

Original Research Article

Phytochemical analysis and screening of Ugandan medicinal plants for antifungal activity against *Candida albicans*.

Kakudidi Esazah¹, Ayorekire Fredric², Okeng- Ogwal Jasper³, Anywar Godwin^{1*}

¹ Department of Biological Sciences, Makerere University (Mak), P. O. Box 7062, Kampala, Uganda

² Department of Biology, Mbarara University of Science and Technology (MUST) P. O. Box 1410, Mbarara, Uganda

³ Department of Pharmacology & Therapeutics, Makerere University (Mak), P. O. Box 7062, Kampala, Uganda

ABSTRACT

Aims: The potential activity against *Candida albicans* of five commonly used medicinal plant species of Bwindi Impenetrable Forest National Park in southwestern Uganda was investigated.

Study design: The phytochemical profiles of *Tetradenia riparia*, *Erucastum arabicum*, *Plectranthus lactiflorus*, *Solanecio mannii* and *Platostoma africanum* were analysed *in vitro*.

Place and Duration of Study: The experiments were carried out in the Department of Pharmacology and Therapeutics, College of Health Sciences, Makerere University and Department of Microbiology and Parasitology, College of Veterinary Medicine and Biosecurity, Makerere University, between September 2012 and January 2013.

Methodology: The Agar well diffusion method was used to measure the antifungal activity against *Candida albicans* (ATCC 10231). The Minimum Inhibitory Concentration (MIC) was determined by the serial dilution method and the phytochemical analyses were made by standard phytochemical tests.

Results: Antifungal activity of both ethanol and diethylether extracts of *Tetradenia riparia* surpassed the Nystatine standard (31 and 28 > 25, respectively) as well as those of other four plant species. The MIC values for the ethanol and ether crude extracts were 0.0312 and 0.0156 g/ml respectively. The plant species tested proved to be positive for different phytochemicals including tannins, coumarins and steroidal glycosides.

Conclusion: Generally, all plant species proved to be active against *Candida albicans*. *Tetradenia riparia* exhibited the highest antifungal activity and it is considered to be a potential source of antifungal drugs.

Keywords: Fungal infections, herbal medicine, Bwindi, *Candida albicans*, Medicinal plants, *Tetradenia riparia*

1. INTRODUCTION

The frequencies of fungal infections are recently increasing [1]. Pathogenic fungi in general and *Candida albicans* in particular, are more difficult to control than non-pathogenic fungi and [2,3,4]. *C. albicans* remains the most frequent cause of fungal infections in an expanding population of immuno-compromised patients [2], and also plays a role in sexual transmitted infections [5]. The pathogenic fungi produce superficial infections, not only in immune-compromised hosts but also in healthy individuals, living under poor sanitary conditions, especially in the developing world [6]. Candidiasis has become a major public health problem as an opportunistic infection of HIV/AIDS [7]. Bacterial and fungal infections have been

reported to be prevalent in Uganda [8]. Fungi widely affect different body parts like skin, hair and nails [9].

There are several challenges that affect the successful treatment of fungal infections. These include limited and costly drugs, microbial drug resistance, toxicity, and relapse of infections [10, 11]. The search for novel antifungal agents relies in great part on ethnobotanical information and ethnopharmacologic exploration [12, 4]. Plants have been found to play a major role in managing dermatological conditions [13, 14]. Svetaz *et al.*, [15] found a significantly higher probability ($p < 0.01$) of detecting plants with antifungal activity ($MIC \leq 1000 \mu g/mL$) against at least one fungus when plants possess reported ethnopharmacological uses related to fungal infections. Therefore medicinal plant species have a great potential as good sources for new and safe antifungal drugs [3].

Kamatenesi *et al.* [8] reported that people in rural parts of Uganda use herbal drugs to treat bacterial and fungal infections. In Bwindi, a typical rural area in southwestern Uganda, the people do not have access to modern medical facilities and services. They, highly depend on traditional medicinal herbs to treat many ailments including fungal infections. The signs or symptoms that give support to a traditional antifungal use are mainly related to skin or mucosal conditions [15]. This study was therefore carried out to investigate the use of selected medicinal plant species used by this community and their potential in treating fungal skin infections.

