Foliar selenium application to improve the tolerance of eggplant grown under salinity stress conditions

Authors' contributions

Both researchers have contributed in all parts of research equally.

ABSTRACT

8 Sea level rise is one of the most risky climate change impacts under Egyptian conditions to increase the salinity of 9 northern Delta. Increasing the tolerance for salinity in current and future crops is strongly desirable. The current experiment was carried out in the experimental station at Agriculture Research Center, Egypt, during the summer seasons of 2014 and 2015, to evaluate the effect of selenium foliar applications (0, 5, 10, 20, 30 µM Na₂SeO₃) on eggplant grown on a sandy soil and irrigated with different concentrations of saline water (0, 30, 60, 120 mM NaCl). The results showed that the Se supplement with 20 µM showed the best effects on vegetative growth and yield of eggplants under different salinity levels of irrigation water. Increasing salinity resulted in increasing N and P contents in the leaves and fruits of eggplant, but K decreased as a result of some sort of antagonism with Na; in spite of that, N, P and K contents in leaves and fruits increased with increasing Se supplements up to 20 µM to be at higher concentrations then decreased. Regarding K/Na ratio in leaves, the treatment of EC_e 13.5 dS m⁻¹ without Se supplements gave the lowest value (0.52); treatment of Se 30 µM under 0 mM NaCl irrigation water gave the highest one (1.71). Also, chlorophyll contents in plant leaves increased with increasing salinity level of irrigation water, but decreased with increasing Se supplements. Regarding the proline contents in fresh leaves, the treatment of EC_e 13.5 dS m⁻¹ without Se supplements gave the highest value (51 mg g^{-1}), but the lowest one (30.9 mg g^{-1}) was observed with control treatment.

Keywords: Sea level rise, Salinity stress, Selenium supplements, Eggplant, proline content.

1. INTRODUCTION

Egypt is very dependent on natural resources that are vulnerable to climate change. A large portion of the arable land in Nile Delta is particularly exposed to sea level rise. <u>Nicholls et al.</u> [1] <u>estimated</u> a mean value of 1 meter global sea level rise by the year of 2100 which would give rise to a 0.37 meter sea level rise at the Nile delta. This, combined with a non climate induced subsidence of the Nile Delta of 0.38 meters would result in the movement of the shoreline to the current 0.75 meter contour and a 5 percent loss of Egyptian agricultural land by the year of 2060 mainly at the coastal area of Nile Delta. <u>El-Raey et al.</u> [2] <u>suggested</u> land losses of 12 to 15 percent of Egypt's current arable land for a one meter sea level rise. Due to salinization and seawater intrusion, the agricultural activities will be difficult below an elevation of 1 meter.

Salinity is an abiotic stress to limit plant growth [3], and is becoming a serious agricultural problem, especially at arid and semi-arid zones, where 20-30% of the land is seriously damaged by salt [4]. High salt concentrations in the soil drastically reduce the yields of a variety of plants worldwide [5]. However, in Egypt, the saline and drainage water is one of the main waster resources for agriculture, which caused degradation of the soil and adversely affect on plant production [6].

Eggplant (*Solanum melongena* L.) is a traditional vegetable crop in many tropical, subtropical and Mediterranean
 countries. Eggplant is classified as a salt moderately sensitive vegetable [7]; <u>Bresler et al.</u> [8], on the other hand,
 classified it as salt sensitive vegetable. This difference in its tolerance classification could be related to differences in

42 used varieties or cultivars and study environmental conditions. Unlukara et al. [9] found a threshold value of salinity lower than 1.5 dS m⁻¹ and a slope value of 4.4% on eggplant. Such authors also reported a decrease in plant water 43 44 consumption due to salinity with a decrease slope of 2.1%.

45 Proline could accumulate in many plant species under a broad range of stress such as water shortage, salinity, 46 extreme temperatures and high light intensity. Proline is considered to be a compatible solute. It protects folded protein 47 structures against denaturation, stabilizes cell membranes by interacting with phospholipids, functions as a hydroxyl 48 radical scavenger and serves as an energy and nitrogen source. However, the contribution of proline to osmotic 49 adjustment and tolerance of plants exposed to unfavourable environmental conditions is still controversial [10]. The 50 metabolic effects of osmolyte accumulation may, however, be equal or even more important than their role in osmotic 51 adjustment, since stress-regulated changes in proline synthesis and degradation may also affect expression of other 52 genes, ensuring that the genetic response to stress is appropriate to the prevailing environmental stress conditions [11]. 53 Proline could accumulate in leaves and roots to protect against the osmotic pressure under salt stress [12].

