

Spectral Discrimination of Coral Reefs on the Small Islands, Spermonde Archipelago, Indonesia

ABSTRACT

Coral reefs play important ecological services such as providing foods, biodiversity, nutrient recycling etc. for human society. On the other hand, they are threatened by human impacts such as illegal fishing and environmental changes such as a rise of sea water temperature and sea level due to global warming. Thus it is very important to monitor dynamic spatial distributions of coral reefs and related habitats such as coral rubber, dead coral, bleached corals, seagrass, etc. Hyperspectral data, in particular, offer high potential for characterizing and mapping coral reefs because of their capability to identify individual reef components based on their detailed spectral response. We studied the optical properties by measuring *in situ* spectra of living corals, dead coral and coral rubber covered with algae. The study site was selected in Spermonde archipelago, South Sulawesi, Indonesia because this area is included in the highest diversity of corals in the world named as Coral Triangle, which is recognized as the global centre of marine biodiversity and a global priority for conservation. Spectra were collected under generally clear skies, between 9:00 a.m. and 15:00 p.m. Central Standard Time, using a LOT-2 Spectra Corpspectroradiometer. The samples comprised living and dead coral covered with alga and coral rubber covered with algae. A total of 90 representative samples of living coral and 26 of dead coral covered with algae and 106 of coral rubber were selected randomly. Correlation analysis and Cluster analysis support that distinct differences in reflectance spectra among categories existed. Common spectral characteristic of living corals, dead corals and coral rubber covered with algae was a reflectance minimum at 674 nm. Healthy corals, dead coral covered with algae and coral rubber covered with algae showed high similarity of spectral reflectance. It is estimated that this is due to photosynthetic pigments.

Keyword : Hyperspectral data, spectral reflectance, coral rubber, living corals, dead corals

1. INTRODUCTION

Coral reefs are very important ecosystem in tropical waters. To conserve coral reefs, it is necessary to monitor them). Therefore, spectra of coral reefs is very important information for dynamic coral reefs. Several researchers have attempted to exploit hyperspectral data (from airborne and close range) to study coral groups (e.g., Mumby et al., 1998; Hochberg and Atkinson, 2000; Mishra et al., 2004, 2007; Clark et al., 1997, 2000; Myers et al., 1999; Schalles et al., 2000; Lubin et al., 2001; Joyce and Phinn, 2003; Kutser et al., 2003; and Mishra et al., 2006. Hochberg et al. (2003) provided a comprehensive literature review on *in situ* remote sensing research pertaining to coral reef ecosystem. They classified types of coral communities —healthy coral, dead coral, algae, and sand but only one of research have attempted in Indonesia by Holden and LeDrew in 1998, 2000 and 2001 to characterize coral groups using field radiometers at close range. Recent studies Hochberg et al. (2006) and; Kutser and Jupp (2006), Stambler and Shashar (2007) and Rundquist et al. (2009) have focused on coral species recognition, and the extent of variability in the reflectance spectra of corals of specific species. Fundamental research regarding the spectral differences between common coral reef features is necessary. Therefore, it is important to determine the spectral characteristics of living coral, dead coral and coral rubber covered with algae.

In this study, we were interested in examining the naturally occurring variations in reflection spectra within a coral rubber, healthy and dead corals covered with algae of a given locality. The objectives of our work were to: (1) document the spectral features of coral rubber, healthy coral and dead corals covered with algae as distributed within four study site in the Spermonde archipelago; and (2) develop spectral library to determine whether living and dead corals covered with algae and coral

rubber covered with algae are spectrally discriminable and how to achieve discrimination. Whereas, this research is expected as baseline information on optical characters of living and dead coral covered with algae and coral rubber covered with algae that can be used as a basic knowledge in interpreting satellite images so that they facilitate in identifying existence and differentiating some healthy coral and dead coral covered algae, especially using hyperspectral.

2. MATERIALS AND METHODS

2.1 Spectra collection

Field data collection was carried out in Samatellu Lompo, Samatellu Pedda, Samatellu Borong Island and Gusung in Spermonde archipelago, Indonesia (9479612,68 N, 757922,94 E and 9478861,44 N, 762635,26 E; Figure 1. Spectra were collected on January and February 2010 under generally clear skies. The data collection occurred between 9:00 a.m. and 15:00 p.m. Central Standard Time, using a LOT-2 Spectra Corps spectroradiometer. The samples comprised living and dead coral covered with alga and coral rubber covered with algae. Spot measurement of individual substrate types were made from about 7 cm above the substrate, resulting in a field of view of about 1.5 cm. Each measurement took about half a minute, capturing over one hundred spectra (depending on the integration time)