2. MATERIAL AND METHODS

2.1 Study area

Bwindi is situated in Kigezi highlands, southwestern Uganda, between $29^{\circ} 35'$ to $29^{\circ} 50'$ E and $0^{\circ} 53'$ to $1^{\circ} 8'$ S [16]. The altitude ranges between 1,190m and 2,607m with a rough terrain characterized by numerous steep sided hills and narrow valleys of a general incline from the south western to the northwestern parts. Geologically, the area is associated with up warping of the Western Rift Valley. The climate is tropical with two rainfall peaks from March to May and September to November. Annual rainfall intensity ranges between of 1,400 mm and 1,900 mm, while the temperature ranges between $7^{\circ} C$ and $15^{\circ} C$ [17].

2.2 Selection of antifungal medicinal plants for bioassay and phytochemical screening

Five of the most frequently used medicinal plant species in areas around BINP were selected for analysis of antifungal activity *in vitro*. The selection of plants for analysis was based on their frequency of mention and preferred use by the traditional healers and the respondents interviewed. The 5 plant species selected were; *Tetradenia riparia* (Hochst.) Codd, *Erucastrum arabicum* Fisch. & C.A.Mey., *Plectranthus lactiflorus* (Vatke) Angew, *Solanecio mannii* (Hook.f.) C. Jeffrey and *Platostoma africanum* P.Beauv. (Table 1).

2.3 Collection and drying of plant materials

Voucher specimens of the plant species the herbalists use to prepare antifungal remedies were collected and taken for identification at the Makerere University Herbarium following the procedures described in [18]. The plant species were classified using the Kew database at www.theplantlist.org accessed on 25/10/2014. In order to collect representative samples of plant specimens for phytochemical and antifungal analyses, procedures described in Hue *et al.*, [19] were followed. The plant materials were collected from the parishes of Rutugunda, Nteko, Rubuguri, Nyamabare, Mpungu, Mukono and Rubimbwa adjacent to the forest. They were air-dried at room temperature for 2 weeks to avoid decomposition of volatile chemical in plant materials. The different dried materials were separately pounded using mortar and pestle then sieved into fine powder, and stored in airtight jars.

2.4 Preparation of extracts

Diethylether and ethanol were used as the extracting solvents for the plant materials. For each plant species, 35g of powder was soaked in petroleum ether in series for 36 hours, with periodic shaking. The extract was filtered off and fresh solvent added. This was repeated 3-5 times until there was almost no

more coloration acquired by the solvent. Each filtrate was then separately concentrated using a rotary evaporator. The dry residue was then further extracted using ethanol in series of 4 to 5 times as above but as the solvent. After recovering the extracting solvents, the concentrated crude extracts were kept in separate containers and labeled appropriately.

2.5 Phytochemical analysis of ethanol crude extracts

Phytochemical screening of crude extracts was conducted to identify the major chemical groups in the extracts using standard tests as described by various authors [20, 21].

2.6 Antifungal susceptibility tests

The Agar well diffusion assay was used to test the activity of each crude extract on a culture of *Candida albicans* (ATCC 10231) obtained from Department of Microbiology and Parasitology, College of Veterinary Medicine, Animal Resources and Biosecurity at Makerere University. Nystatin was used as the positive control since it is a polyene-macrolide antifungal antibiotic that is widely available for the topical treatment of localized fungal infections and is known to be effective against a variety of fungal infections in humans [22, 23].

Dehydrated Mueller Hinton agar (38g) was suspended in 1 liter of water and mixed well. The contents were boiled with frequent agitation for one minute to enable the powder dissolve completely. The resultant medium was autoclaved at 121°C for 25 minutes and then cooled in a water bath at 45 to 50°C, before being introduced into the sterilized Petri dishes. A representative sample of the medium was incubated at 27 to 30°C for 24 hours and examined to verify its sterility. Four wells of approximately 4 mm diameter were dug into the Petri dishes. The entire surface of the agar was inoculated with *C. albicans*. After inoculation, the plates were incubated at 27 to 30°C for 24 hours to allow the inoculum establish fully. One gram of extract was dissolved in 2 ml of Dimethyl sulphoxide (DMSO) separately introduced into the wells in the culture and incubated at 30 °C for 24 hours. The diameters of the zones of inhibition around the wells were measured.

2.7 Minimum Inhibitory Concentration (MIC) of extracts

MIC was defined as the lowest concentration of extract at which no fungal growth was observed after incubation. The MIC of each plant extract was determined by serial double dilution method using Brain Heart Infusion Broth (BHIB).

3. RESULTS

The commonly mentioned fungal diseases in Bwindi according to an earlier ethnobotanical study by Kakudidi et al. [24] were *Candidiasis*, *Tinea capitis*, *T.pedis*, *T. corporis*, *T. versicolor*, *T. manuum*, *T. unguium*, and *T. cruris*.