54 Although selenium (Se) is not considered an essential nutrient for plant growth, it is a vital element for human and animal nutrition in trace amounts [13]. However, a diet containing 1 mg Se kg⁻¹ dry weight (DW) may lead to chronic 55 Se poisoning in humans and animals, and one-time ingestion of plant material containing 1,000 mg Se kg⁻¹ DW can lead 56 57 to acute Se poisoning and death [14] as it is shown from the conducted experiments (becoming toxic for human and 58 animals fed with these plants). Selenium is a constituent of seleno-proteins, many of which have important functions, 59 including antioxidant protection, energy metabolism and redox regulation during transcription and gene expression [15]. 60 Selenium supplementation to plants enhance the production and quality of edible plant products, by increasing 61 antioxidant activity of plants, as shown in tea leaves [16], and rice [17]. Foliar application of selenium was shown to be 62 several times more efficient than application in soil fertilizers [18], but strongly dependent on spraying conditions. Also, 63 [19] showed that foliar spray gave a high recovery. However, [20] found foliar application to be less efficient than 64 application to soil at planting.

65 66

Thus, the main objective of this study is to evaluate the protective effect by foliar application of selenium supplements (0, 5, 10, 20, 30 µM Na₂SeO₃) on eggplant (vegetative growth, yield, proline and some elements content) grown on a sandy soil and irrigated with different concentrations of saline water (0, 30, 60, 120 mM NaCl). 67

68 69

2. MATERIALS AND METHODS

The current experiment was carried out in the experimental station at the Central Laboratory for Agricultural
Climate (CLAC), Agriculture Research Center (ARC), Egypt, during the summer seasons of 2014 and 2015. The
climatic data at Dokki site during the studied seasons of 2014 and 2015 were shown in Figure 1; these data collected
from automated weather station allocated at the site.





100 **2.1 Plant materials**

99

Eggplant (*Solanum melongena* L. cv. Baladi) seeds were sown on 20th and 18th January of 2014 and 2015,
 respectively, in polystyrene trays. After the fifth true leaf stage (26th and 23th February, respectively), the eggplant
 seedlings were transplanted into bedding system of sandy soil.

104 2.2 System materials

Open system of sandy soil from Siwa oasis - Matroh governorate, *Typic Torripsamments*, was used and the
physical and chemical characteristics of the soil were listed in Table 1. The system bed performed of bricks on cement
base (60cm width x 25cm height x 7.5m length). The final plant spacing was 50cm in the row and 40cm in-between.
Black polyethylene (1mm) was used to create the main gully which was filled by the soil. A layer of 2-3cm of gravel
takes a place in the bottom of gully bin for leaching the drainage water easily.

110 No organic matter or manure was applied to the soil to avoid the effect of organic matter on the salinity impacts111 under the investigated different treatments.

Different salinity irrigation water levels were pumped via submersible pump (110 watt). A plastic tank 120 L (one per each bin system) and submersible pump (one per each tank) were used to pump the nutrient solution and different salinity irrigation water levels via polyethylene pipe (16mm) with 2 liters per hour dripper. The nutrient solution [21] was adjusted by using EC meter to the required level (2.5 dS m⁻¹) in all treatments. The fertigation was programmed to work 2 - 4 times/day and the duration of irrigation time depended upon the season.

- 117
- 118

Table 1. Some physical and chemical characteristics of the studied soil						
Particle size distribution, %		Soluble cations, meq L ⁻¹				
Sand	97.5	Ca ²⁺	3.40			
Silt	1.50	Mg^{2+}	3.50			
Clay	1.00	Na^+	1.50			
Texture class	Sandy	\mathbf{K}^+	0.60			
CaCO ₃ , %	5.80	Soluble anions, meq L ⁻¹				
OM, %	0.05	CO_{3}^{2}	0.00			
CEC, cmolc kg ⁻¹	9.30	HCO ₃ ²⁻	3.20			
pH (1:2.5 soil:water suspension)	8.19	Cl	4.10			
ECe, dS m ⁻¹	0.90	SO_4^{2-}	1.70			

119

120 2.3 Investigated treatments

121 The application of different treatments, after 2 weeks of transferring the eggplant seedlings, was applied. The 122 study investigated the effects of different selenium (Se) concentrations (0, 5, 10, 20, 30 μ M Na₂SeO₃) as foliar 123 application on eggplant cultivated under different saline irrigation water levels (0, 30, 60, 120 mM NaCl) on the studied 124 soil. The EC_w concentrations and Se supplements were applied according to [22, 23, 24, 25].

Eggplants were harvested on 26th and 17th June 2014 and 2015, respectively,... The collected samples were dried at
 70°C in an air forced oven for 48hrs, and then digested by H₂SO₄/H₂O₂ mixture according to the method described by
 [26] and kept for the elements determination.