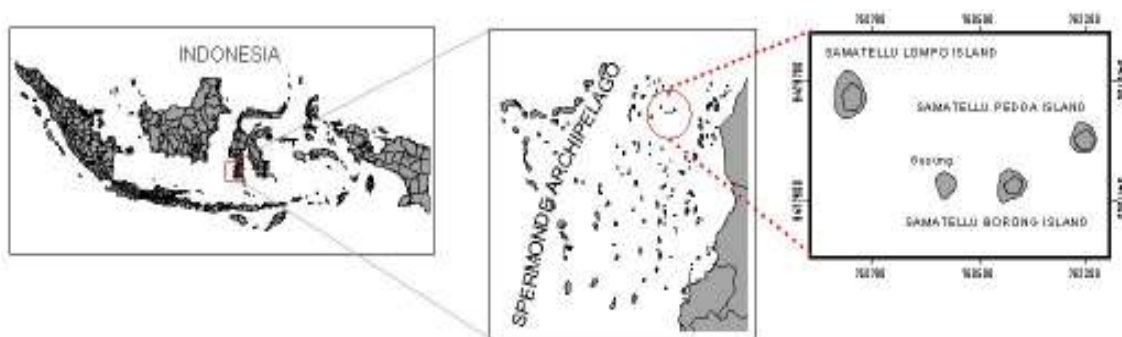


Fig1. Location of the Samatellu Lompo, Samatellu Borong, and Samatellu Pedda island study sites in Spermonde Archipelago, South Sulawesi, Indonesia.

We studied the optical properties of the *in situ* measured living and dead coral covered with algae and decided to divide the substrates into the following classes: live corals (*Acropora formosa*, *Seriatopora stelata*, *Acropora macrostomata*, *Acropora sarmentosa*, *Porites columnaris*, *Porites meyeri*), dead corals covered with algae (dead *Porites* and dead *Acropora*), coral rubber (<3 months and >3 months). In the present study the coral rubber were divided into recently (less than 3 month) and long time (considerably greater than 3 months) coral rubber. Dead coral was greater than 3 months. Recently coral rubber began covered with turf algae and coralline white is visible clearly. Long time coral rubber are mainly covered with turf algae that growing rapidly and significantly darker than recently coral rubber and therefore more easily separable.

A total of 90 representative samples of living coral and 26 of dead coral covered with algae and 106 of coral rubber were selected randomly. The reflectance spectra were taken over each sample between 1 and 3 m in depth, and each spectrum was the result of averaging of individual scans compiled over approximately 30 seconds total. The spectral range of the instrument is 300 - 1100 nm. Spectra are sampled with 3 nm intervals. The total number of substrates for which reflectance spectra were collected was 222.

2.2 Data Analysis

Correlation analysis and Cluster analysis were applied to data analysis. Cluster analysis was used to determine spectral similarity in and among coral species based on spectral responses at observed wavelengths. Similarity scale used was euclidean distance. Distance scale determined spectral

similarity and dissimilarity in which object with shorter distance would be more similar each other compared to object with longer distance.

3. RESULTS AND DISCUSSION

3.1 Spectral Analysis

Our efforts focused primarily on measurements of living corals, dead corals covered with algae, and coral rubber. Reflectance spectra (Avg± Std, range 0-1) from 10 categories are shown in Fig.2. The coral rubber were divided into recently (less than 3 month) and long time (considerably greater than 3 month) coral rubber. Recently coral rubber began covered with turf algae and coralline white is visible clearly. Long time coral rubber are mainly covered with turf algae that growing rapidly and significantly darker than recently coral rubber and therefore more easily separable. The live corals, dead coral and coral rubber had a similar reflectance. This indicated that living corals couldn't be easily separated from dead corals and coral rubber.

Dead *Porites* appeared similar in spectral magnitude and shape to dead *Acropora* (Fig.3B). They have a peak reflectance at 605nm and reflectance minima at 674nm. Coral rubber display measured spectra with reflectance minima at 674nm. All of the measured parts of the live corals showed distinctive spectral features (Fig. 3C), such as a peak reflectance at 579nm, and drop at 674nm, and a steep rise around 700nm. Furthermore, with few exception, *Steriotopora stellate* and recently coral rubber spectra displayed highest reflectance (9%). The average maximum reflectance values were between 4% and 9% for all categories. There were certain spectral features common to most living corals, dead coral covered with algae and rubber covered with algae. Curva of mean apparent reflectance spectra showed that there are more spectral variations in the shape of the curves of living coral measurements than dead coral covered with algae measurements (Fig.3A,B), but there do not appear to be any significant differences in spectral reflectance within dead coral covered with algae category (Fig.3D). Reflectance was generally lower in the shorter wavelength region (400-500nm) and in most cases there were not distinctive features between substrates in this region.

