

## Original Research Article

# Metalaxyl induced changes in protein metabolism during germination of Maize

## Abstract

The present work was carried out to investigate the effect of fungicide metalaxyl as a seed treatment on the protein metabolism of maize during early germination. The study was carried out for 7 days after soaking the seeds in different concentrations of metalaxyl and a control was maintained. Biochemical analyses of whole seedlings were done up to 7 days. Changes in the amount of total soluble protein, total free amino acids, and the protease activity were performed using standard methods. The results indicate that protein content was maximum on 4<sup>th</sup> and 6<sup>th</sup> day of germination in both control and treated seeds. Metalaxyl treatment resulted in the decreased protein content (20-50% inhibition) over the untreated in a dose dependent manner till the 4<sup>th</sup> day of germination. But as the growth proceeded a proportionate increase in the protein content was observed on the 6<sup>th</sup> day of germination in fungicide treated seeds compared to the control. The specific activity of protease was decreased by 46%, 81%, 88%, 97%, and 100% with 1.5mg, 3mg, 4.5mg, 6mg and 7mg concentration of metalaxyl on 3<sup>rd</sup> day of germination when compared to the control. An increase in the total free amino acids occurred during the germination and maximum free amino acids content was observed on the 5<sup>th</sup> day. Metalaxyl treatment resulted in the dose dependent depletion of free amino acids content. A significant increase in proline content was found to occur in treated seeds. From the overall findings, the present study gives an insight into a protective effect of the system with an increased production of proline and decreased protease activity, free amino acids and at the same time a higher protein content during later stages of germination for a particular concentration which may be due to the synthesis of novel proteins as a defense mechanism indicating the dual role of metalaxyl.

**Keywords:** *Metalaxyl; germination; protease; proteins; free amino acids; proline.*

## 1. INTRODUCTION

The mobilization of seed storage proteins represents one of the most important post-germination events in the growth and development of seedling. Proteolytic enzymes play a central role in the biochemical mechanism of germination [1]. During germination period, the storage proteins are degraded by a variety of proteases which convert the insoluble storage proteins in to soluble peptides and these peptides are further hydrolyzed to free amino acids. These free amino acids are mobilized to the embryonic axis to support its growth and also to provide energy [2]. Downey mildew of maize (*Zea mays.L*) caused by *Peronosclerospora* is one of the most destructive diseases of this crop in the tropical Asian countries [3]. Integrated approaches are used to manage the disease including crop rotation, planting resistant cultivars and the application of fungicides like mancozeb and metalaxyl. Metalaxyl (methyl N-(methoxyacetyl)-N-(2, 6-xyllyl)-DL-alaninate) compounds have been widely used for the control of Downey mildew in a number of crops. Metalaxyl

is a systemic fungicide acts by suppressing sporangial formation, mycelia growth and establishment of new infection [4]. Much of our knowledge of reserve mobilization and its control processes during germination is very well understood. In contrast, available information on metalaxyl is limited to its effect on the pathogen and not much data is available on the effect of metalaxyl in protein metabolism of maize seeds. Hence, the present work is an attempt for furthering our knowledge towards better understanding of the effect of metalaxyl in germinating maize seeds focusing on the protein metabolic changes during germination.

## 2. MATERIAL AND METHODS

### 2.1 Collection of seeds and Treatments

Maize seeds were procured from VC farm, University of Agriculture Science, Mandya, Karnataka. All the chemicals were purchased from SLR and MERCK and the chemicals were of analytical grade. Seeds were surface sterilized with 0.1% mercuric chloride for 10 minutes and repeatedly washed with distilled water for 4-5 times. Seeds of uniform size were selected and soaked for 24 hours in distilled water (control) and with different concentrations (mg/g) of metalaxyl, 1.5, 3, 4.5, 6 and 7 mg/gm of the seeds (1:5 weight/volume) for 24 hours. Five seeds in triplicate were placed on petridish with 8-10 layer of soaked filter paper and incubated at 25 °C both in light and dark condition. Uniform seedlings were selected and processed for further studies. Everyday filter paper was wetted with 10ml of distilled water. [5]

### 2.2 Preparation of crude extract

Around 1-2 gram of seedling treated with different concentrations of the metalaxyl and the untreated seeds were taken each day, (up to 7 days) homogenized in ice cold saline (5ml) using pestle and mortar. The solution was centrifuged for 10 minutes and supernatant were used for further analysis.