128 2.4 Experiment design

The experimental design was a split plot with 3 replicates. Each experimental plot contained 10 plants. The saline
 irrigation levels were assigned as main-plots and Se concentrations as sub-plots as Figure 2 illustrates.



1.10	
140	
141	
142	
143	Figure 2. The layout of experimental design
144	2.5 Measurements
145	The vegetative and yield characteristics beside the chemical analysis of eggplants were measured as follows:
146	- Plant height (cm), before starting the flowering stage
147	- Number of leaves per plant, before starting the flowering stage
148	- Fresh weight of total fruits per plant (g/plant)
149	- Number of fruits per plant
150	- Chlorophyll content in leaves, SPAD:
151	Total chlorophyll of the fifth mature leaf from top was measured using Minolta chlorophyll meter Spad-501.
152	- Proline content in 0.5 g of fresh leaves, at harvest:
153	Proline content can increase upon exposure of plants to drought, salinity, cold, heavy metals, or certain pathogens.
154	Thus, determination of proline levels is a useful assay to monitor physiological status and to assess stress tolerance of
155	higher plants. Proline content was determined according to the method of [27] modified by [28], and was expressed as
156	mg g^{-1} fresh weight (FW).
157	- Total N, P, K and Na contents in leaves and fruits, at harvest:
158	Total nitrogen in plant was determined using Kjeldahl method according to the procedure described by [26]; total
159	phosphorus was determined using spectrophotometer according to [29] and both total potassium and sodium in plant
160	was determined using Flame photometer as described by [30].
161	Statistical analysis was performed using the analysis of variance adopting a SAS software package [31].
162	Significance among treatments was evaluated using Duncan's approach ($P \le 0.05$).
163	
164	3. RESULTS AND DISCUSSION
165	3.1 Vegetative growth and yield of eggplants
166	Data in Table 2 showed that the plant height was different under the studied treatments, with decreasing along
167	increasing irrigation water salinity. However, the plant height increased with increasing Se supplements. Regarding the
168	interaction between irrigation water salinity and Se supplements, the irrigation with tap water and Se 20 μ M were found
169	to give the highest plant; the irrigation water with EC_e 13.5 dSm ⁻¹ and Se 0 μ M gave the lowest one. Also, number of
170	leaves per plant went hand by hand with the previous findings on the plant height.
171	The total fruit fresh weight and the number of fruits per plant (Table 2) were agreed with the findings on the plant
172	height and number of leaves per plant. Generally, the Se supplement with 20 μM showed the best effects on vegetative
173	growth and yield of eggplants under different irrigation water salinity treatments, with higher effect for irrigation with
174	tap water and decreased with increasing salinity in irrigation water. These findings may be due to: (1) Se 20 μM is
175	suitable to counter act salinity problems inside the plant. (2) The vegetative growth and yield of eggplants decreased
176	with increasing salinity in irrigation water. Kabata-Pendias and Pendias [32] mentioned that the mean Se content in clay
177	soils was 0.29 mg kg ⁻¹ and in coarse mineral soils 0.17 mg kg ⁻¹ , and in plant Se at 10 ppm DW is considered as
178	phytotoxic. Yassen et al. [25] reported that Se interaction with plants depended on its concentration. At lower rates, Se
179	stimulated growth of ryegrass seedlings, while at high doses it acted as pro-oxidant reducing yields and inducing
180	metabolic disturbances. Terry et al. [13] found that there was a small decrease in shoot accumulation of Se with
	5

181 increasing salt level. <u>Unlukara et al.</u> [9] added that vegetative dry weight of the eggplants decreased with increasing soil

182 salinity and with fruit yield being more sensitive.

Table 2. Effect of irrigation water salinity and selenium supplements on average vegetative growth and yield of eggplants during the two studied seasons