3.2 Correlation analysis

To examine the similarities between the categories, Pearson correlation coefficients were calculated as summarized in Table 1. The Pearson correlation coefficient considers the profile, or spectral, shape (Wilkinson et al., 1996), so the coefficients represent the similarities of the entire spectrum as a whole. The average spectra for each 10 categories that comprise living and dead coral covered with algae and coral rubber were included in the correlation analysis. When the entire average spectrum is considered, the correlations between categories are high suggesting a high degree of similarity overall.

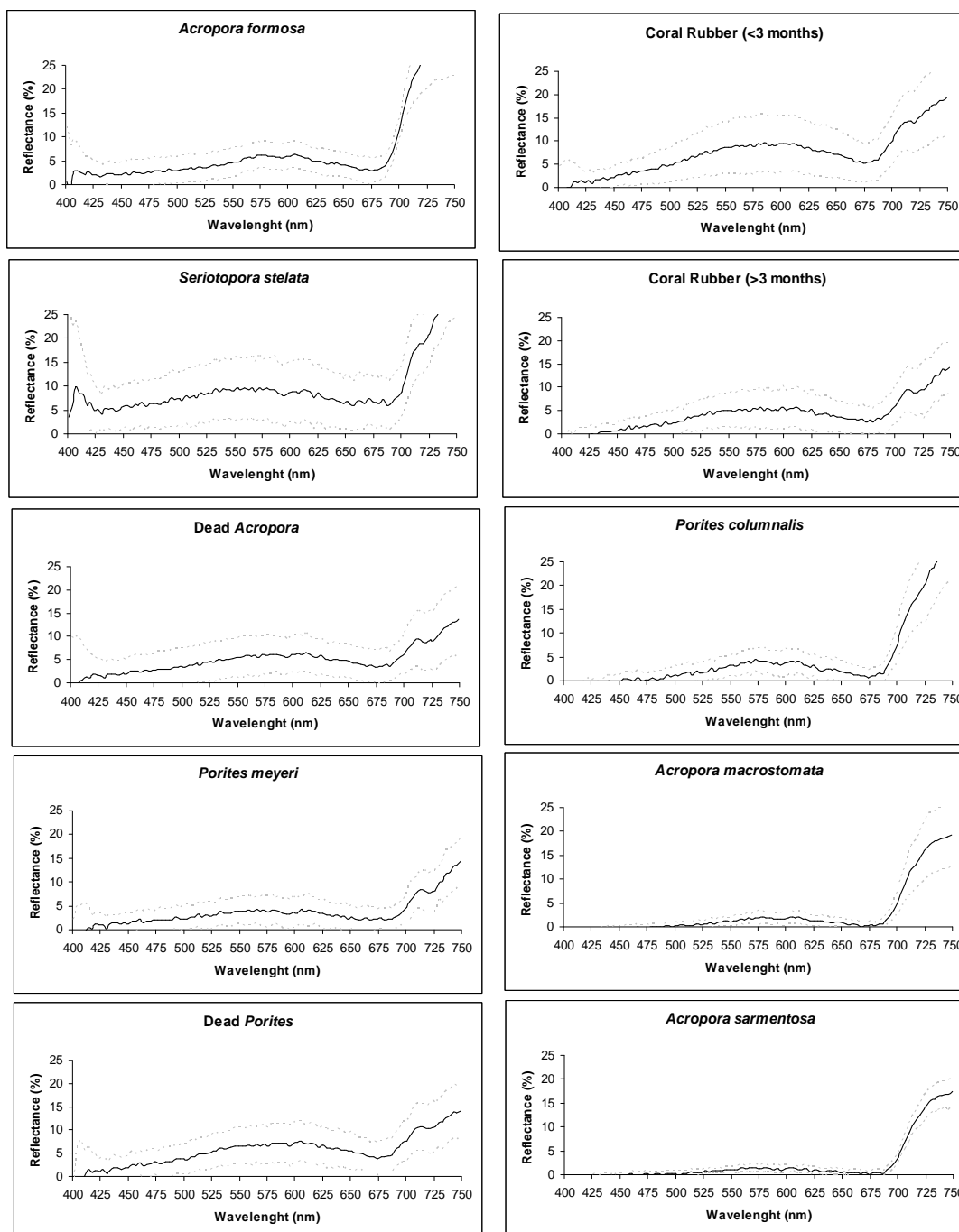


Fig.2 Reflectance spectra (Avg \pm Std, range 0-1) from 10categories

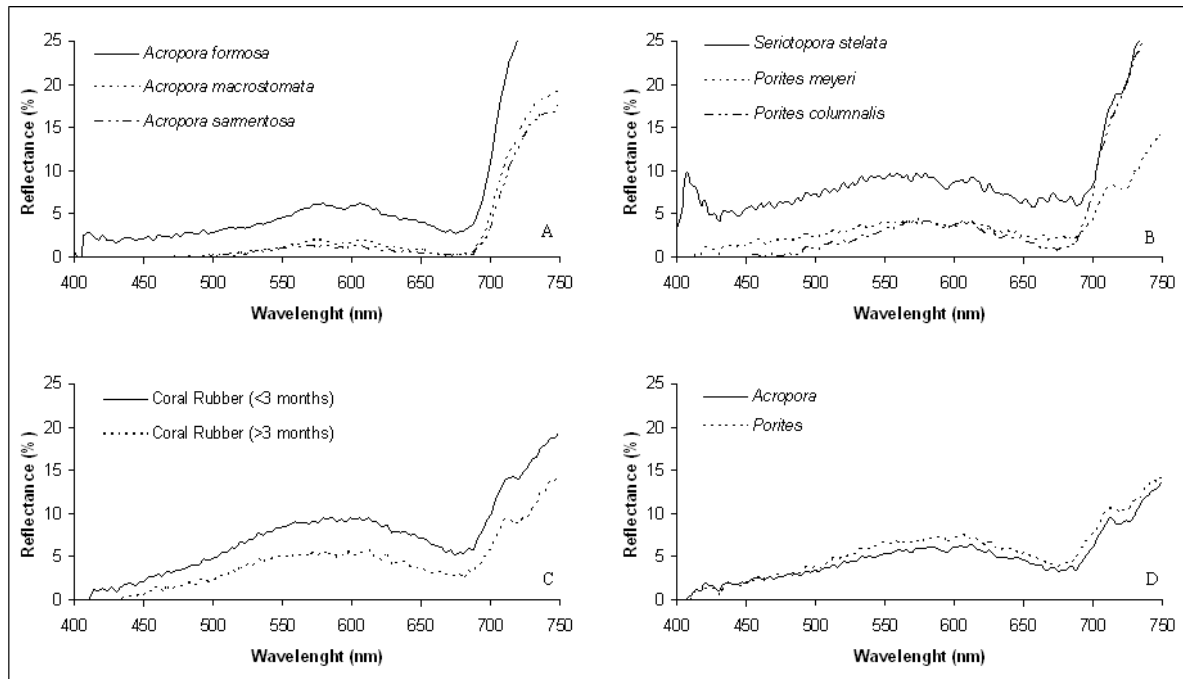


Fig. 3 Mean apparent reflectance spectra (%) of live coral acropora (A), live coral non acropora (B), rubber covered with algae (C), dead coral covered with algae (D). Dead coral covered with algae are represented by a reflectance spectrum of *Porites* and *Acropora*, live corals by *Acroporaformosa*, *Seriotopora stelata*, *Acroporamacrostromata*, *Acroporasarmentosa*, *Poritescolumnaris*, and *Poritesmeyerii* spectrum

Table 1. Correlation coefisien for 10 categories of live corals, dead corals covered with algae, coral rubber covered with algae