### 2.3 Biochemical studies

Protein was estimated as described by Lowry et al. [6] using BSA as standard. The activity of protease was determined following the procedure of Kunitz [7]. Total free amino acids were extracted and determined following the method of Sugano et al [8]. Free proline content was estimated following the method of Bates et al [9].

### 2.4 Statistical Analysis

The data are expressed as the mean  $\pm$  SEM analyzed by one-way analysis of variance (ANOVA) and Dunnett's *t*-test was used as the test of significance. P value < 0.05 was considered as the minimum level of significance. All statistical tests were carried out using SPSS statistical software.

## 3. RESULTS AND DISCUSSION

### 3.1 Total Protein

The protein content during different days of germination in control and fungicide treated seedlings is presented in Table-1. The protein content was maximum on 4<sup>th</sup> and 6<sup>th</sup> day of germination in both control and treated seeds. Metalaxyl treatment resulted in the decreased protein content (20-50% inhibition) over the untreated in a dose dependent manner till the 4<sup>th</sup> day of germination. The treated seeds showed similar pattern of protein content on different days of germination. However an increase in the protein content was seen on the 6<sup>th</sup> day of germination in treated seeds.

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96 **Table 1- Effect of metalaxyl on the Total Protein (mg/g) in the seedlings of Maize.**

Concentration of metalaxyl (mg/g)	Seedling age in days							
	0	1	2	3	4	5	6	7
<b>Control</b>	12.6 ±0.9	24.2±1	26.69±1. 4	10.75±0 .6	42.48±2. 1	29.52±1. 4	57±3.1	50.65±1
<b>1.5</b>	10.7±0. 7	22.82±1	24.45±1. 5	9.21±0. 3	34.09±1. 4	21.54±1. 2	76.4±2	60.49±1. 9
<b>3.0</b>	13.1±0. 5	25.43±1. 2	25.91±1. 2	9.71±0. 4	30.8±1.2	33.15±1. 5	96.06±2.4	31.96±1. 1
<b>4.5</b>	34.4±1. 2	27.61±1	25.69±0. 4	2.71±0. 1	26.78±0. 9	33.69±1. 6	67.51±2.3	52.4±2.1
<b>6.0</b>	12.6±1	25.18±1. 1	12.43±0. 3	12.14±0 .6	20.16±0. 5	17.87±0. 9	63.19±2.3	22.33±0. 8
<b>7.0</b>	10±0.5	21.12±0. 9	23.15±1. 1	7.65±0. 3	25.45±0. 7	15.91±0. 7	47.52±1.2	31.21±0. 9