Salinity levels,		Selenium c						
$dS m^{-1}(A)$	Se 0	Se 5	Se 10	Se 20	Se 30	Mean (A)		
Plant height, cm								
$EC_{w} = 0.75$	55.2 g	59.7 d	62.1 c	71.7 a	65.0 b	62.7 A		
$EC_w = 3$	52.3 h	57.0 f	61.2 c	64.7 b	61.3 c	59.3 B		
$EC_w = 7$	45.3 j	49.7 i	58.0 e	60.3 cd	58.0 e	54.3 C		
$EC_{w} = 13.5$	35.01	43.7 k	45.7 ј	56.0 g	52.7 h	46.6 D		
Mean (B)	47.0 D	52.5 C	56.7 B	63.2 A	59.3 B			
			No. of leaves	/plant				
$EC_{w} = 0.75$	68.3 i	72.8 h	84.0 f	128 a	113 b	93.2 A		
$EC_w = 3$	62.7 k	72.0 h	76.7 g	98.3 d	101 c	82.1 B		
$EC_w = 7$	60.31	63.7 k	67.7 i	100 c	87.0 e	75.7 C		
$EC_{w} = 13.5$	59.31	63.3 k	66.3 j	83.0 f	72.3 h	68.9 D		
Mean (B)	62.7 E	68.0 D	73.7 C	102 A	93.3 B			
		Fr	uit fresh weigl	ht, g/plant				
$EC_{w} = 0.75$	1149 e	1222 d	1342 c	1686 a	1475 b	1375 A		
$EC_w = 3$	748 h	880 g	1019 f	1125 e	1036 f	962 B		
$EC_w = 7$	517 k	597 ј	675 i	850 g	734 h	675 C		
$EC_{w} = 13.5$	356 m	4331	513 k	573 jk	544 k	484 D		
Mean (B)	693 D	783 C	887 B	1058 A	947 B			
No. of fruits/plant								
$EC_{w} = 0.75$	19.5 d	20.0 d	21.0 c	24.4 a	22.0 b	21.4 A		
$EC_w = 3$	14.7 h	15.7 g	17.3 f	18.4 e	17.0 f	16.6 B		
$EC_w = 7$	11.7 ij	11.8 i	12.3 i	14.7 h	13.8 h	12.8 C		
$EC_{w} = 13.5$	9.15 k	9.60 k	10.8 j	10.8 j	11.2 ј	10.3 D		
Mean (B)	13.8 C	14.3 C	15.3 B	17.1 A	16.0 B			

183 # Each value is the mean of 6 replications

184 This is a factorial experiment from two factors: salinity levels (A), selenium concentrations (B) in a spilt plot design,

letters A B C D among the main factors, letters a b c d ... among the interaction between the two factors (A×B), and
different letters means significant.

187

188 3.2 N, P and K contents in leaves and fruits of eggplants

189 Data in Table 3 showed the effect of irrigation with saline water on N, P and K contents in leaves of eggplants 190 under Se supplements, compared with the control (without any treatments). Increasing salinity resulted in increased N 191 and P contents in the leaves, but decreased K. Almost, N, P and K contents in leaves increased with increasing Se 192 supplements up to 20 µM then decreased with higher concentrations. Regarding the interaction between irrigation water 193 salinity and Se supplements, Se 20 µM with all saline water treatments generally gave the highest value of N, P and K 194 contents in plant leaves; almost all saline water treatments without Se supplement gave the lowest ones.

Salinity levels,						
$dS m^{-1}(A)$	Se 0	Se 5	Se 10	Se 20	Se 30	Mean (A)
			%N			
EC _w = 0.75	2.801	3.36 k	3.50 ј	4.34 e	3.78 h	3.56 D
$EC_w = 3$	3.78 h	3.76 h	3.64 i	4.48 d	3.99 g	3.93 C
	4. 48	4.8	4. 22	4. 90	4. 01	
$EC_w = 7$	d	5 C	f	с	g	4.49 B
EC _w =	5. 88	4.9 4.c	4. 90	5. 32	4. 20	
13.5	a	C	с	b	f	5.05 A
Meen	4. 24	4.2	4. 07	4. 76	4.	
(B)	B	3 B	C C	A	C	
			%P			
$EC_{w} = 0.75$	0.61 h	0.62 h	0.65 g	0.70 f	0.65 g	0.65 C
$EC_w = 3$	0.64 g	0.67 f	0.69 f	0.75 e	0.68 f	0.69 C
$EC_w = 7$	0.69 f	0.74 e	0.79 d	0.82 c	0.76 e	0.76 B
$EC_{w} = 13.5$	0.81 c	0.86 b	0.87 b	0.93 a	0.86 b	0.87 A
Mean (B)	0.69 D	0.72 C	0.75 B	0.80 A	0.74 B	
			%K			
EC _w = 0.75	1.62 e	1.81 c	1.94 b	1.84 c	1.70 d	1.78 A
$EC_w = 3$	1.50 g	1.77 c	1.79 c	1.62 e	1.65 e	1.67 B
$EC_w = 7$	1.26 i	1.57 f	1.58 f	2.20 a	1.58 f	1.64 B
$EC_{w} = 13.5$	1.09 k	1.16 j	1.23 i	1.40 h	1.53 g	1.28 C
Mean (B)	1.37 D	1.58 C	1.63 B	1.77 A	1.62 B	

Table 3. Effect of irrigation water salinity and selenium supplements on average N, P and K contents of eggplant leaves during the two studied seasons

195 # Each value is the mean of 6 replications

196 This is a factorial experiment from two factors: salinity levels (A), selenium concentrations (B) in a spilt plot design,

197 letters A B C D among the main factors, letters a b c d ... among the interaction between the two factors (A×B), and

198 different letters means significant.