	<i>Acroporaformosa</i>	<i>Acroporamacrostromata</i>	<i>Acroporasarmentosa</i>	<i>Seriotopora stelata</i>	<i>Poritesmeyerii</i>	<i>Poritescolumnalis</i>	Coral Rubber (<3 months)	Coral Rubber (>3 months)	Dead <i>Acropora</i>	Dead <i>Porites</i>
<i>Acroporaformosa</i>	1									
<i>Acroporamacrostromata</i>	0.97	1								
<i>Acroporasarmentosa</i>	0.97	0.99	1							
<i>Seriotopora stelata</i>	0.96	0.93	0.94	1						
<i>Poritesmeyerii</i>	0.89	0.96	0.95	0.90	1					
<i>Poritescolumnalis</i>	0.96	0.99	0.99	0.95	0.98	1				
Coral Rubber (<3 months)	0.86	0.91	0.89	0.84	0.95	0.93	1			
Coral Rubber (>3 months)	0.77	0.88	0.86	0.76	0.96	0.90	0.96	1		
Dead <i>Acropora</i>	0.85	0.92	0.90	0.85	0.98	0.94	0.99	0.98	1	
Dead <i>Porites</i>	0.81	0.89	0.87	0.80	0.95	0.91	0.99	0.98	0.99	1

3.3 Spectral clustering

Cluster analysis is the generic name for a multivariate procedure of clumping similar objects into categories enabling identification of (1) outliers, and (2) the basic structure of the dataset. No satisfactory general method has been developed for deciding how many clusters exist in data set of unknown structure (Wilkinson et al., 1996). Therefore, the number of cluster is a subjective decision based on knowledge of the dataset characteristics. The objective of cluster analysis is to determine which objects are similar and dissimilar and categorize them accordingly (Holden and LeDrew., 1998).