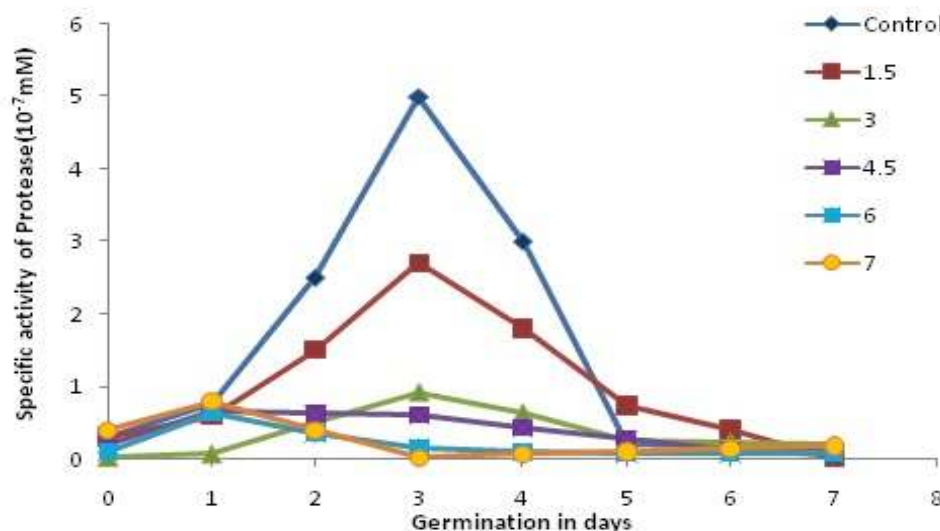
97 In the present study the protein content in germinated seeds were found to be increased ,due to the  
 98 mobilization of storage nitrogen for the production of protein needed for the development of the young  
 99 seedling and the observation was well in agreement with the studies conducted in germinated seeds  
 100 [10]. A decline in the protein content was observed in treated seeds till the 4<sup>th</sup> day in a dose dependent  
 101 fashion. The decrease in the protein content in fungicide treated maize may be due to osmotic shock  
 102 effect by the fungicides which results in the release of protein and loss of membrane transport ability [11].  
 103 It has been suggested that the toxicant produced in the treated seeds may inhibit the protein synthesis by  
 104 binding to the larger ribosomal subunit inducing change in the enzyme system ceasing ATP and NADP  
 105 formation .The results of our studies were in parallel, as the concentration of fungicide increases, the  
 106 amount of protein gradually decreased. But as the growth proceeded a proportionate increase in the  
 107 protein content was observed on the 6<sup>th</sup> day of germination in fungicide treated seeds compared to the  
 108 control. The enhanced protein content may be attributed to the fact that as growth proceeds, Metalaxyl  
 109 may induce the expression of many defense related genes in plants which results in the synthesis of  
 110 novel proteins. The results of our study correlates with the recent proteomic analysis which revealed the  
 111 occurrence of novel proteins in several plant species under heavy metal stress [12].

### 112 **3.2 Activity of Protease**

113 Protease is a hydrolytic enzyme which acts on proteinaceous substances to produce amino acids and  
 114 amides. The specific activities of protease during germination of maize seeds treated with Metalaxyl are  
 115 shown in Fig 1. Both control and treated seeds showed increasing specific activity up to 3<sup>rd</sup> day of  
 116 germination and decreased then onwards. Specific activity of protease in maize seedling at different  
 117 concentration of Metalaxyl on 3<sup>rd</sup> day were found to be 27, 9.1, 6, 1.6 and 0.3 ×10<sup>-7</sup> mM whereas in control

the activity is found to be  $50 \times 10^{-7}$  mM . The specific activity was decreased by 46%, 81%, 88%, 97%, and 100% with 1.5mg, 3mg, 4.5mg, 6mg and 7mg concentration of Metalaxyl on 3<sup>rd</sup> day of germination when compared to the control.

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126 Fig.1a. Effect of metalaxyl on the <sup>\*</sup> Specific activity of protease in the seedlings of Maize

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128 <sup>\*</sup>Specific activity of Protease is expressed as  $10^{-7}$  mM of amino acids released/mg of protein/min

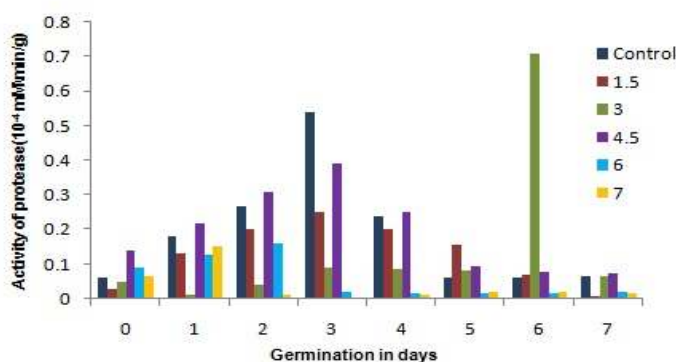


Fig.1b. Effect of metalaxyl on the activity of protease in the seedlings of Maize

129 Increasing in proteolytic activity with concomitant reserve protein depletion agrees with the findings of

130 earlier studies on other seeds; *Phaseolus vulgaris* [13], *Lupinus albus* [14] *Vicia sativa* [15] and