199

200 Regarding the fruits of eggplant, data in Table 4 showed the effect of irrigation water salinity on N, P and K 201 contents under Se supplements which have similar trends with those of plant leaves. Increasing N and P contents in 202 leaves and fruits of eggplant with increasing salinity of irrigation water may be due to increase the amino acids inside 203 the plant with increasing the stress; amino acids also interact with phospholipids to adjust the osmotic potential 204 according to [33]. Also, they reported that Se had a high ability to induce antioxidant and hormon balance in the plant. 205 Yassen et al. [25] found that foliar application of Se on potato plants increased % N, P, K and protein contents in the 206 yield of tubers. Decreasing K content with increasing salinity of irrigation water, on the other hand, may be due to the increase of NaCl concentration; Na⁺ content increased in leaves and fruits indicating that the eggplant (which has a 207 208 glycophytic reaction) could not control uptake of Na⁺ [34]. Kong et al. [15] reported that the major influences on Se 209 uptake by plants were soil pH and salinity; Cl⁻ which inhibit uptake by affecting plant metabolism. In general, 210 increasing N, P and K contents in leaves and fruits of eggplant by increasing foliar Se supplements under irrigation with 211 saline water may be due to the role of Se in increasing antioxidant activity of the plant to face the stress.

Salinity levels,	Selenium concentrations, µM (B)							
$dS m^{-1}(A)$	Se 0	Se 5	Se 10	Se 20	Se 30	Mean (A)		
	%N							
$EC_{w} = 0.75$	2.38 k	2.41 k	2.52 ј	2.55 ј	2.94 g	2.56 C		
$EC_w = 3$	2.40 k	2.57 i	2.68 h	3.10 f	3.02 j	2.75 C		
$EC_w = 7$	3.50 d	3.08 f	3.06 f	3.22 e	3.61 c	3.29 B		
EC _w = 13.5	3.96 a	3.94 b	3.20 e	3.50 d	3.94 b	3.71 A		
Mean (B)	3.06 B	3.00 C	2.87 D	3.09 B	3.38 A			
				%P				
$EC_{w} = 0.75$	0.48 i	0.51 h	0.54 g	0.57 f	0.51 h	0.52 C		
$EC_w = 3$	0.55 g	0.60 e	0.62 e	0.68 c	0.65 d	0.62 C		
$EC_w = 7$	0.61 e	0.65 d	0.68 c	0.73 b	0.69 c	0.67 B		
$EC_{w} = 13.5$	0.67 c	0.72 b	0.73 b	0.79 a	0.72 b	0.73 A		
Mean (B)	0.58 D	0.62 C	0.64 B	0.69 A	0.64 B			
%K								
$EC_{w} = 0.75$	1.16 i	1.19 i	1.24 g	1.60 b	1.57 b	1.35 A		
$EC_w = 3$	1.11 i	1.14 i	1.23 hi	1.52 c	1.52 c	1.31 B		
$EC_w = 7$	1.06 j	1.16 i	1.21 i	1.67 a	1.48 d	1.32 B		
EC _w = 13.5	0.99 k	1.06 j	1.17 i	1.33 f	1.41 e	1.19 C		
Mean (B)	1.08 D	1.14 C	1.21 B	1.53 A	1.50 A			

Table 4. Effect of irrigation water salinity and selenium supplements on average N, P and K contents of eggplant fruits during the two studied seasons

Each value is the mean of 6 replications

213 This is a factorial experiment from two factors: salinity levels (A), selenium concentrations (B) in a spilt plot design,

214 letters A B C D among the main factors, letters a b c d ... among the interaction between the two factors (A×B), and

215 different letters means significant.

216

217 3.3 Some stress markers in leaves of eggplants

218 Data in Table 5 showed that chlorophyll content in plant leaves increased with increasing salinity of irrigation water, 219 but decreased with increasing Se supplements, significant decrease in the treatment of Se 20 µM being found under 220 irrigating with tap water. Increasing chlorophyll content in plant leaves indicates that plant suffered from saline stress 221 compared to the control (irrigation with tap water). It may be due to a reduction in extension growth under saline stress, 222 increased leaf thickness and its color became darker, which gave high readings with SPAD. Khattab [35] studied the 223 metabolic and oxidative responses associated with exposure of rocket plants (Eruca sativa L.) to different levels of Se 224 (0, 5, 10, 100, 1000, 2000 and 3000 µM of sodium selenate) for 10 days. He found that Se up to 10 µM enhanced the 225 growth and levels of chlorophylls, sugar and amino acids. However, high levels of Se (100 µM and up) exert toxic 226 effects. Germ et al. [36] reported that Se protected chloroplasts during stress.