Results of visualization at curve of reflectance curve were tested with cluster analysis applied to all of those coral species. In cluster analysis, it was found groups based on the spectral responses of each sample to eight wavelength groups and based on scale of similarity distance among these samples in which an object with shorter distance among samples would be more similar one to others compared to the objects having longer distance. Display of group division at overall wavelengths was presented in dendrogram graph as shown at Figure 4.

Based on Figure 3, there are many clusters as spectra, while on the right there is only one cluster. Therefore, moving from left to right denotes an encreasing degree of difference between spectra where a small Euclidean distance suggests that the spectra are most similar. Based on the reflectance values and formation of spectral curve pattern at the six live corals, two dead coral covered with algae, and 2 coral rubber, it can be observed that there is a similar spectral among categories.

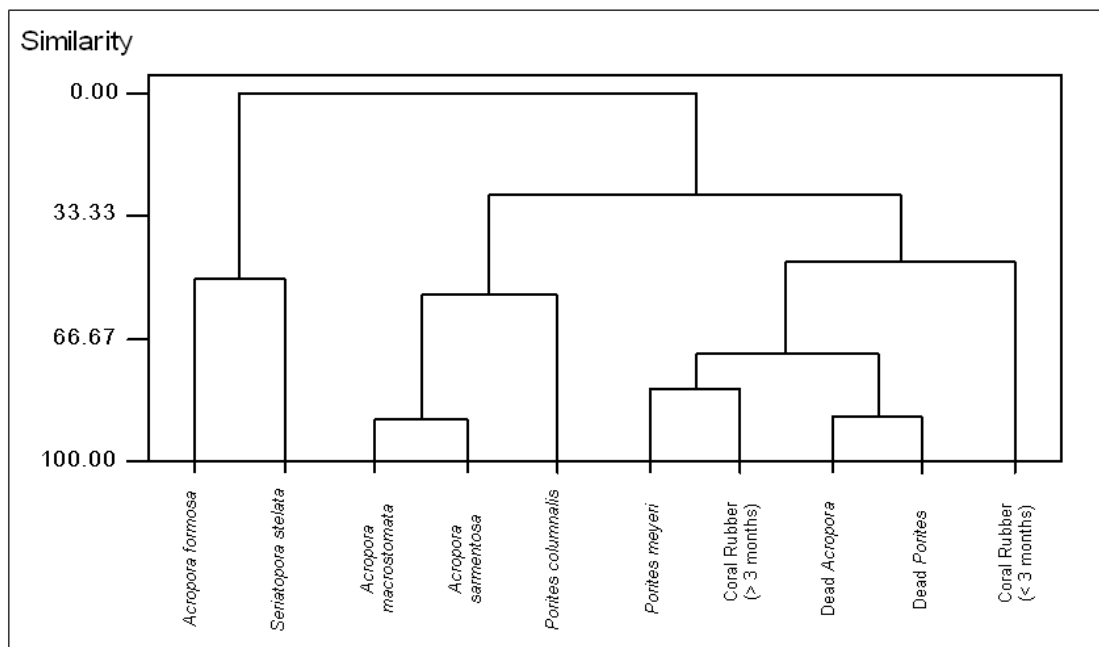


Fig.4. Dendrogram Graph of ten categories divided into six live corals (*Acroporaformosa*, *Acroporamacrostromata*, *Acroporasarmentosa*, *Seriotoporastelata*, *Poritesmeyer*, and *Poritescolumnaris*), 2 dead coral covered with algae (*Acropora*, and *Porites*) and 2 coral rubber (<3 months, and >3 months)

There were six main groups at similarity distance of 70,87% i.e. *Acroporamacrostromata* and *Acroporasarmentosa* having spectral similarity, categorized in one group. *Poritesmeyer* having spectral similarity with old coral rubber, dead *Acropora* and dead *Porites*, categorized in one group. However, *Acroporaformosa*, *Seriotoporastelata*, *PoritesColumnaris*, recently coral rubber have formed groups with himself. They have not shown a spectra similarity with another categories. At similarity distance of 87,81%, it was show eight groups i.e. *Acroporamacrostromata* and *Acroporasarmentosa* categorized in one group. Dead *Acropora* and dead *Porites* having spectral similarity categorized in one group. *Acroporaformosa*, *Seriotoporastelata*, *PoritesColumnaris*, *Poritesmeyer* recently coral rubber, and old coral rubber have not shown a spectra similarity with another categories. They have formed groups with himself.