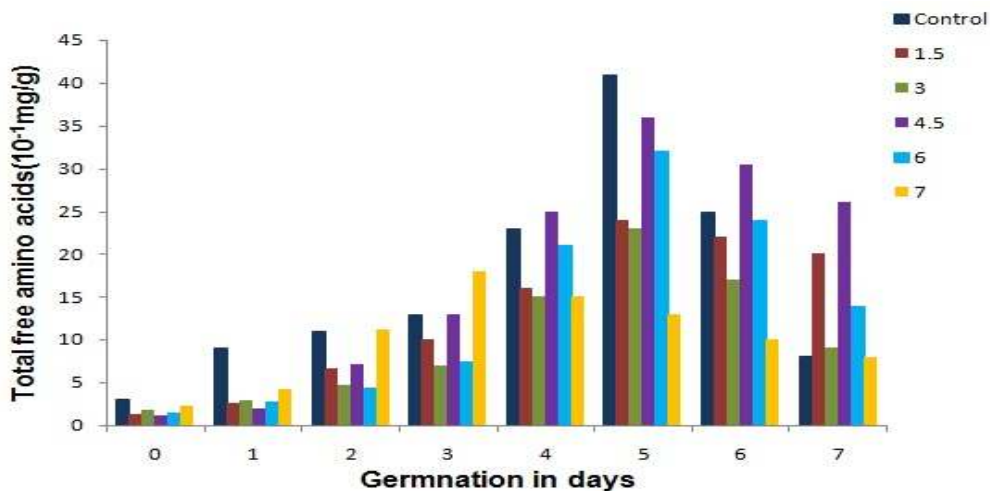
131 *Macrotyloma uniflorum* [16].The result from our study is not parallel with the above data as there is a dose

132 dependent inhibition of protease activity in metalaxyl treated maize seeds on 3<sup>rd</sup> day of germination.

133 Impairment of proteasome functionality and decreased protease activities seems to be a common  
feature  
134 involved in metal toxicity in plants [17].

### 135 3.3 Free amino acid

136 The amino acid content in control and treated seeds during different days of germination is  
presented in  
137 the Fig.2. A gradual increase in the amino acid content is seen and the maximum content of  
amino acid  
138 ( $41 \times 10^{-1}$  mg/g ) was seen on the 5<sup>th</sup> day in control seed and a gradual decrease was seen till  
the 7<sup>th</sup> day  
139 of germination. The free amino acid content was decreased effectively with the increasing  
concentration of fungicides with the maximum inhibition was seen with the highest concentration  
(7mg/g). Only 30% of  
140 free amino acids were present in that concentration. An increase in the total free amino acids  
occurred  
141 during the germination and maximum free amino acids content was observed on the 5<sup>th</sup> day.  
142 Metalaxyl treatment resulted in the dose dependent depletion of free amino acids content. There  
was no  
143 change in the pattern of amino acid profile on different days of germination in both treated and  
control  
144 seeds. These result suggests, the protective effect of amino acids against unfavorable condition,  
145 increasing the tendency of the system to maintain the homeostasis.