227Regarding the proline content in plant fresh leaves, data also showed that proline content increased with increasing228salinity of irrigation water (indication of stress), but decreased with increasing Se supplements compared to the control.229The treatment of EC_e 13.5 dS m⁻¹ without Se supplements gave the highest value of proline content (51 mg g⁻¹), control230treatment being the lowest one (30.9 mg g⁻¹). These findings agreed with those obtained by [33] regarding the effect of231Se on proline content in cucumber seedlings grown under saline conditions. They explained the accumulation of proline232in plant under the stress by increasing biosynthesis or inhibition of proline degradation. Nowak [22] reported that Se233enhanced the salt tolerance of cucumber seedlings by protecting the cell membrane against lipid peroxidation. However,

he explained the growth-promoting effect of low Se concentrations (5 and 10 μ M) under saline conditions due to the

antioxidative activity of Se, increase in proline accumulation and/or decrease in content of Cl⁻ ions in the shoots tissues.

236

- 237
- 238
- 239

240

during the two studied seasons								
Salinity levels,		Selenium						
$dS m^{-1}(A)$	Se 0	Se 5	Se 10	Se 20	Se 30	Mean (A)		
Chlorophyll content, SPAAD								
$EC_{w} = 0.75$	51.31	53.5 k	54.2 ј	55.2 i	55.1 i	53.9 C		
$EC_w = 3$	61.2 d	59.3 f	56.4 i	55.3 i	57.7 h	58.0 C		
$EC_w = 7$	62.3 b	60.7 e	60.5 e	57.7 h	58.9 g	60.0 B		
$EC_{w} = 13.5$	65.4 a	62.5 b	60.3 e	59.5 f	61.7 c	61.9 A		
Mean (B)	60.1 A	59.0 B	57.9 B	56.9 C	58.4 B			
			Proline con	tent, mg g ⁻¹	FW			
$EC_{w} = 0.75$	30.9 n	32.3 m	36.01	36.3 k	36.5 j	34.4 D		
$EC_w = 3$	45.3 g	42.1 h	40.1 i	45.3 g	45.5 f	43.7 C		
$EC_w = 7$	47.7 b	45.7 e	45.4 g	45.7 e	47.4 c	46.4 B		
$EC_{w} = 13.5$	51.0 a	50.9 a	45.9 d	47.4 c	47.8 b	48.6 A		
Mean (B)	43.7 B	42.8 C	41.9 D	43.7 B	44.3 A			
K/Na ratio								
$EC_{w} = 0.75$	1.41 e	1.47 d	1.54 c	1.65 b	1.71 a	1.56 A		
$EC_w = 3$	0.95 i	1.03 h	1.03 h	1.08 g	1.14 f	1.05 B		
$EC_w = 7$	0.86 j	0.88 j	0.94 i	0.98 i	0.89 j	0.91 C		
$EC_{w} = 13.5$	0.52 m	0.77 k	0.78 k	0.88 j	0.711	0.73 D		
Mean (B)	0.94 E	1.04 D	1.07 C	1.15 A	1.11 B			

Table 5. Effect of irrigation water salinity and selenium supplements on some stress markers in eggplant leaves

241 # Each value is the mean of 6 replications

242 This is a factorial experiment from two factors: salinity levels (A), selenium concentrations (B) in a spilt plot design,

letters A B C D among the main factors, letters a b c d ... among the interaction between the two factors (A×B), and
different letters means significant.

245 246

Data in Table 5 showed values of K/Na ratio in leaves of eggplants as an important indicator on salinity stress. K/Na ratio decreased with increasing salinity of irrigation water, but increased generally with increasing Se supplements. Regarding the interaction between irrigation water salinity and Se supplements, the treatment of EC_e 13.5 dS m⁻¹ without Se supplements gave the lowest value of K/Na ratio (0.52), the treatment of Se 30 μ M under irrigation with tap water being the highest one (1.71). <u>Akinci et al.</u> [34] <u>reported</u> that increasing NaCl in the solution led to a decrease in the K/Na ratio and increased Na in several eggplant varieties.