231

232 3.4 Discussion

233

234 The Pearson correlation analysis revealed that when the entire spectral curve is considered, there is a
235 strong correlation between and within the living corals, dead coral covered with algae and coral
236 rubble covered with algae. Coral rubble is covered with epiphytic algae after bleaching by blast fishing
237 activities. As time goes on, epiphytic algae become thicker. Reflectance of coral rubble is changed
238 with period after bleaching. Epiphytic algae become visible size by three months after bleaching. By
239 virtue of change in algae cover over time, it was possible to discriminate between coral rubble
240 bleached less than 3 months, and more than 3 months.

241 Therefore coral rubble showed the similar shape but have a different of spectral magnitude and peak
242 position between two groups of coral rubble. These differences could be used as precise indicators
243 and predictors for identifying coral rubble conditions reflecting the chlorophyll concentration. It
244 contributed significantly to the increase in value of reflectance. All photosynthetic organisms (living
245 corals, dead corals covered with algae, and coral rubble covered with algae) displayed a reflectance
246 minimum at approximately 674nm, a feature related to the presence of chlorophyll.

247 In generally, living corals, dead corals covered with algae and coral rubber spectra display a
248 reflectance minimum at 674nm. Additionally, all of the live coral and coral rubber spectra have a peak
249 in reflectance at 579nm. However, dead coral have a peak reflectance at 605nm. The object will tend
250 to reflect the same colour as its colour and to absorb the other colours so that the intended object will
251 have higher reflectance values at similar wavelength colour to the original colour. Similarly perceived
252 colors may be the outcome of different spectra. Therefore, it is beneficial, if often difficult, to examine
253 spectra rather than colors when attempting to classify corals (Clark et al., 1997; Holasek et al., 1998;
254 Holden and LeDrew, 1999; Hochberg and Atkinson, 2000; Hochberg et al., 2003; Mazel et al. 2003;
255 Mazel and Fuchs, 2003).

256 Reflectance spectra of their study are similar in shape and magnitude to those of the our study. The
257 highest reflectance value of live corals was achieved at wavelengths of 579nm. Dead coral was
258 achieved a highest reflectance at 605nm. This is in agreement with a study by Kutser *et al* (2003)
259 that hard coral had high mean reflectance value at wavelength 550-700nm. Similar trend was found
260 by Nurdin and Rani (2008) which take measurement in the laboratory that reflectance peak for hard
261 corals was at wavelength 550-620nm. Low value at blue and green wavelengths are largely the result
262 of absorption by photosynthetic and photoprotective (Salih et al., 2000). Similarly, higher values at red
263 wavelength indicate lack of absorption or presence of active fluorescence (Mazel, 1995).

264 Chlorophyll in the zooxanthellae is an efficient absorber of light at the wavelength transmitted by
265 seawater, but its fluorescence emission at 685nm and longer wavelengths is strongly absorbed by
266 seawater (Mazel and Fuchs, 2003). Our results are in general agreement with other studies, which
267 measured light signals returned from corals. High spectral resolution measurements provide
268 opportunities for more refined assessments, primary because of pigment specific absorbance bands
269 and the great impact of pigments on intercepting light in the coral (Holden and LeDrew 1998).

270 The results of cluster analysis are encouraging with respect to the separability of live corals, dead
271 coral covered with algae and coral rubber reflectance. Similarity level among groups formed was high
272 or on the other word spectral reflectance variability among ten categories was low. *Porites meyeri* was
273 in the same group with dead coral covered with algae and old coral rubber had a similar reflectance.
274 This indicated that *Porites meyeri* couldn't be easily separated from dead corals and old coral
275 rubber. All photosynthetic organisms (living and dead corals covered with algae, coral rubber covered
276 with algae) displayed a reflectance minimum at approximately 595 nm and 674nm, a feature related
277 to the presence of chlorophyll. The living coral and coral rubber reflectance spectra showed the most
278 variation in shape and magnitude in comparison with the other groups. Within dead coral had the the
279 least variation in spectral shape. The results of study by Holden and LeDrew (1998) showing that the
280 healthy cluster include 2 bleached spectra and 2 macroalgae spectra. Thus, there is a certain degree
281 confusion between healthy coral, bleached coral and algae. Since the macro algae contain
282 photosynthetic pigments, the confusion with healthy coral containing zooxanthellae is understandable.
283 The overall results of this cluster analysis suggest good separability based on measured reflectance.