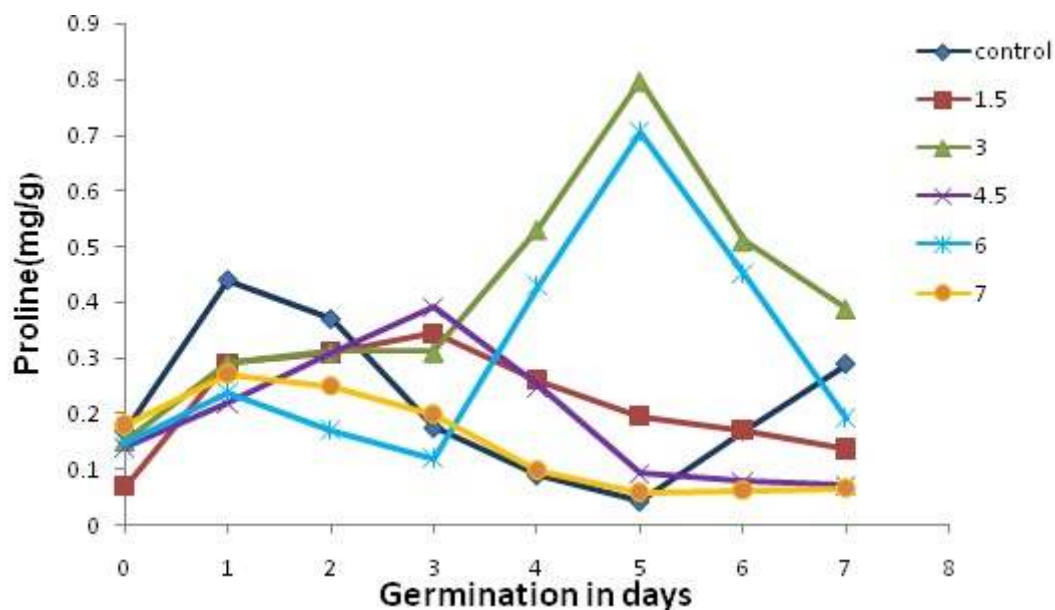
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149 **Fig.2. Effect of Metalaxyl on the Amino acid content ( $10^{-1}$  mg/g) in the seedlings of Maize**

## 150 3.4 Proline content

151 Proline content in control and treated seeds are presented in the Fig.3. The proline content increased in  
 152 treated seeds and maximum proline content was seen with 3mg/g and in 6mg/g concentration of  
 153 metalaxyl (0.797mg/g, 0.705mg/g) on 5<sup>th</sup> day of germination and decreased thereafter. The proline  
 154 content was lowest in control (0.043mg/g) on 5<sup>th</sup> day and in treated (7mg/g) and it was 0.06mg/g  
 155 on 7<sup>th</sup> day. A significant increase in proline content was found to occur in treated seeds.



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158 **Fig.3. Effect of Metalaxyl on the Proline content (mg/g) in the seedlings of Maize**

159 Proline is considered as an antioxidant free radical scavenging and biochemical indicator of  
 stress. It is  
 160 considered as a metabolic measures of abiotic stress [18]. In the present work the stress caused  
 by  
 161 metalaxyl resulted in an increase in proline levels. In general water stress has been known to  
 increase

162 free proline in leaves[19]. Plant growing in stress condition need to produce specific proteins having higher  
 163 proline content (hydroxyproline rich glycopeptide or proline rich glycopeptides) [20]. The high content of  
 164 free proline is also observed in different cultivars of wheat under salt stress[21]. Effect of this change is  
 165 subsequently contributed to higher production of glutamic acid which stressed plants need  
 166 to create phytochelatin[22].

## 166 4. CONCLUSION

167 The present study gives an insight into a protective effect of the system with an increased production of  
 proline and decreased protease activity, free amino acids and at the same time a higher protein content  
 during later stages of germination for a particular concentration may be due to the synthesis of novel  
 proteins as a defense mechanism indicating the dual role of metalaxyl. Further work is in progress in  
 investigating the role of increased protein content in maize during germination in treated. This may throw  
 light on understanding the signaling molecules its mechanism and regulation.