3.1 Growth inhibition zone

The activity of both ethanol and diethylether extracts of *T. riparia* against *C. albicans* surpassed Nystatin (Table 1). However, Nystatin had a greater diameter of the zone of inhibition than all the other plant extracts. Ethanol extract of *T. riparia* had the highest diameter (31 mm) of all plants screened. The extracts of other plants except the ether extract of *P. lactiflorus*, and the ethanol extracts of *P. africanum* and *S. maniis*, were active against *C. albicans*.

3.2 Minimum inhibition concentration

Ethanol extract of *T. riparia* had the lowest MIC against *C. albicans* (0.0156 g/ml) of all plants (Table 2). This was followed by the ether extract of the same plant *T. riparia* (0.0312 g/ml). All extracts apart from the ether extract of *P. latiflorus* and *S. manii* were active against *C. albicans*. The DMSO had not effect on the growth of *C. albicans*

3.3 Qualitative phytochemical analysis

Different plant parts of the different plant species were analysed (Table 3). Tannins were present in ethanol extracts of all the plant species analysed. Coumarin derivatives and steroid glycosides were also detected in all cases. None of the plant species analysed was positive for emodols. Anthocyanin pigments were only present in the hydrolysed ethanol extract of *T. riparia*. None of the plant species analysed was positive for alkaloid salts. Only *P. latiflorus*, *S. manii* and *P. africanum* contained basic alkaloids. Flavanosides were present only in the ether extract of the roots of *P. africanum* and the hydrolysed ethanol leaf extract of *S. manii*.

Table 1: Diameters (mm) of zones of growth inhibition of the plant extracts on *C albicans*

Plant Name	<i>T. riparia</i>		<i>E. arabicum</i>		<i>P. lactiflorus</i>		<i>S. manii</i>		<i>P. africanum</i>	
Plant Extracts	DE	Et	DE	Et	DE	Et	DE	Et	DE	Et
Diameter of zone of Inhibition (mm)	28	31	21	12	n.a	12	13	R	14	R

Key: DE – Diethyl Ether, Et– Ethanol, n.a – Not active, standard drug Nystatin 25 mm

Table 2: Minimum inhibitory concentration (MIC) of extracts (g/ml) on *Candida albicans*

Plant Name	<i>T. riparia</i>		<i>E. arabicum</i>		<i>P. lactiflorus</i>		<i>S. manii</i>		<i>P. africanum</i>	
Plant Extracts	DE	Et	DE	Et	DE	Et	DE	Et	DE	Et
MIC mg/g	0.0156	0.0312	0.0625	0.5	n.a	0.5	0.125	n.a	0.5	0.5

Key: DE – Diethyl Ether, Et – Ethanol, n.a – Not active

Table 3: Major Phytochemical compounds in the respective plant extracts

Plant name Parts used	<i>T. riparia</i> Leaf			<i>E. arabicum</i> Leaf			<i>P. lactiflorus</i> Leaf			<i>S. manii</i> Stem & Leaf			<i>P. africanum</i> Root		
Plant extracts	DE	Et	HE	DE	Et	HE	DE	Et	HE	DE	Et	HE	DE	Et	HE
Phytochemicals															
Basic alkaloids	-	-	-	-	-	-	+	-	-	+	-	-	+	-	-
Alkaloid salts	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Anthocyanin	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
Coumarin derivatives	-	-	+	+	-	+	+	-	+	+	-	+	+	-	-
Emodols	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Flavone aglycones	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
Flavanosides	-	-	-	-	-	-	-	-	-	-	-	+	-	-	+
Higher fatty acids	+	-	-	+	-	-	-	-	-	-	-	-	-	-	-
Reducing compounds	-	+	-	-	+	-	-	+	-	-	-	-	-	-	-
Steroid glycosides	-	-	+	-	-	+	-	-	+	-	-	+	-	-	+
1. Sterols & triterpenes	+	-	-	+	-	-	+	-	-	+	-	-	-	-	-
2. Tannins	-	+	-	-	+	-	-	+	-	-	+	-	-	+	-

Key: Compound present (+), Compound absent (-), DE – Diethyl Ether, Et – Ethanol, EH – Hydrolysed Ethanol

4. DISCUSSION

Considering zones of inhibition as a measure of plant extracts activity, both the ether and ethanol extracts of *T. riparia* were more active against *C. albicans* than Nystatin. *T. riparia* has a great potential for the development of an antifungal agent. Previous ethnobotanical studies have mentioned some of these plant species to be used in treating fungal, bacterial or viral infections and other diseases in different parts of Africa [25, 26, 27, 28]. The active compounds of *E. arabicum*, *S. angulatus* and *P. africanum* could also be a potential source of antifungal drugs if they are purified.