252 Germ et al. [36] reported that, in the senescing plants, the addition of Se strengthens the antioxidative capacity by
 253 preventing the reduction of tocopherol concentration and by enhancing superoxide dismutase (SOD) activity.
 254 Senescence processes are partly delayed due to enhanced antioxidation, which is associated with an increase of
 255 glutathione peroxidase (GPx) activity. In ryegrass (*Lolium perenne*) up to Se addition of 1.0 mg kg⁻¹, the decreased lipid

256 257

peroxidation was connected with Se-induced increase in GPx activity [37]. It was shown that Se has the ability to regulate the water status of plants under conditions of drought [38], and that the protective effect of Se under drought 258 stress conditions was achieved by increasing the water uptake capacity of the root system.

259

260 4. CONCLUSION

261 Under mitigation and adaption strategy of climate change impacts with the expected increase in the salinity of irrigation 262 water especially in the Northern Egypt as a result of sea level rise, the present study recommends the applying selenium 263 as foliar application at the concentration 10 - 20 μ M to increase the tolerance of eggplants against salinity of irrigation 264 water and to avoid salinity stress on the yield. These concentrations of Se supplementation present a promising potential 265 for use in conditions of relatively high levels of NaCl in the irrigation water, due to its antioxidative activity. More work 266 is required to investigate the effect of Se on different crops, the real role of it inside the plant in physiological stages and 267 its content; besides studying the impact of sea level rise on the irrigation water and soil salinities.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

273 REFERENCES

274

268 269

270 271

272

- 275 1. Nicholls RJ, Leatherman SP. Global sea-level rise. In: Strzepek KM, Smith JB (eds.), When Climate Changes: 276 Potential Impact and Implications. Cambridge Univ. Press, Cambridge 1995.
- 277 2. El-Raey M, Nasr S, Frihi S, Desouki S, Dewidar K. Potential impacts of accelerated sea level rise on Alexandria 278 governorate, Egypt. J. Coastal Res. 1995;14:190-204.
- 279 3. Sengupta S, Majumder AL. Insight into the salt tolerance factors of a wild halophytic rice, *Porteresia coarctata*: a 280 physiological and proteomic approach. Planta 2009;229:911-929.
- 281 4. FAO. Crops and drops. Making the best use of water for agriculture. Food and Agriculture Organization of the 282 United Nations. Roma 2002;p 26.
- 283 5. Gorai M, Neffati M. Germination responses of *Reaumuria vermiculata* to salinity and temperature. Annals Appl. 284 Biol. 2007;151:53-59.
- 285 6. Abd-Elrahman Shaimaa H. Remediation of some degraded soils using new techniques. Scholars' Press, 286 OmniScriptum GmbH & Co. KG, Germany 2013;p 8.
- 287 7. Heuer B, Meiri A, Shalevet J. Salt tolerance of eggplant. Plant Soil 1986;95:9-13.
- 288 8. Bresler E, McNeal BL, Carter DL. Saline and sodic soils. Springer-Verlag, Berlin 1982.
- 289 9. Unlukara A, Kurunc A, Kesmez GD, Yurtseven E, Suarez DL. Effects of salinity on eggplant (Solanum melongena 290 L.) growth and evapotranspiration. Irrig. & Drain. 2010;59:203-214.
- 291 10. Hare PD, Cress WA. Metabolic implications of stress-induced proline accumulation in plants. Plant Growth 292 Regulation 1997;21:79-102.
- 293 11. Claussen W. Proline as a measure of stress in tomato plants. Plant Sci. 2005;168:241–248.