284 Similarity level among groups of live corals, dead coral covered with algae and coral rubber showed
285 high similarity level or low variability of signal reflectance. Reflectance of coral is readily
286 distinguishable from that of other reef bottom types (Hochberg et al. 2003). This indicates significant
287 spectral differences between corals and other bottom types that are independent of coral groupings

(e.g., taxa), which further implies that variability in reflectance of coral must not be random (Hochberg et al. 2004). In this respect, corals share a high degree of similarity in reflectance. At the same time, it is possible that live corals and dead coral covered with algae groups themselves are distinguishable from each other. Discrimination between corals and other types may rely on spectral features that are independent of those features that might discriminate between coral groups (Hochberg et al. 2004). This was caused by difference in composition structure of coral corallite and corallite size (big or small). We have documented that a differences in the magnitude and shapes of spectral curves from different categories and a strong absorption of them at 674nm region (Fig. 3). However, the interpretation of the peaks and shoulders of our spectra as areas of lower pigment activity and absorbance is probably fluorescence features associated with the coral polyp host tissue (Mazel, 1995).

Our results described 9-38 spectral pattern within one category. For example, *Porites meyeri* (non acropora) has 30 spectral pattern and *dead Porites* has 12 spectral pattern. At these categories demonstrated that within and between live coral, dead coral and old coral rubber can present such a range of colors, making spectral discrimination between them difficult. According to a research by Karpouzli and Malthus (2003), spectral reflectance among coral species showed high variability. This was caused by difference in pigment content of each substrates. Different pigments would reflect and absorb light at different wavelengths, so that affecting their reflectance values. Longer wavelength than 600nm will be absorbed by chlorophyll-a, whereas the shorter wavelengths will be absorbed by accessory pigments (Hochberg and Atkinson, 2004). Thus it is not surprising that reflectance of living and dead corals and coral rubber also shows variability at these study.

We developed spectral library to determine whether live coral, dead coral covered with algae and coral rubber covered with algae are spectrally discriminable and how to achieve discrimination. More investigation is required to statistically determine the degrees to which various categories groupings are spectrally discernable. Basically, every material has different structure or particle composition and this difference influences its electromagnetic response pattern. We have shown that basic live corals, dead corals and coral rubber have characteristic reflectance, that within them, and that they are spectrally separable from each other. It was difficult to find a completely dead coral without colonization by algae. Dead coral are not actually pure because trace amounts of macroalgae were visible on the surface. The stage of dead coral is temporary and the affected corals will either recover to their normal pigmentation or die and be colonized by macroalgae (Holden and LeDrew, 2001). However, dead corals and coral rubber are rapidly colonised by algae whose pigmentation may be similar to that of the coral's zooxanthellae, making the distinction between live corals, dead corals and coral rubber more complicated.

References

- Burke L, Selig E, Spalding M (2002) Reef at Risk in Southeast Asia. Washington, D.C. World Resources Institute (WRI)
- Clark CD, Mumby PJ, Chisholm JRM, Jaubert J, Andréfouët S (2000) Spectral discrimination of coral mortality states following a severe bleaching event. *Int J Remote Sens* 21:2321–2327
- Hedley JD, Mumby PJ (2002) Biological and remote sensing perspectives of pigmentation in coral reef organisms. In: Southward AJ, Young CM, Fuiman LA (eds) *Advances in Marine Biology*. Academic Press, San Diego, pp 279–317
- Holden H, LeDrew E (1998) Spectral discrimination of healthy and non-healthy corals based on cluster analysis, principal components analysis, and derivative spectroscopy. *Remote Sens Environ* 65:217–224
- Holden H, LeDrew E (1999) Hyperspectral identification of coral reef features. *Int J Remote Sens* 20:2545–2563
- Hochberg EJ, Atkinson MJ (2000) Spectral discrimination of coral reef benthic communities. *Coral Reefs* 19:164–171
- Hochberg EJ, Atkinson MJ (2003) Capabilities of remote sensors to classify coral, algae and sand as pure and mixed spectra. *Remote Sens Environ* 85:174–189
- Hochberg EJ, Atkinson MJ, Andréfouët S (2003) Spectral reflectance of coral reef bottom-types worldwide and implications for coral reef remote sensing. *Remote Sens Environ* 85:159–173

346 Hochberg E.J, Atkinson M.J, Apprill A, Andréfouët S (2004) Spectra reflectance of coral. *Coral Reefs*
347 23, 84–95.

348 Hochberg E.J, Apprill A.M, Atkinson M.J, Bidigare R.R (2006) Bio-optical modeling of photosynthetic
349 pigments in corals. *Coral Reefs* 25, 99–109.

