronosequence in Michigan jack pine forests. *Oecologia*. 2006; 149:690-700

- 1185 173. Smithwick EAH, Ryan MG, Kashian DM, Romme WH, Tinker DB, and Turner MG. Modeling the
1186 effects of fire and climate change on carbon and nitrogen storage in lodgepole pine (*Pinus*
1187 *contorta*) stands. Glob Chang Biol. 2009; 15:535-548
- 1188 174. Rustad LE, Campbell JL, Marion GM, Norby RJ, Mitchell MJ, Hartley AE, et al. A meta-analysis of
1189 the response of soil respiration, net nitrogen mineralization, and aboveground plant growth to
1190 experimental ecosystem warming. Oecologia. 2001; 126(4):543-562
- 1191 175. Driscoll KG, Arocena JM, and Massicotte HB. Post-fire soil nitrogen content and vegetation
1192 composition in Sub-Boreal spruce forests of British Columbia's central interior, Canada. For Ecol
1193 Manage. 1999; 121:227-237
- 1194 176. Chen J, Chen WJ, Liu J, Cihlar J, and Gray S. Annual carbon balance of Canada's forests during
1195 1895-1996. Global Biogeochem Cycles. 2000; 14:839-849 DOI: doi:10.1029/1999GB01207.
- 1196 177. Kurz WA, Stinson G, and Rampley G. Could increased boreal forest ecosystem productivity offset
1197 carbon losses from increased disturbances? Phil. Trans. R. Soc. B. 2008; 363:2261-2269 DOI:
1198 doi:10.1098/rstb.2007.2198.
- 1199 178. Flannigan MD, Logan KA, Amiro BD, Skinner WR, and Stocks BJ. Future area burned in Canada.
1200 Clim. Change. 2005; 72(1-2):1-16 DOI: doi:10.1007/s10584-005-5935-y.
- 1201 179. Talbot JM and Treseder KK. Controls over mycorrhizal uptake of organic nitrogen. Pedobiologia.
1202 2010; 53:169-179 DOI: doi:10.1016/j.pedobi.2009.12.001.
- 1203 180. Schimel JP and Weintraub MN. The implications of exoenzyme activity on microbial carbon and
1204 nitrogen limitation in soil: a theoretical model. Soil Biol Biochem. 2003; 35(4):549-563 DOI:
1205 10.1016/s0038-0717(03)00015-4.
- 1206 181. Chalot M and Passard C. Ectomycorrhiza and nitrogen provision to the host tree. in Ecological
1207 aspects of nitrogen metabolism in plants, Polacco JC and Todd CD, Editors, John Wiley & Sons;
1208 2011
- 1209 182. Hobbie EA and Hobbie JE. Natural abundance of N-15 in nitrogen-limited forests and tundra can
1210 estimate nitrogen cycling through mycorrhizal fungi: A review. Ecosys. 2008; 11(5):815-830 DOI:
1211 10.1007/s10021-008-9159-7.
- 1212 183. Alberton O, Kuyper TW, and Gorissen A. Competition for nitrogen between *Pinus sylvestris* and
1213 ectomycorrhizal fungi generates potential for negative feedback under elevated CO₂. Plant Soil.
1214 2007; 296(1-2):159-172 DOI: 10.1007/s11104-007-9306-5.
- 1215 184. Aerts R. The advantages of being evergreen. Trends Ecol Evol. 1995; 10(10):402-407
- 1216 185. Meerts P. Mineral nutrient concentrations in sapwood and heartwood: a literature review. Ann
1217 For Sci. 2002; 59(7):713-722 DOI: 10.1051/forest:2002059.
- 1218 186. Reich PB, Wright IJ, Cavender-Bares J, Craine JM, Oleksyn J, Westoby M, et al. The evolution of
1219 plant functional variation: Traits, spectra, and strategies. Int J Plant Sci. 2003; 164(3):S143-S164
- 1220 187. Warren CR and Adams MA. Evergreen trees do not maximize instantaneous photosynthesis.
1221 Trends Plant Sci. 2004; 9(6):270-274
- 1222 188. Hikosaka K. Interspecific difference in the photosynthesis-nitrogen relationship: patterns,
1223 physiological causes, and ecological importance. J Plant Res. 2004; 117(6):481-494 DOI:
1224 10.1007/s10265-004-0174-2.
- 1225 189. Coates KD, Lilles EB, and Astrup R. Competitive interactions across a soil fertility gradient in a
1226 multispecies forest. J Ecol. 2013; 101:806-818 DOI: doi:10.1111/1365-2745.12072.
- 1227 190. Grantz DA, Garner JHB, and Johnson DW. Ecological effects of particulate matter. Env Int. 2003;
1228 29(2-3):213-239 DOI: 10.1016/s0160-4120(02)00181-2.
- 1229 191. Duchesne L and Houle D. Impact of nutrient removal through harvesting on the sustainability of
1230 the boreal forest. Ecol Appl. 2008; 18(7):1642-1651
- 1231 192. Matson P, Lohse KA, and Hall SJ. The globalization of nitrogen deposition: Consequences for
1232 terrestrial ecosystems. Ambio. 2002; 31(2):113-119

1233 193. Thiffault E, Pare D, Belanger N, Munson A, and Marquis F. Harvesting intensity at clear-felling in
1234 the boreal forest: Impact on soil and foliar nutrient status. *Soil Sci Soc Am J.* 2006; 70(2):691-701
1235 DOI: 10.2136/sssaj2005.0155.

1236 194. Litton CM, Ryan MG, and Knight DH. Effects of tree density and stand age on carbon allocation
1237 patterns in postfire lodgepole pine. *Ecol Appl.* 2004; 14:460-475

1238 195. Rossi S, Tremblay M-J, Morin H, and Savard G. Growth and productivity of black spruce in even-
1239 and uneven-aged stands at the limit of the closed boreal forest. *For Ecol Manage.* 2009;
1240 258:2153-2161

1241 196. Messier C, Doucet R, Ruel JC, Claveau Y, Kelly C, and Lechowicz MJ. Functional ecology of
1242 advance regeneration in relation to light in boreal forests. *Can J For Res.* 1999; 29:812-823

1243 197. Chesworth W. Encyclopedia of Soil Science. in *Encyclopedia of Soil Science*, Springer, Editor
1244 2008: Dordrecht, Netherlands.

1245 198. Brown KR, Thompson WA, Camm EL, Hawkins BJ, and Guy RD. Effects of N addition rates on the
1246 productivity of *Picea sitchensis*, *Thuja plicata*, and *Tsuga heterophylla* seedlings .2.
1247 Photosynthesis, ¹³C discrimination and N partitioning in foliage. *Trees.* 1996; 10(3):198-205

1248

1249