- 294 12. Chen Z, Cuin TA, Zhou M, Twomey A, Naidu BP, Shabala S. Compatible solute accumulation and stress mitigating
 295 effects in barley genotypes contrasting in their salt tolerance. J. Exp. Bot. 2007;58:4245-4255.
- 296 13. Terry N, Zayed AM, de Souza MP, Tarun AS. Selenium in higher plants. Annual Rev. Plant Physiol. Plant Mol.
 297 Biol. 2000;51:401–432.
- 298 14. Pilon-Smits, Elizabeth AH, Quinn, Colin F. Selenium metabolism in plants. In: Hell R, Mendel RR. (eds.), Cell
 299 Biology of Metals and Nutrients, Plant Cell Monographs 2010;17:225-241.
- Kong L, Wang M, Bi D. Selenium modulates the activities of antioxidant enzymes, osmotic homeostasis and
 promotes the growth of sorrel seedlings under salt stress. Plant Growth Regulation 2005;45:155-163.
- 302 16. Xu J, Yang F, Chen L, Hu Y, Hu Q. Effect of selenium on increasing the antioxidant activity of tea leaves harvested
 303 during the early spring tea producing season. J. Agric. Food Chem. 2003;51:1081-1084.
- 304 17. Xu J, Hu Q. Effect of foliar application of selenium on the antioxidant activity of aqueous and ethanolic extracts of
 305 selenium-enriched rice. J. Agric. Food Chem. 2004;52:1759-1763.
- 306 18. Aspila P. History of selenium supplemented fertilization in Finland. Proceedings, 20 Years of Selenium
 307 Fertilization; Helsinki, Finland 2005;8–9:8-13.
- 308 19. Curtin D, Hanson R, Lindley TN, Butler RC. Selenium concentration in wheat grain as influenced by method, rate,
 309 and timing of sodium selenate application. New Zealand J. Crop Hort. Sci. 2006;34:329-339.
- 20. Lyons GH, Lewis J, Lorimer MF, Holloway RE, Brace DM, Stangoulis JCR. High selenium wheat: agronomic biofortification strategies to improve human nutrition. Food Agric. Environ. 2004;2:171-178.
- 21. El-Behairy UA. The effect of levels of phosphorus and zinc in the nutrient solution on macro and micronutrients
 uptake and translocation in cucumber (*Cucumus sativus* L.) grown by nutrient film technique. Ph.D thesis, London
 University 1994; p. 299.
- 22. Nowak HB. Beneficial effects of exogenous selenium in cucumber seedlings subjected to salt stress. Biol. Trace
 Elem. Res. 2009;132:259-269.
- 317 23. Yao X, Chu J, Wang G. Effects of selenium on wheat seedlings under drought stress. Biol. Trace Elem. Res.
 318 2009;130:283-290.
- 24. Chu J, Yao X, Zhang Z. Responses of wheat seedlings to exogenous selenium supply under cold stress. Biol. Trace
 Elem. Res. 2010;136:355-363.
- 321 25. Yassen AA, Safia M Adam, Sahar M Zaghloul. Impact of nitrogen fertilizer and foliar spray of selenium on growth,
 322 yield and chemical constituents of potato plants. Aust. J. Basic & Appl. Sci. 2011;5:1296-1303.
- 323 26. <u>Chapman HD, Pratt PF. Methods of analysis for soils, plants and waters. Division of Agric. Sci., Berkeley, Univ.</u>
 324 <u>California, USA 1961;pp 150-152.</u>
- 325 27. Troll W, Lindsley J. A photometric method for the determination of proline. J. Biol. Chem. 1955;215:655-660.
- 326 <u>28</u>. Petters W, Piepenbrock M, Lenz B, Schmitt JM. Cytokinine as a negative effector of phosphoenolpyruvate
 327 carboxylase induction in *Mesembryanthemum crystallinum*. J. Plant Physiol. 1997;151:362-367.
- Watanabe FC, Olsen SR. Test of an ascorbic acid method for determining phosphorus in water and NaHCO₃
 extracts from soils. Soil Sci. Soc. Amer. Proc. 1965;29:677-678.
- 30. Jackson ML. Soil chemical analysis. Prentice-Hall, Inc. Englewood Cliffs, NJ Library of Congress, USA 1958;pp
 38-388.
- 332 31. SAS Institute. The SAS System for Windows; Release 6.12; Statistical Analysis System Institute Inc., Cary, NC
 333 1996.

- 334 32. Kabata-Pendias A, Pendias H (Eds.). Trace elements in soils and plants. 2nd Ed. CRC Press, Inc. Boca Raton,
 335 Florida 1992.
- 33. Abd El-Nasser Walaa, Shatlah MA, Hossien M, Srorr HAM. Selenium induces antioxidant defensive enzymes and
 promotes tolerance against salinity stress in cucumber seedlings (*Cucumis sativus*). Arab Univ. J. Agric. Sci.
 2010;18:65-76.
- 339 34. Akinci IE, Akinci S, Yilmaz K, Dikici H. Response of eggplant varieties (*Solanum melongena*) to salinity in
 340 germination and seedling stages. New Zealand J. Crop Hort. Sci. 2004;32:193–200.
- 341 35. Khattab HI. Metabolic and oxidative responses associated with exposure of *Eruca sativa* (Rocket) plants to different
 342 levels of selenium. Int. J. Agric. Biol. 2004;6:1101-1106.
- 343 36. <u>Germ M, Stibilj V, Kreft I. Metabolic importance of selenium for plants. The Europ. J. Plant Sci. Biotech. 2007;</u>
 344 1:91-97.
- 345 37. <u>Hartikainen H, Xue T, Piironen V. Selenium as an antioxidant and prooxidant in ryegrass. Plant Soil 2000;225:193-</u>
 346 200.
- 347 38. Kuznetsov VV, Kholodova VP, Kuznetsov VIV, Yagodin BA. Selenium regulates the water status of plants exposed
 348 to drought. Doklady Biological Sciences 2003;390:266-268.