350 Joyce KE, Phinn SR (2003) Hyperspectral analysis of chlorophyll content and photosynthetic capacity
351 of coral reef substrates. *LimnolOceanogr* 48:489–496

352 Karpouzli A.E, Malthus T.J, Place C (2004) Hyperspectral discrimination of coral reef benthic
353 communities in the western Caribbean, *Coral Reefs*, 2004, 23, 141-151

354 Kirk, J. T. O (1994) *Light and photosynthesis in aquatic ecosystems*, 2nd ed. Cambridge Univ. Press

355 Kutser T, Dekker AG, Skirving W (2003) Modeling spectral discrimination of Great Barrier Reef
356 benthic communities by remote sensing instruments. *LimnolOceanogr* 48:497–510

357 Kutser T, &Jupp D. L. B (2006) On the possibility of mapping living corals to the species level based
358 on their optical signatures. *Estuarine Coastal Shelf Sci.*, 69, 607-614.

359 Mazel CH (1995) Spectral measurements of fluorescence emission in Caribbean cnidarians. *Mar*
360 *EcolProgSer* 120: 185-191

361 Mazel CH (1996) Coral fluorescence characteristics: excitationemission spectra, fluorescence
362 efficiencies, and contribution to apparent reflectance. *SPIE Volume 2963 - Ocean Opt XIII*
363 1:240–245

364 Mazel CH, Fuchs E (2003) Contribution of fluorescence to the spectral signature and perceived color
365 of corals. *LimnolOceanogr* 48:390–401

366 Mishra D, Narumalani S, Bahl R, Rundquist D, Lawson M (2006) Predicting the Percent Live Cover of
367 Corals: An *in situ* Remote Sensing Approach. *Geophysical Research Letters*, 33, L06603

368 Mishra D, Narumalani S, Lawson M, and Rundquist D (2004) Bathymetric Mapping Using IKONOS
369 Multispectral Data. *GIScience and Remote Sensing*, 41(4):301–321.

370 Mishra D, Narumalani S, Rundquist D, Lawson M (2005) High Resolution Ocean Color Remote
371 Sensing of Benthic Habitats: A Case Study at the Roatan Island, Honduras. *IEEE Transactions*
372 *in Geosciences and Remote Sensing*, 43(7):1592–1604.

373 Mishra D, Narumalani S, Rundquist D, Lawson M (2006) Benthic Habitat Mapping in Tropical Marine
374 Environments Using QuickBirdImagery. *PhotogrammetricEngineering and Remote Sensing*,
375 72(9):1037–1048.

376 Mishra D, Narumalani S, Rundquist D, Lawson M (2007) Enhancing the Detection and Classification
377 of Coral Reef and Associated Benthic Habitats: A Hyperspectral Remote Sensing Approach.
378 *Journal of Geophysical Research*, 112, C08014

379 Mumby P, Green E, Clark C, Edwards A (1998) Digital Analysis of Multispectral Airborne Imagery of
380 Coral Reefs. *Coral Reefs*, 17(1):59–69.

381 Mumby, P.J., Green, E.P., Edwards, A.J. and Clark, C.D. (1999). The cost-effectiveness of remote
382 sensing for tropical coastal resources assessment and management. *Journal of Environmental*
383 *Management* 55: 157-166.

384 Myers MR, Hardy JT, Mazel CH, Dustan P (1999) Optical spectra and pigmentation of Caribbean
385 Reef corals and macroalgae. *Coral Reefs* 18: 179-186

386 Rundquist D, Gitelson A, Lawson M, Keydan G, Leavitt B, Perk R, Jennifer Keck, Mishra DR,
387 Narumalani S (2009). Proximal Sensing of Coral Features: Spectral Characterization of
388 *Siderastrea* and *Acropora*. *GIScience & Remote Sensing*, 46, No. 2, p. 139–160.

389 Schalles JF, Rundquist DC, Gitelson AA and Keck J (2000) Close range, hyperspectral reflectance
390 measurements of healthy Indo-Pacific and Caribbean Corals. In: Proc of the Sixth Intern Conf,
391 Remote Sensing for Marine and Coastal Environments, Veridian ERIM Intern, Ann Arbor, MI,
392 USA, I: 431- 440.

393 Stambler N, Sharhar N (2007) Variation in spectral reflectance of the
394 hermatypic corals, *Stylophora pistillata* and *Pocillopora damicornis*. *Journal of Experimental*
395 *Marine Biology and Ecology* 351: 143–149

396 Wilkinson L, Blank G, Grunber, C (1996) *Desktop analysis with Systat*. New Jersey: Prentice Hall.