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## 180 REFERENCES

- 181 1. Muntz K, Belozersky MA, Dunaevsky YE, Schlereth A, Tiedemann J. Stored proteinases and  
 182 the initiation of storage protein mobilization in seeds during germination and seedling growth.  
 183 *Journal of Experimental Botany*. 2001; 52:1741–1752.
- 184 2. Bewley JD, Black M. *Physiology and Biochemistry of Seeds in Relation to Germination*. Vol. I.  
 185 *Springer – Verlag*, Berlin. 1983.
- 186 3. Bonde MR. Epidemiology of downy mildew diseases of maize, sorghum and pearl millet. *Tropical*  
 187 *Pest Management*. 1982; 28(1):49-60.
- 188 4. Fisher DJ, Hayes AL. Mode of action of the systemic fungicides furalaxyl, metalaxyl and ofurace.  
 189 *Pestic. Sci*. 1982; 13:330–339. DOI: 10.1002/ps.2780130316
- 190 5. ISTA. International rules for seed testing. In: Daper SR.(Ed), Rules 2003. International seed  
 191 testing association, Zurich, Switzerland, 1-520
- 192 6. Lowry, O.H., Rosenbrought, N.J., Farr, F.L. and Randall, R.J. 1951. Protein measurement with  
 Folin phenol reagent. *J. Biol. Chem*. 193: 265-275.
7. Kunitz, M, 1947 isolation of a crystalline protein compound of trypsin and of soybean trypsin-  
 Inhibitor. *J. Gen. Physiol.*, 30(4), 311-320.
8. Sugano N, Tanaka T, Yamamoto E, Nishi A (1975) Phenylalanine ammonia lyase in carrot cells  
 in suspension cultures. *Phytochemistry* 14:2435–2440
- 182 9. Bates, L.S., Waldran, R.P. and Teare, I.D. 1973. Rapid determination of free proline for water-  
 183 stress studies. *Plant Soil* 39: 205-207.
- 184 10. Gernah DI, Ariahe CC, Ingbian EK. Effects of malting and lactic acid fermentation on some  
 185 chemicals and functional properties of maize. *Am.J.Food. Technol*. 2011;6:404-412
- 186 11. Amar L, Reinhold L. Loss of membrane transport ability in leaf cells and release of protein as a  
 187 result of an osmotic shock. *Plant Physiol*. 1973;51:620-625



12. Elisabetta Gianazza, Robin Wait, Andrea Sozzi, Simona Regondi, Dolores Sac, Massimo Labra and Elisabetta Agradi. Growth and protein profile changes in *Lepidium sativum* L. plantlets exposed to cadmium. *Environmental and Experimental Botany*. 2007; 59:179–187.

13. Taneyama M, Okamoto T, Yamauchi D, Minamikawa T. Development of endopeptidase activity in cotyledons of *Vigna mungo* seedling: Effects of exogenously applied end products and plant hormones. *Plant Cell and Physiology*. 1996; 37:19–26.

14. Ferreira RB, Malo TS, Teixeira AN. Catabolism of the seed storage proteins from *Lupinus albus*: Fate of globulins during germination and seedling growth. *Australian Journal of Plant Physiology*. 1995; 22:373–381.

15. Schlereth A, Standhardt D, Mock HP, Muntz K. Stored cysteine proteinases start globulin breakdown in protein bodies of embryonic axis and cotyledons of germinating vetch (*Vicia sativa* L.) seeds. *Planta*, 2001; 212:718–727.

16. Rajeswari J, Ramakrishna Rao P. Storage protein degradation in germinating horse gram seeds. *Indian J Plant Physiol*. 2002; 7:314–320.

17. Wang C, Tian Y, Wang X, Geng J, Jiang J, Yu H, Wang C. Lead-contaminated soil induced oxidative stress, defense response and its indicative biomarkers in roots of *Vicia faba* seedlings. *Ecotoxicology*. 2010 Aug; 19(6):1130–9.

18. Andurwulan N, Fardiaz D, Wattimenia GA, Setty K. Antioxidant activity associated with lipid and phenolic mobilization during seed germination of *Pangium edule Reinw*. *J Agric Food Chem*. 1999; 47:3158–63.

19. Stewart CK, Lee JA. The role of proline accumulation in halophytes. *Planta* 1974; 120:1279–289.

20. Ueda A, Yamamoto-Yamane Y, Takabe T. Salt stress enhances proline utilization in the apical region of barley roots. *Biochemical and Biophysical Research Communication*. 2007; 355:61–66.

21. Poustini K, Siosemardeh A, Ranjbar M (2007). Proline accumulation as a response to salt stress in 30 wheat (*Triticum aestivum* L.) cultivars differing in salt tolerance, *Genet. Resour. Crop. Evol.* 54(5): 925–934.

22. Pavlíková D, Pavlík M, Staszková L, Motyka V, Száková J, Tlustoš P, Balík J. Glutamate kinase as a potential biomarker of heavy metal stress in plants. *Ecotoxicology and Environmental Safety*. 2008; 70:223–230.