The low MIC values of *T. riparia* of 0.0156 and 0.0312 mg/ml, for the ether and ethanol extracts respectively are indicative of the high activity the plant given low concentrations are required to inhibit the growth of *C. albicans*. Whereas the ethanolic extract of *S. manii* showed no observable effects on the growth of *C. albicans*, its ether was active against. Similarly, the ether extract of *P. lactiflorus* showed no observable effects on the growth of *C. albicans* while its corresponding ethanolic extract was active. This is indicative of the fact that the solvent used in extraction and preparation of herbal medicines an effect on the active ingredients extracted and their therapeutic efficacy. This therefore has implications on the efficacy of herbal remedies.

In general, ethanolic extracts of *T. riparia* and *P. lactiflorus* were found more active against *C. albicans* than their corresponding ether extracts. The ether extracts of *E. arabicum*, *P. africanum* and *S. manii* were more active than their corresponding ethanolic extracts. This implies that the active ingredients of *T. riparia* and *P. lactiflorus* are polar and therefore more soluble in a polar solvent while for *E. arabicum*, *P. africanum* and *S. angulatus* their active ingredients are non polar and therefore more soluble in ether which is a non polar solvent. Different researchers have reported the use of these plant species for various ethnomedicinal uses; *T. riparia* has been reported to be taken as a leaf decoction once a day for de-worming in Kabale district, western Uganda [25]. The leaves are used for treating tuberculosis in Uganda [26], conjunctivitis, meningitis, malaria, skin diseases, stomach ache, ulcers, bilhazia, worms, rheumatism, dyspepsia, fever, colds, cough, toothache, kwashirko, blocked fallopian tubes in the Rwenzori region, western Uganda [27], febrile convulsions in south western Uganda [28], bacterial and fungal infections as well as herpes zoster in queen Elisabeth Biosphere Reserve, western Uganda [8], cough and chest infections in the Eastern Cape province of South Africa [29].

The leaves of *Erucastrum arabicum* have been reported to be used for treating constipation and ringworm in western Uganda [30], red eyes in south western Uganda (Ssegawa & Kasenene 2007) [28], while the roots used to treat diarrhea in Kenya [31]. A root infusion of *Plectranthus lactiflorus* is used to treat rheumatism [30] and bacterial and fungal infections in queen Elisabeth Biosphere Reserve, western Uganda [8]. A stem infusion of *Solanecio manii* is drunk or bathed for relaxation of pelvic region for child birth in south western Uganda (Ssegawa & Kasenene 2007) [28], whereas a root decoction *Platostoma africanum* is used as an aphrodisiac and for headaches and used in Tanzania [32].

4.1 Major chemical groups

Some phytochemical compounds already known to have antifungal properties were present in some of the plants screened. Tannins have been demonstrated to have a powerful antifungal action in clinical studies and also have wound healing and antiseptic effects [33, 34], therefore, the tannins present in all the plant species analysed in this study could be responsible for their antifungal activity and can justify the ethnobotanical uses of the plant species by the local people. Cowan [35] also reported that several plant products utilized as antimicrobial agents to include quinones, flavonoids, tannins, coumarins, alkaloids and terpenoids. Flavonoids have also been shown to have antiviral, antiviral and antibacterial [36, 37].

A GC/MS analysis of the essential oils from the leaves and stems of *T. riparia* from South Africa identified 35 components, with the main constituents being α -terpineol (22.6%), fenchone (13.6%), β -fenchyl alcohol (10.7%), β -caryophyllene (7.9%), and perillyl alcohol (6.0%) [38]. The yields of essential oils have also been shown to range from 0.17% to 0.26% [39].

5. CONCLUSION

The people of Bwindi use several medicinal plants to treat fungal infections. Generally, all plant species examined were active against *Candida albicans*. *T. riparia* exhibited the highest antifungal activity of all the plant species analysed with inhibition diameters zones of 28 and 31 mm for the ether and ethanol extracts respectively. Both these zones were higher than those exhibited by the positive control – Nystatin, which had a diameter of zone of inhibition of 25mm. *T. riparia* also had the lowest MIC values for both extracts further confirming its potency even at low concentrations and therefore can guide in the development of alternative treatment for antifungal treatments especially in *Candida albicans*.

CONSENT

Prior informed consent as obtained from all the participants verbally before they were interviewed.

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